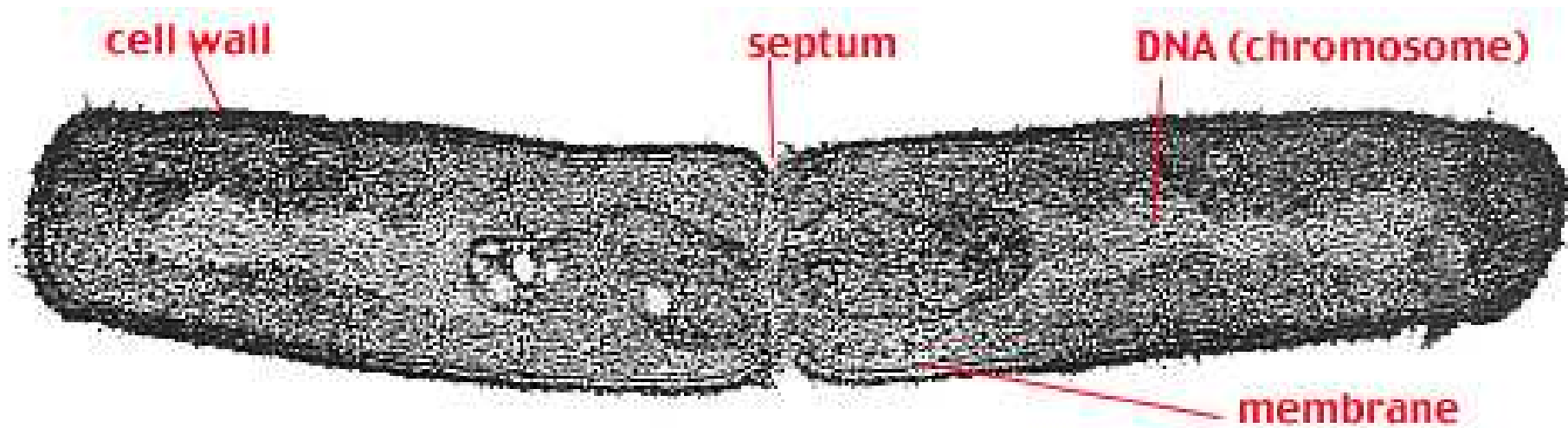


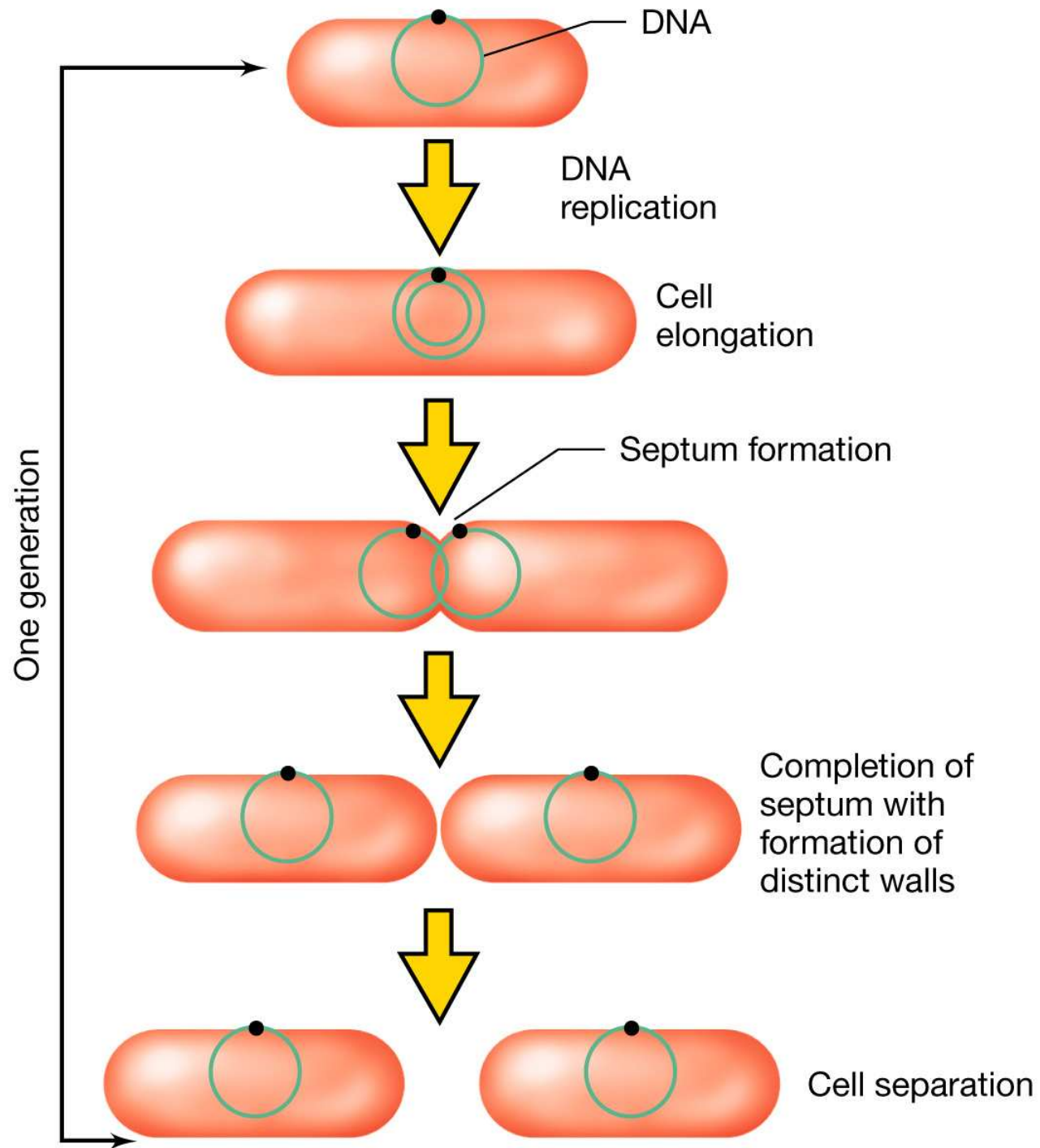
MICROBIOLOGIA GENERALE

Microbial growth 1

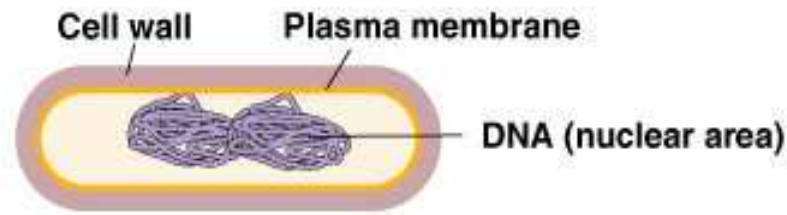
Microbial growth:
cell growth and binary fission

Bacterial growth by binary fission

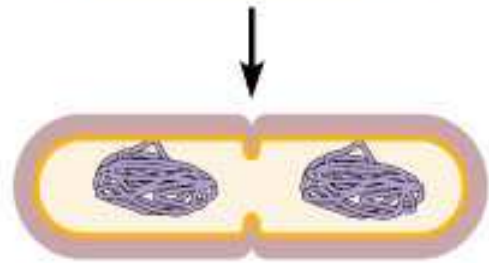




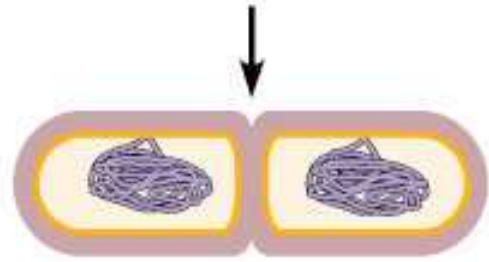
1 Cell elongates and DNA is replicated



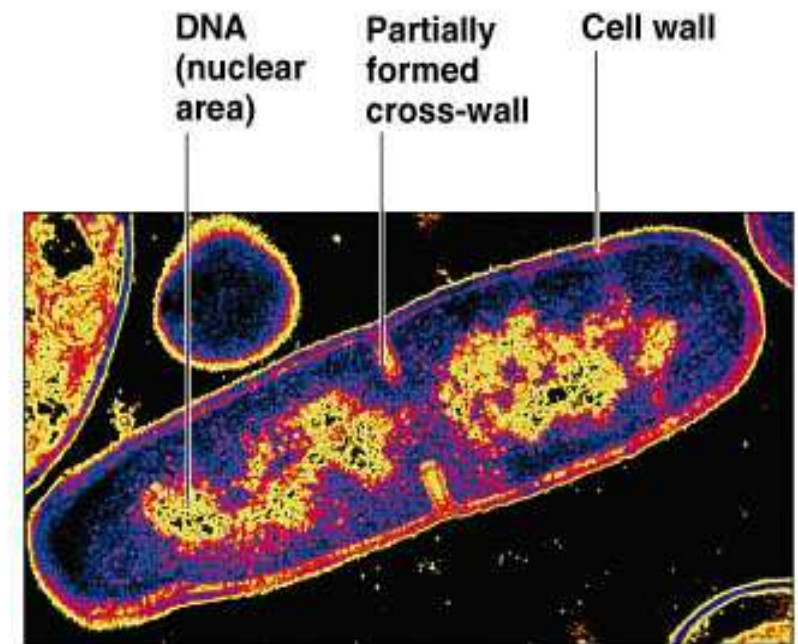
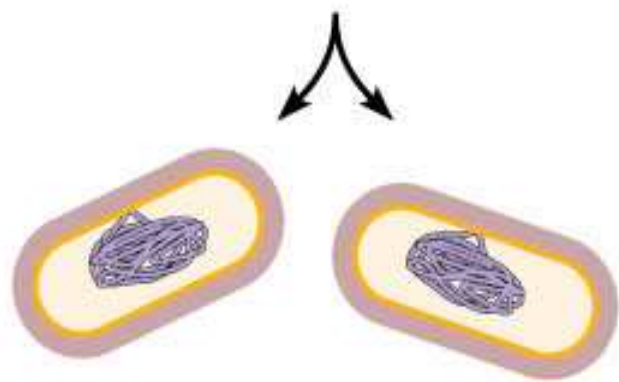
2 Cell wall and plasma membrane begin to divide



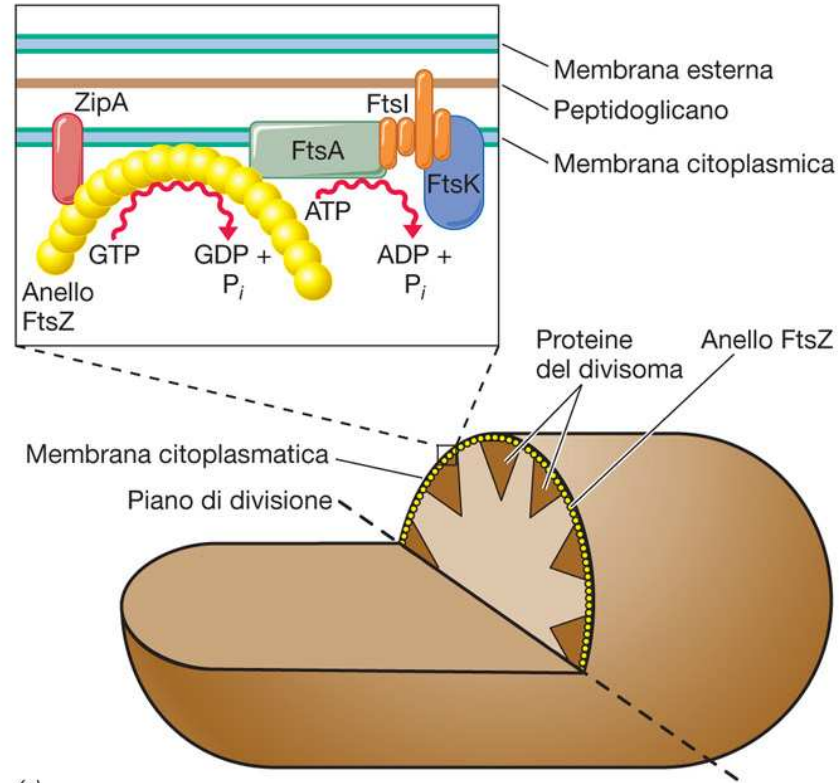
3 Cross-wall forms completely around divided DNA



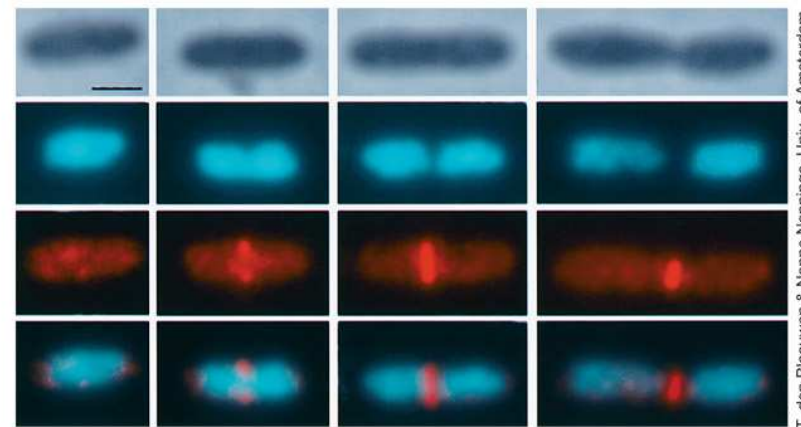
4 Cells separate



The FtsZ protein and cell division



(a)



(b)

Phase contrast

Nucleoid stain

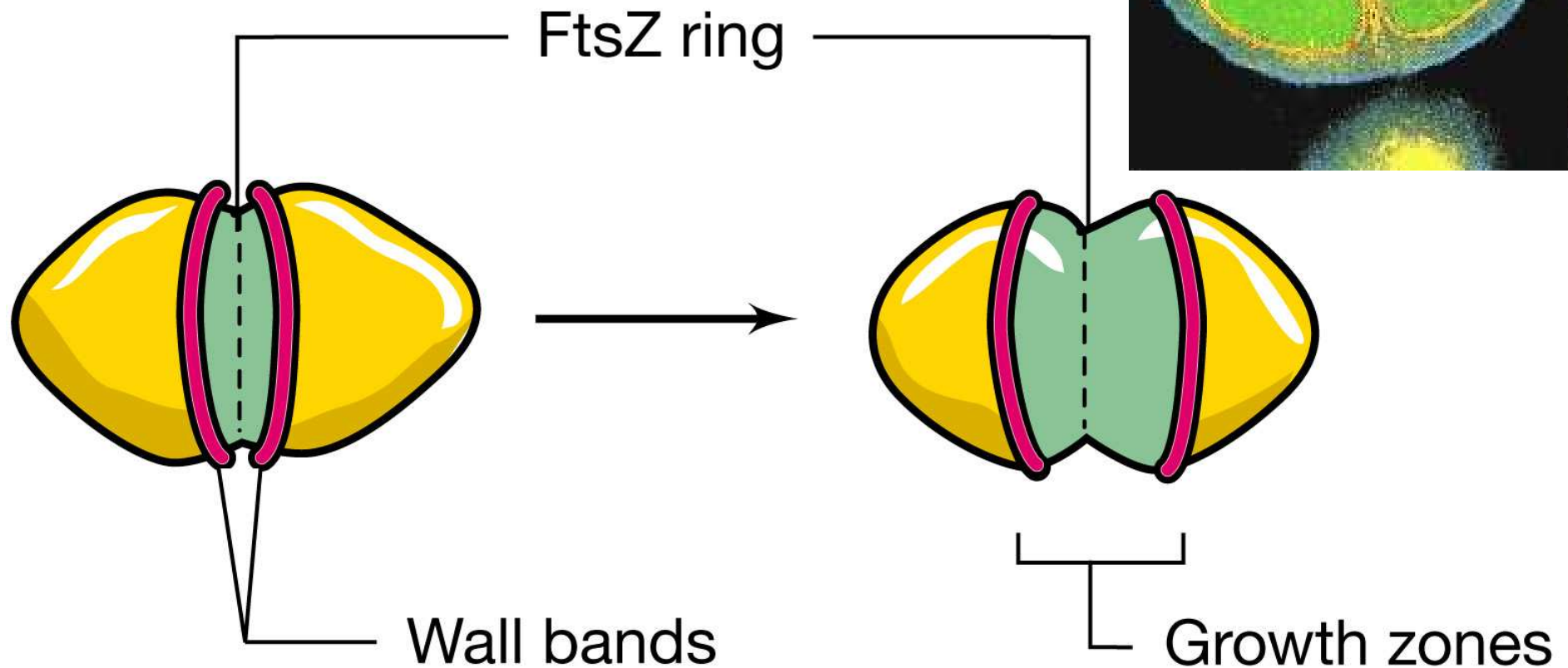
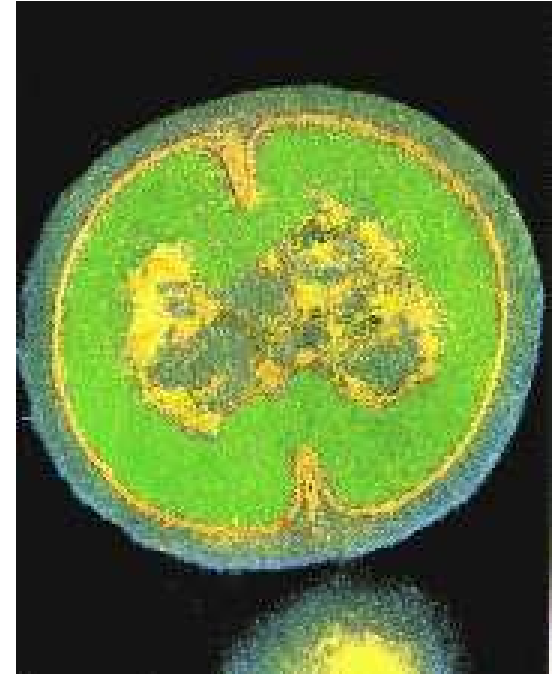
FtsZ stain

Merged

T. den Blaauwen & Nanne Nanninga, Univ. of Amsterdam

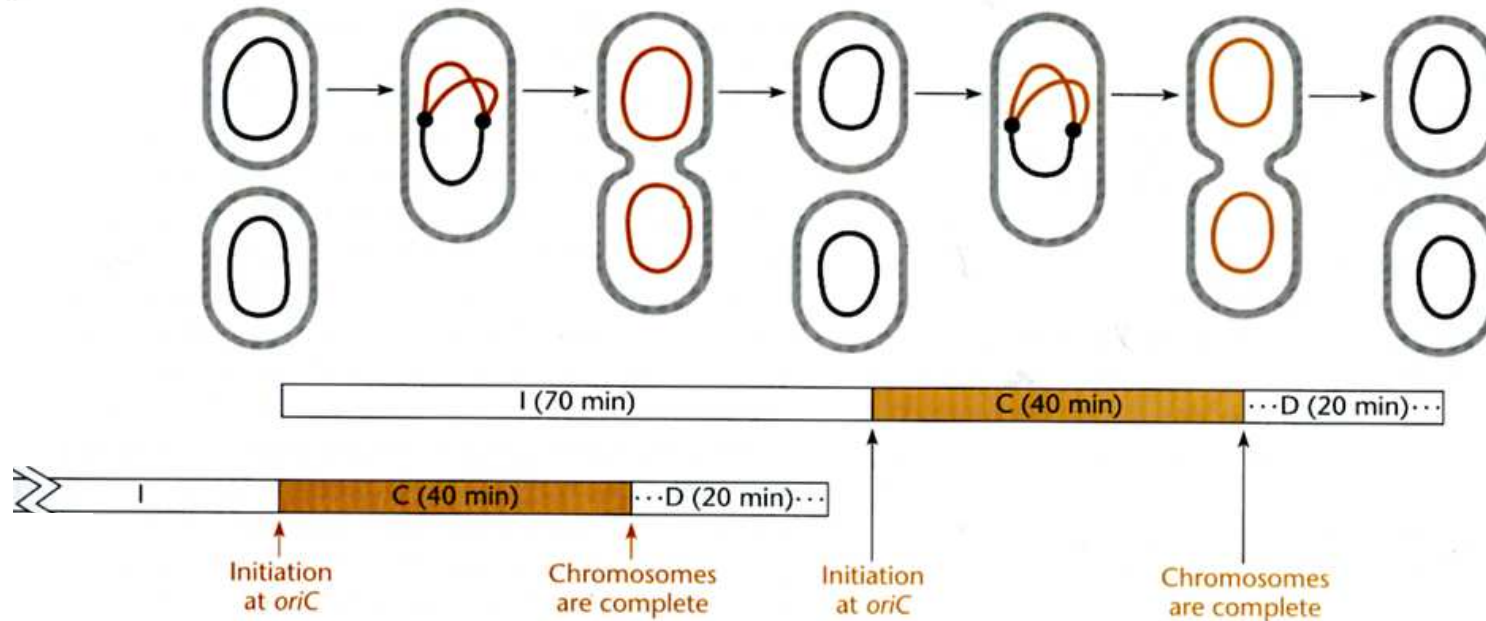
Cell wall synthesis during cell division in gram-positive Bacteria.

- In cocci, new cell wall synthesis (shown in green) is localized at only one point



Generation times for some bacteria under optimal conditions

Bacterium	Medium	Generation time (min)
<i>Escherichia coli</i>	Glucose-salts	17
<i>Bacillus megaterium</i>	Sucrose-salts	25
<i>Streptococcus lactis</i>	Milk	26
<i>Streptococcus lactis</i>	Lactose broth	48
<i>Staphyloc. aureus</i>	Heart infusion broth	27-30
<i>Lactobacillus acidophilus</i>	Milk	66-87
<i>Rhizobium japonicum</i>	Mannitol-salts-yeast extract	344-461
<i>Mycobacterium tuberculosis</i>	Synthetic	792-932
<i>Treponemapallidum</i>	Rabbit testes	1980



Slow growth $I = C + D = 70 \text{ min}$

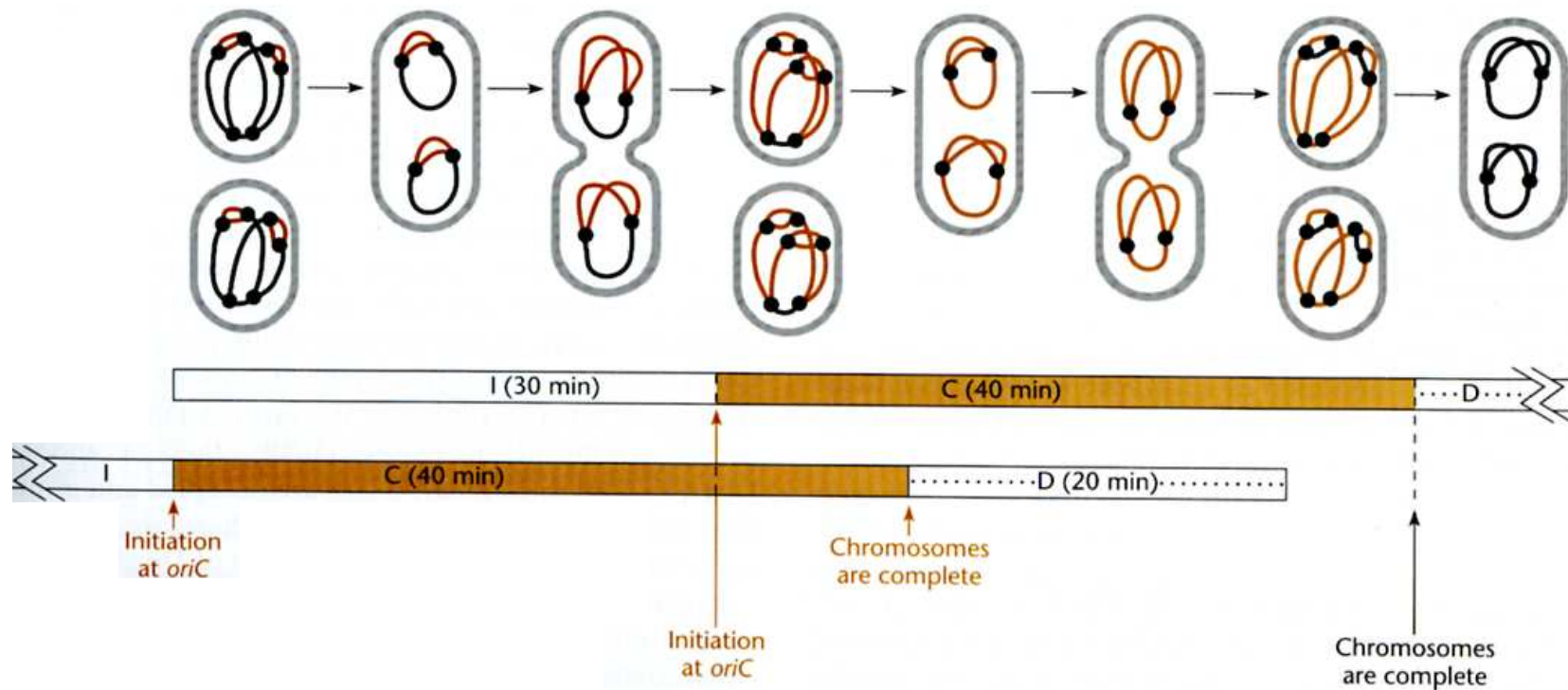
$C = 40 \text{ min}$ (DNA replication)

$D = 20 \text{ min}$ (septum formation
and cell division)

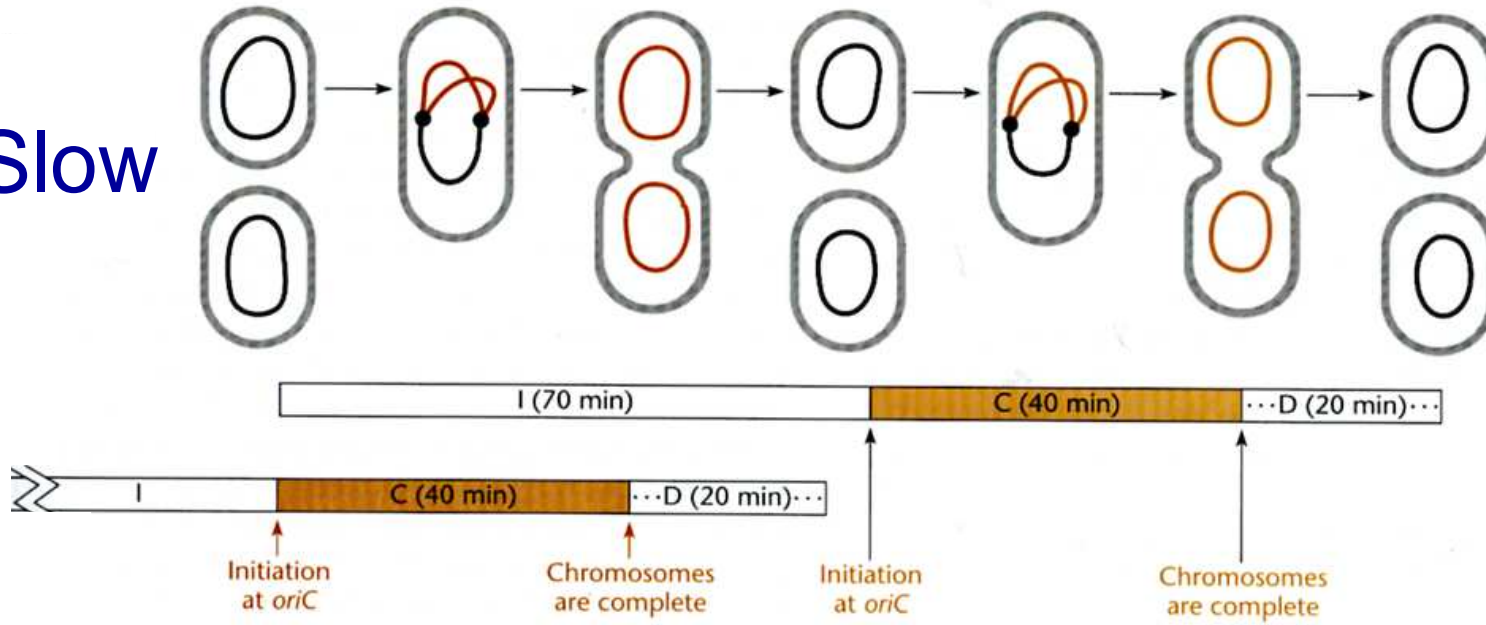
Rapid growth $I = C + D = 30$ min

C = 40 min (DNA replication)

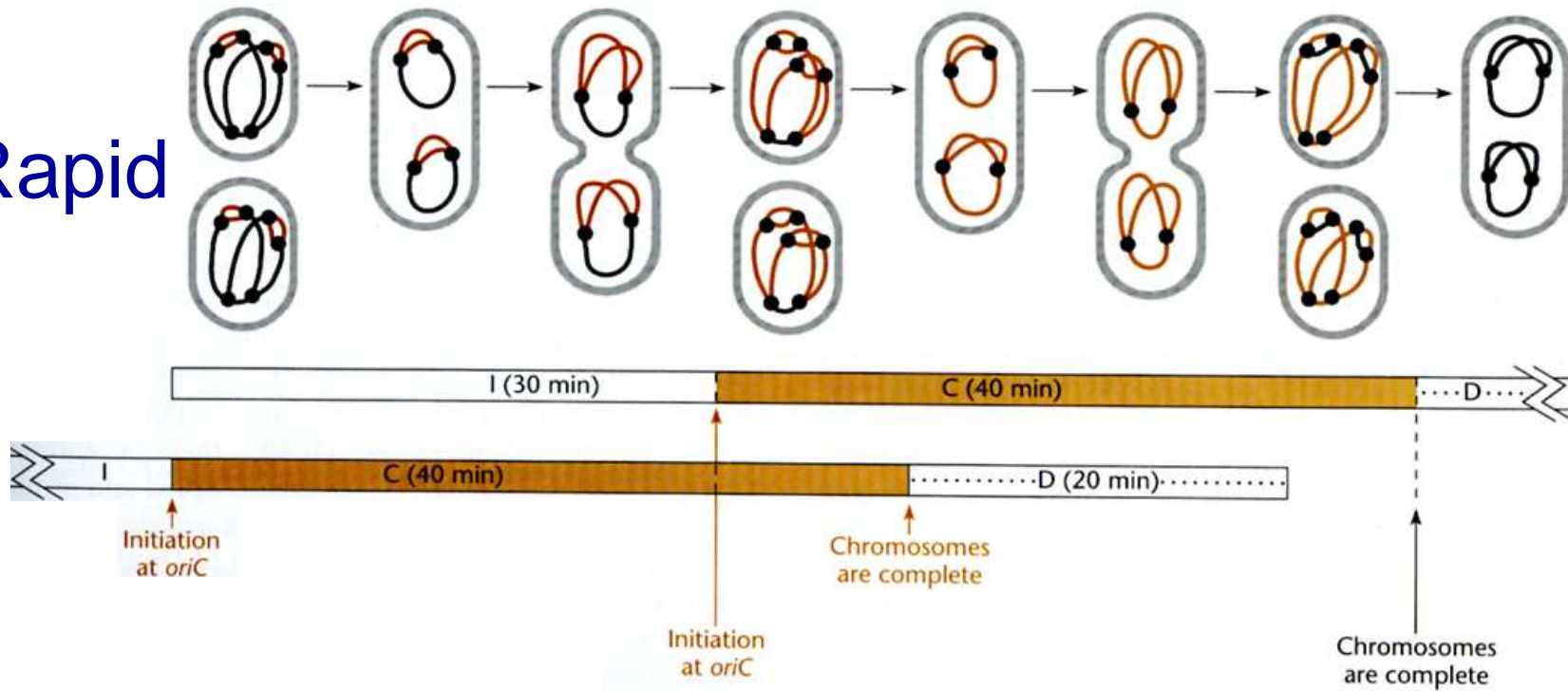
D = 20 min (septum formation
and cell division)



Slow



Rapid

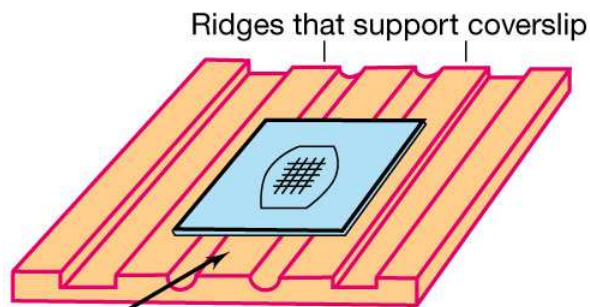


Microbial growth:
how to measure the growth

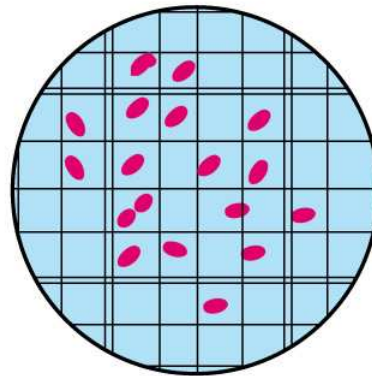
Some methods used to measure bacterial growth

Method	Application	Comments
Direct microscopic count	Enumeration of bacteria in milk or cellular vaccines	Cannot distinguish living from nonliving cells
Viable cell count (colony counts)	Enumeration of bacteria in milk, foods, soil, water, laboratory cultures, etc.	Very sensitive if plating conditions are optimal
Turbidity measurement	Estimations of large numbers of bacteria in clear liquid media and broths	Fast and nondestructive, but cannot detect cell densities less than 10^7 cells per ml
Measurement of total N or protein	Measurement of total cell yield from very dense cultures	Only practical application is in the research labs
Measurement of Biochemical activity (O ₂ uptake, CO ₂ , ATP)	Microbiological assays	Requires a fixed standard to relate chemical activity to cell mass and/or cell numbers
Measurement of dry weight or wet weight of cells or volume of cells after centrifugation	Measurement of total cell yield in cultures	probably more sensitive than total N or total protein measurements

A measurement of the microbial concentration: the **direct microscopic counting**



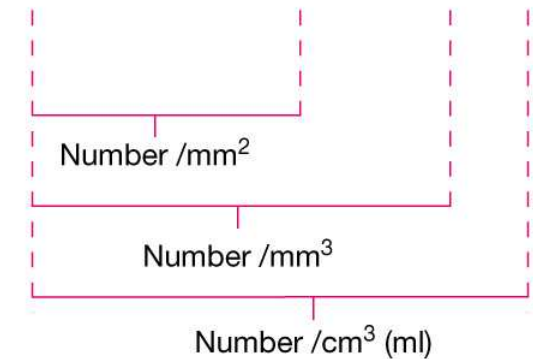
Sample added here; care must be taken not to allow overflow; space between coverslip and slide is 0.02 mm ($\frac{1}{50}$ mm). Whole grid has 25 large squares, a total area of 1 mm² and a total volume of 0.02 mm³.



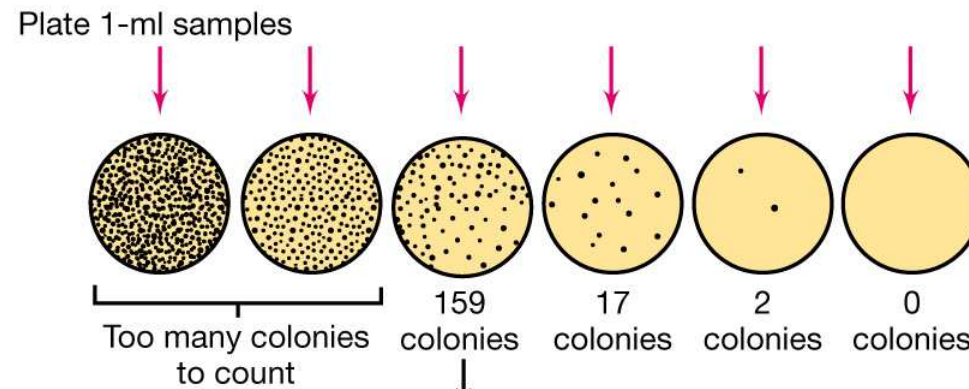
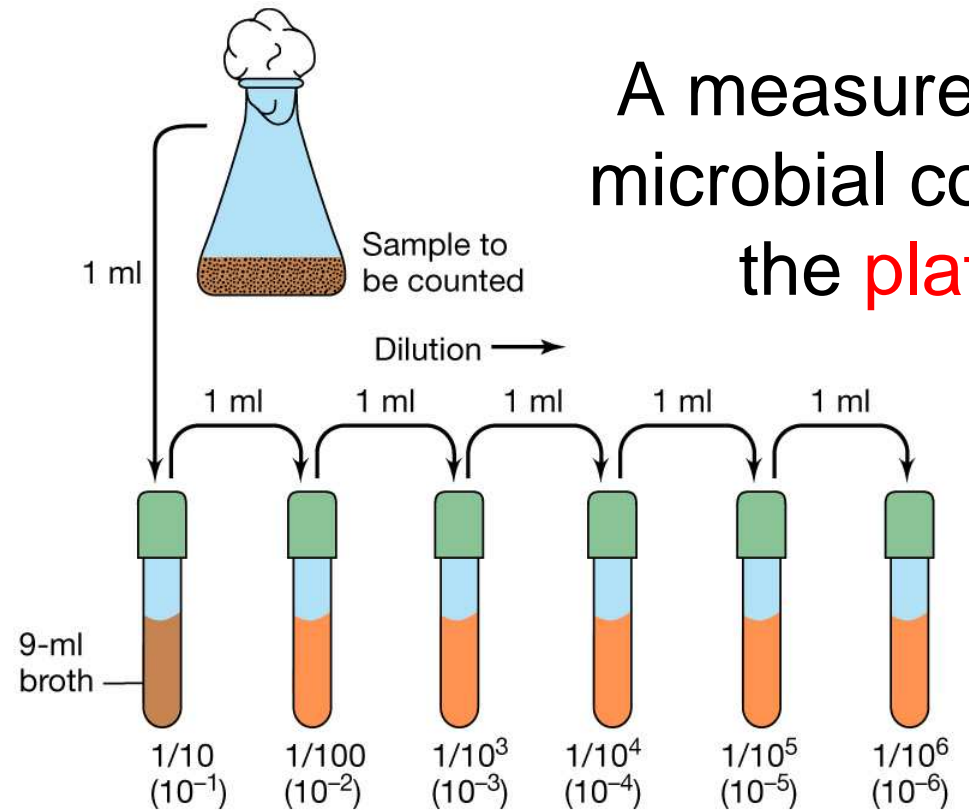
Microscopic observation; all cells are counted in large square: 12 cells (in practice, several squares are counted and the numbers averaged.)



To calculate number per milliliter of sample:
12 cells x 25 large squares x 50 x 10³
= 1.5 x 10⁷



A measurement of the microbial concentration: the **plate count**

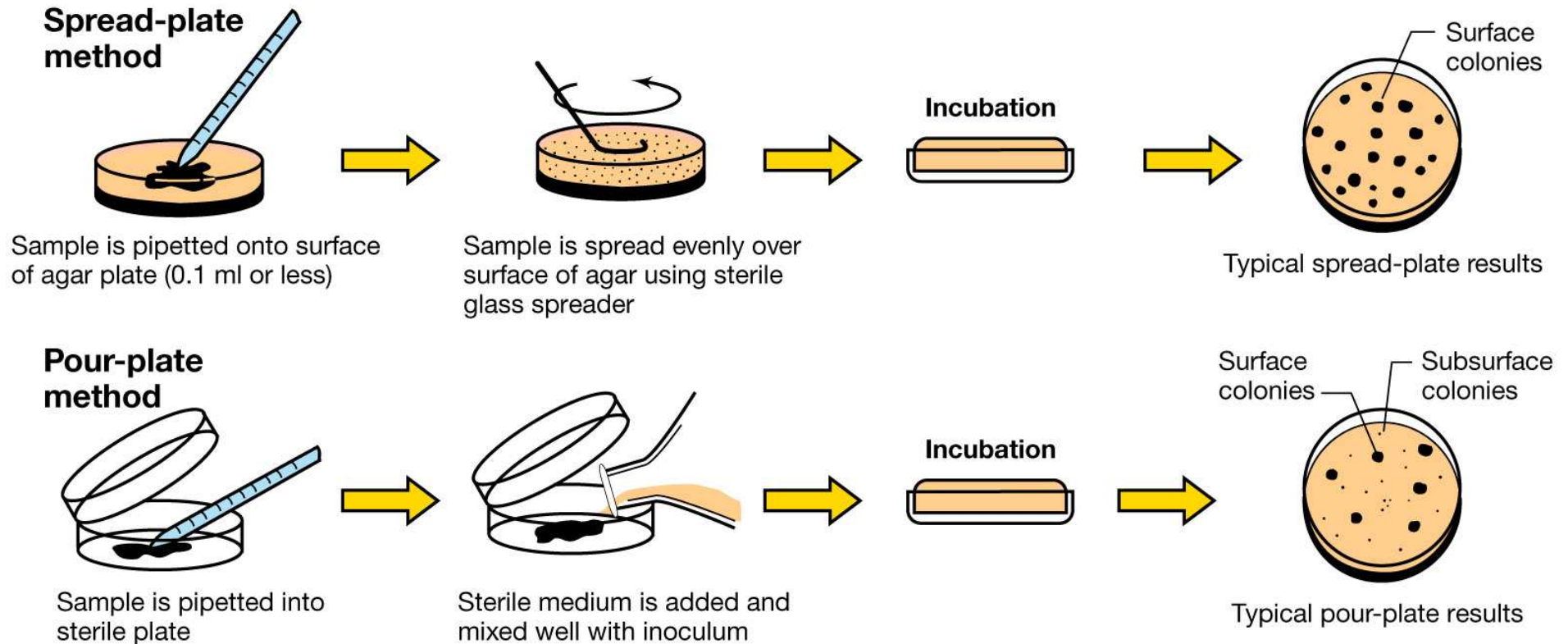


$$159 \times 10^3 \text{ Plate count} \times \text{Dilution factor} = 1.59 \times 10^5 \text{ Cells (colony-forming units) per milliliter of original sample}$$

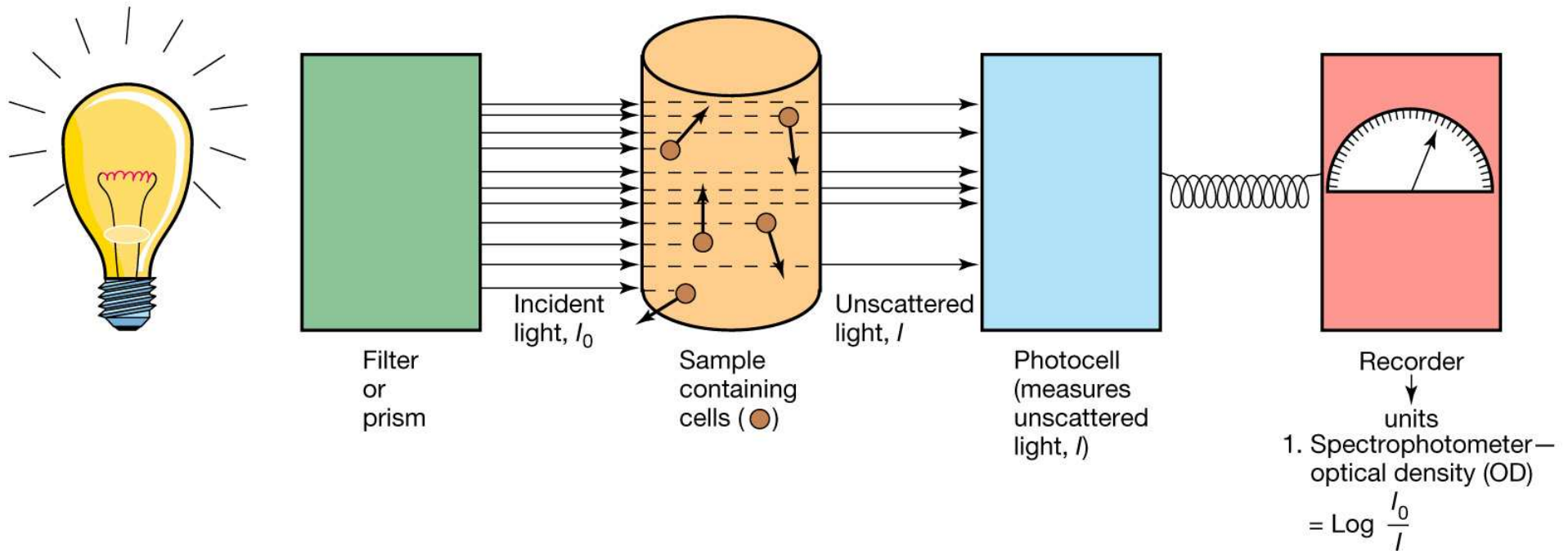
CFU/ml

Two methods of performing a viable count (plate count).

In either case the sample must usually be diluted before plating



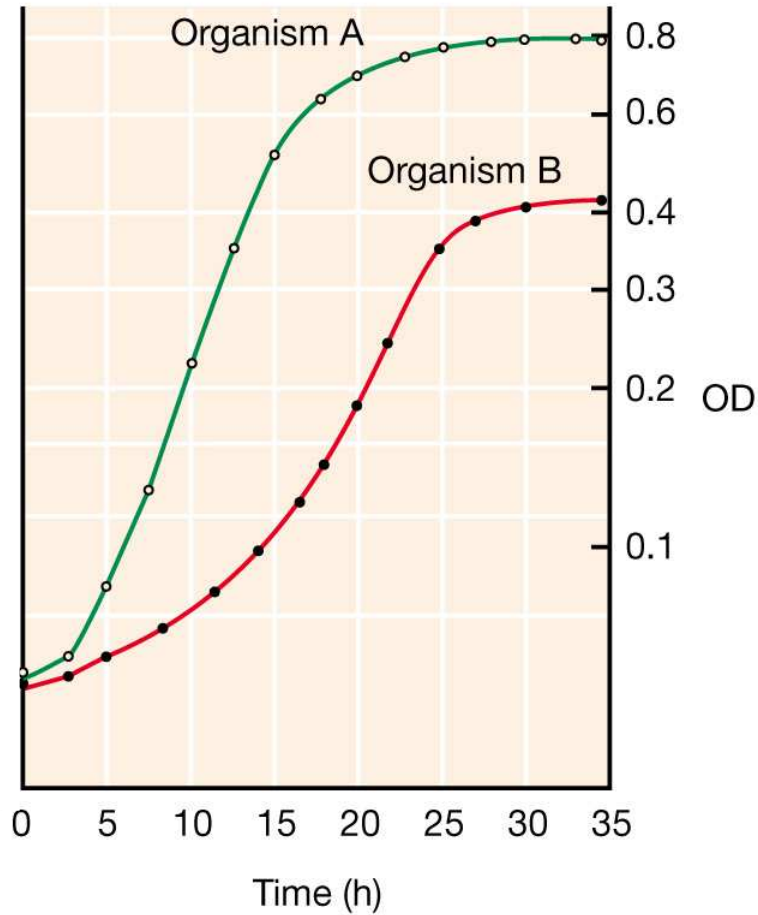
A measurement of the microbial mass: the **turbidity** procedure



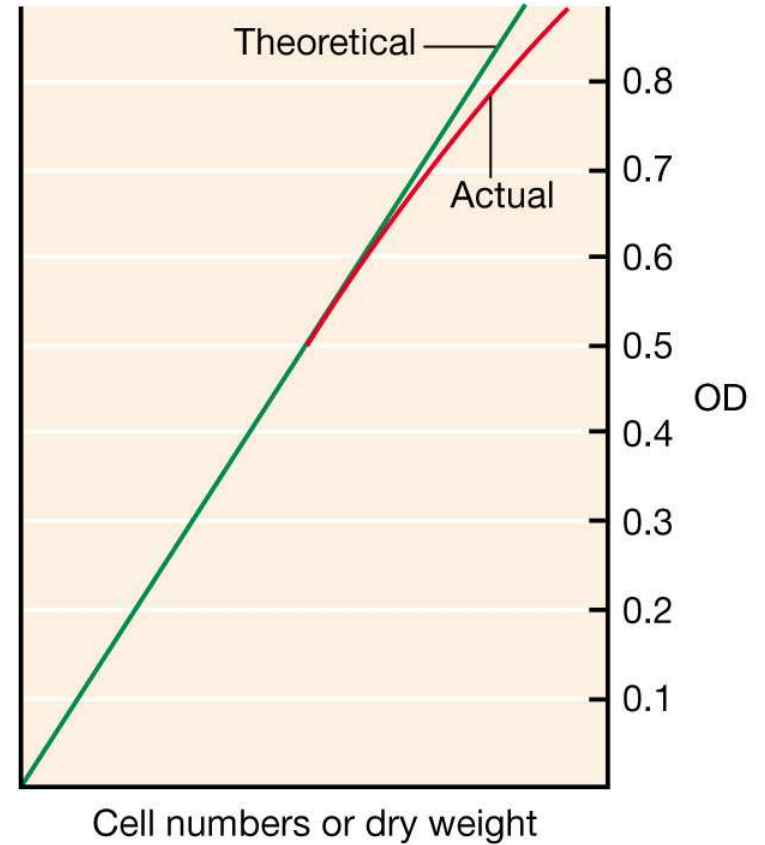
(a) $I_t = I_0 - (I_r + I_a)$ $T = I_t / I_0 \times 100$

O.D. (Optical Density) = $-\log T = \log I_0 / I_t$

Turbidity measurements of microbial growth.



Typical growth curves of two bacteria growing at different growth rates



Relationship between cell number or dry weight and turbidity readings

Microbial growth:
**the growth curve of a bacterial
population**

Time (h)	Total number of cells	Time (h)	Total number of cells
0	1	4	256
0.5	2	4.5	512
1	4	5	1,024
1.5	8	5.5	2,048
2	16	6	4,096
2.5	32	.	.
3	64	.	.
3.5	128	10	1,048,576

$I=30$ min

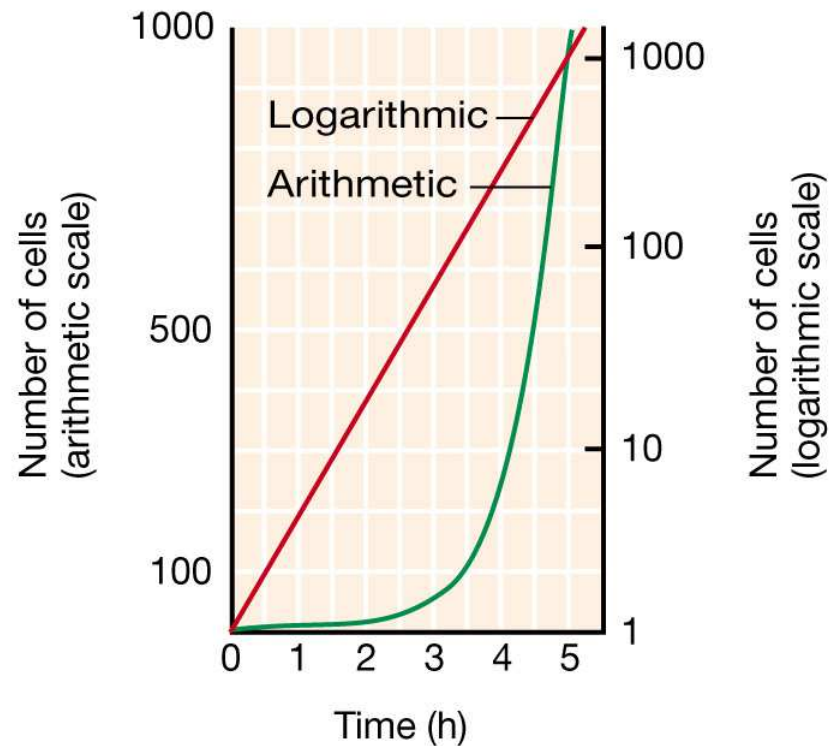
(a)

The rate of growth of a bacterial culture

Time (h)	Total number of cells	Time (h)	Total number of cells
0	1	4	256
0.5	2	4.5	512
1	4	5	1,024
1.5	8	5.5	2,048
2	16	6	4,096
2.5	32	.	.
3	64	.	.
3.5	128	10	1,048,576

$I=30$ min

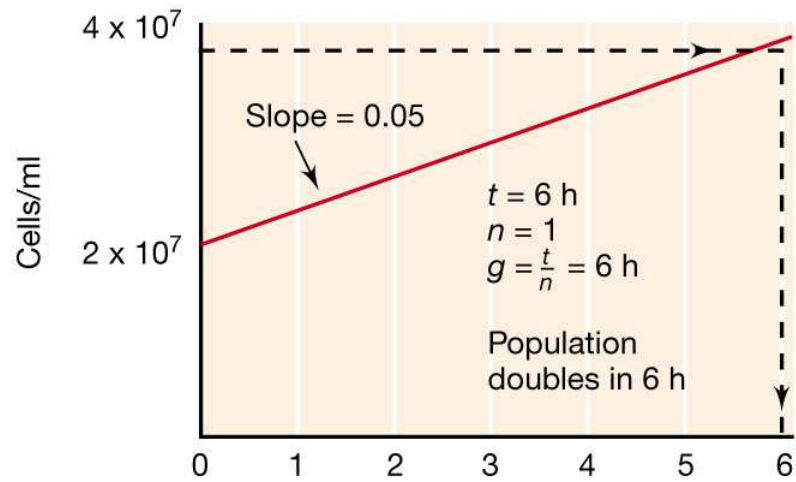
(a)



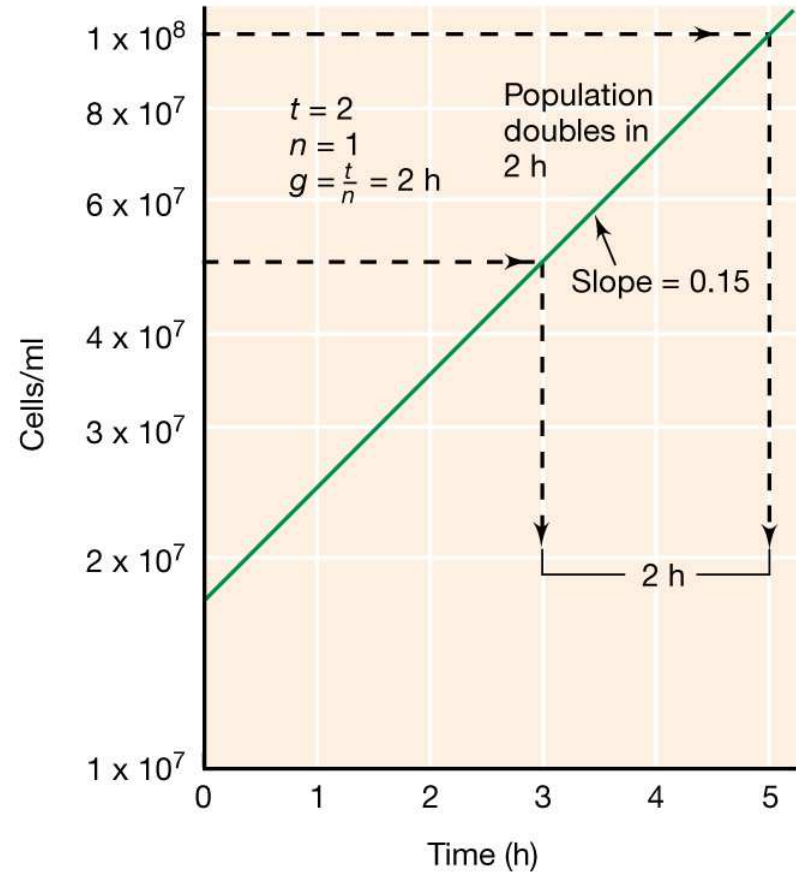
The rate of growth of a bacterial culture

(b)

Method of estimating the generation times (g)

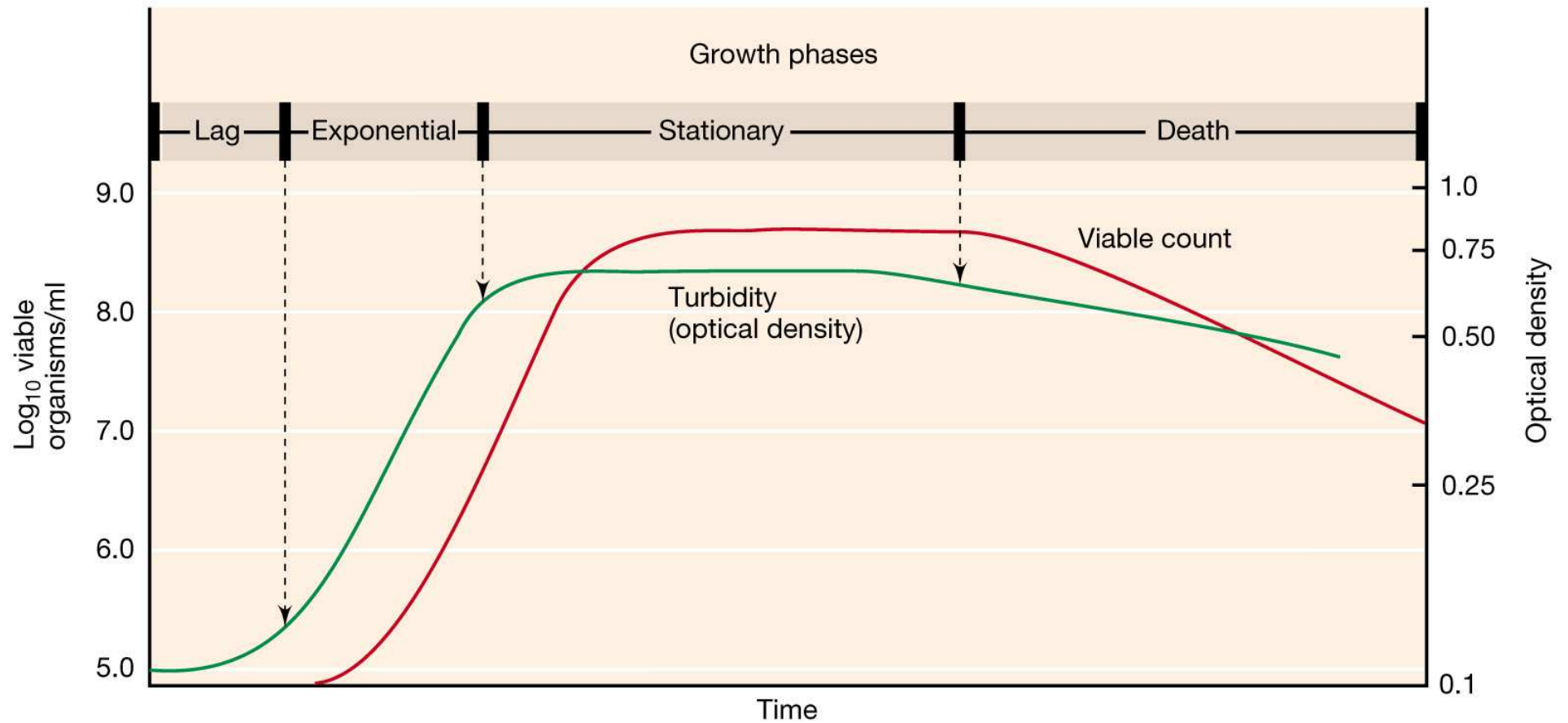


(a)

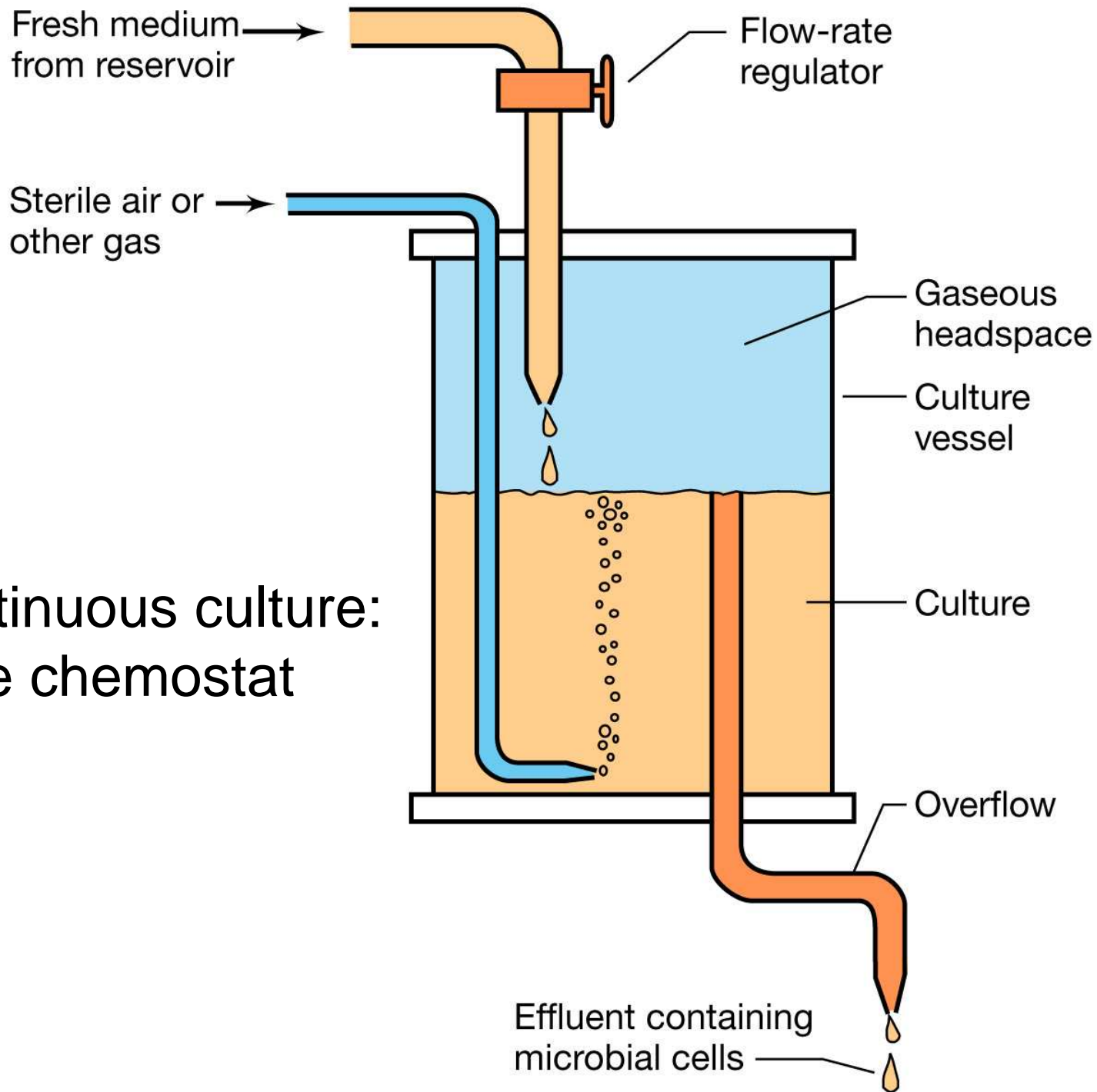


(b)

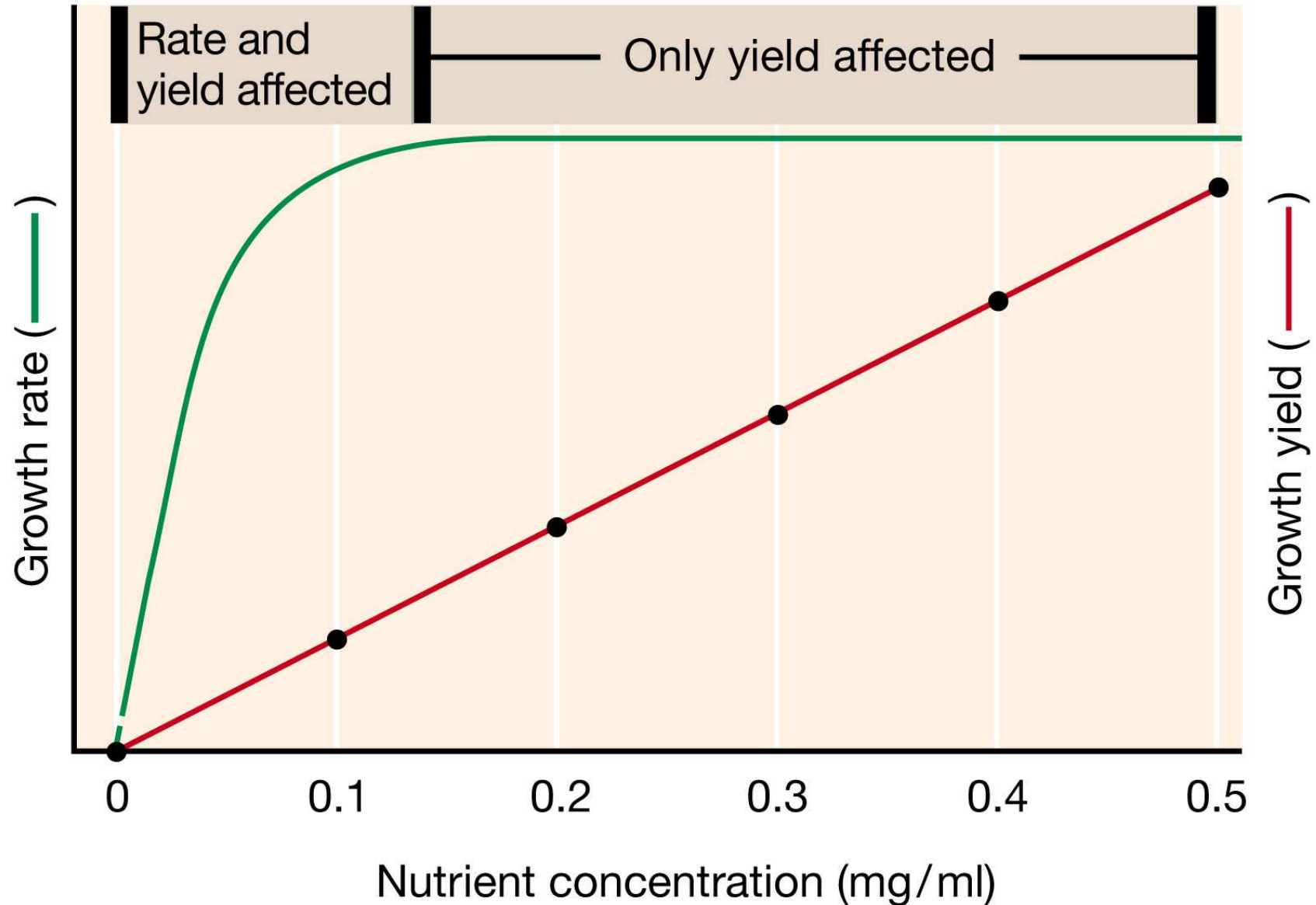
Typical **growth curve** for a bacterial population



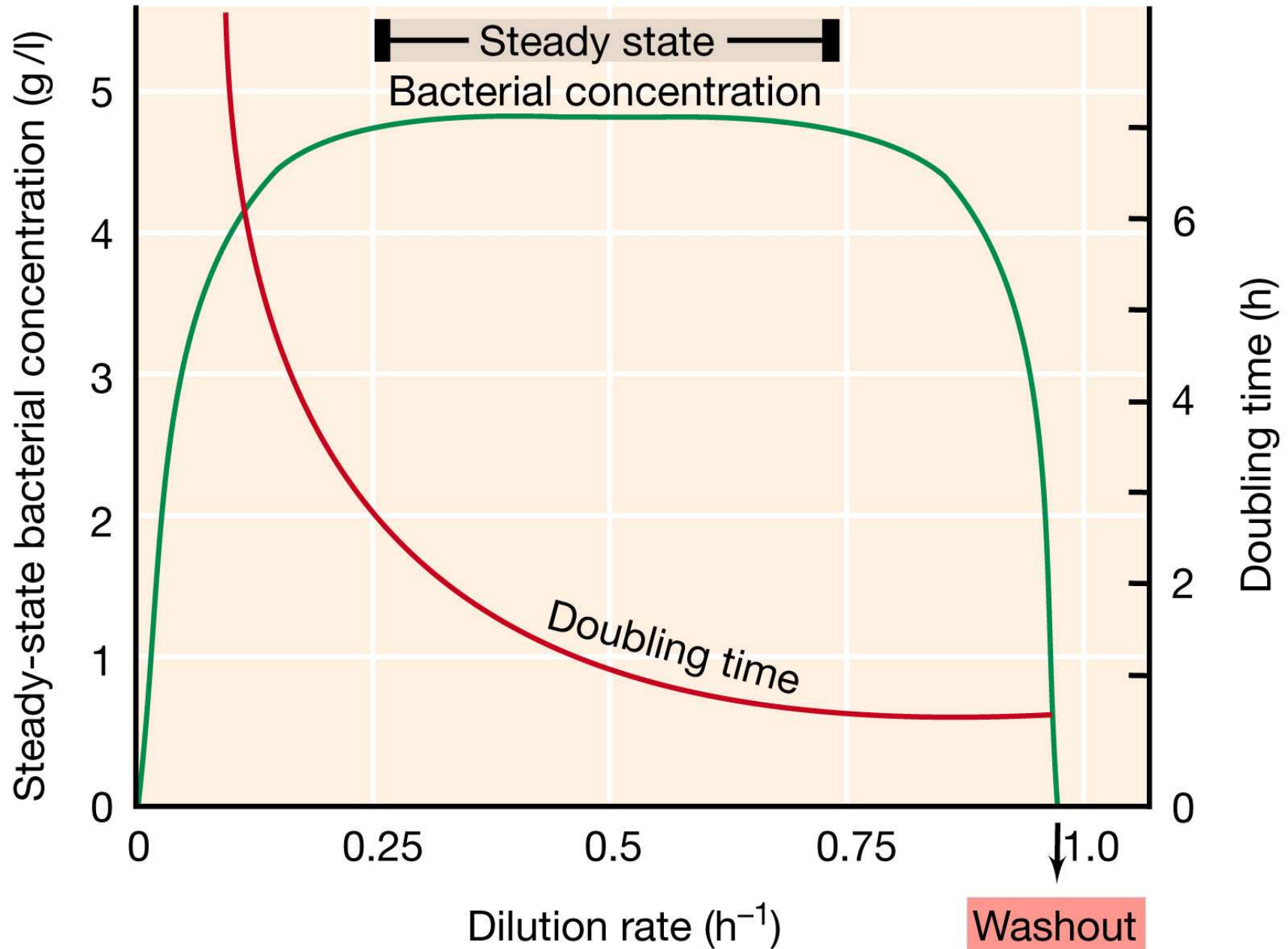
A continuous culture: the chemostat



Relationship between nutrient concentration, growth rate (green curve), and growth yield (red curve)

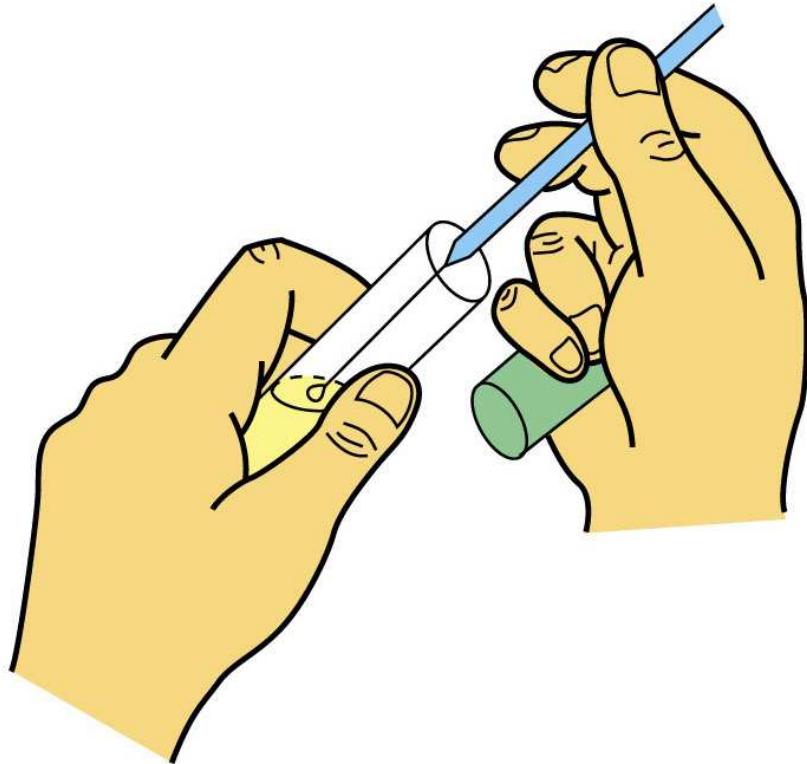


Steady-state relationships in the chemostat



Microbial growth:
the growth on solid media

Growth on solid media: **colonies** and **pure cultures**



(a)



(b)

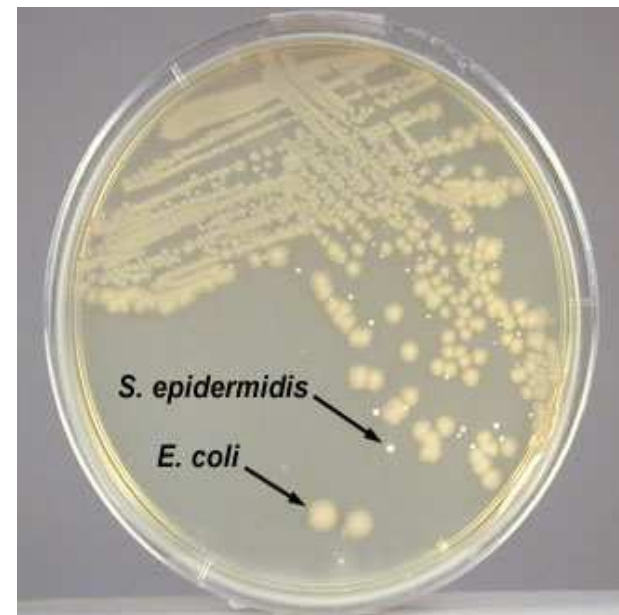
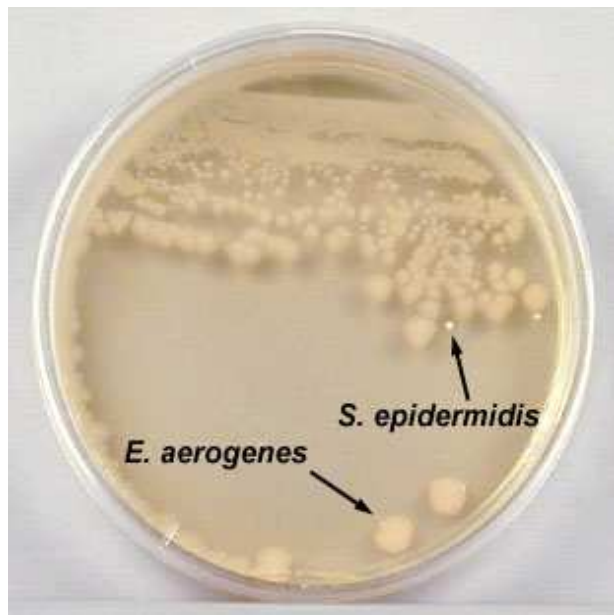
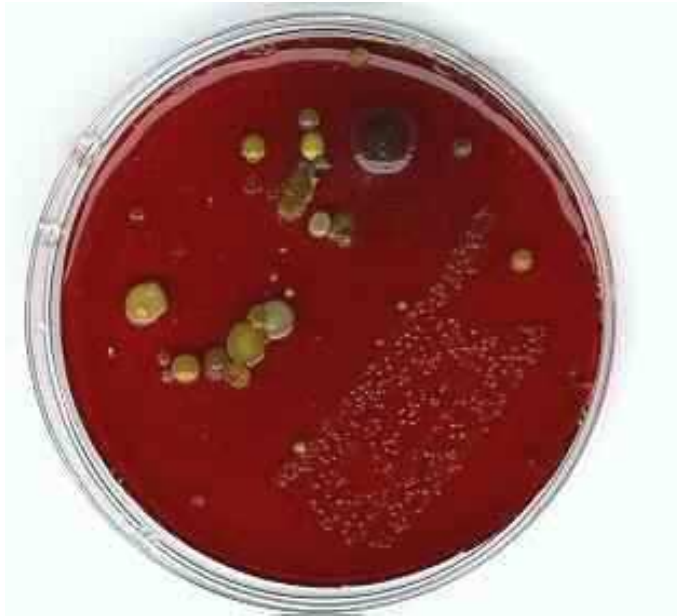
A bacterial colony contains $1-10 \times 10^6$ cells

Procedure:

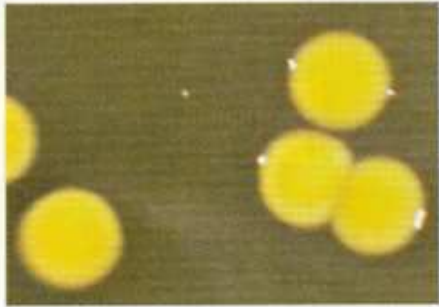
1. Flame the loop and wire and streak a loopful of broth as at **A** in the diagram.
2. Reflame the loop and cool it.
3. Streak as at **B** to spread the original inoculum over more of the agar.
4. Reflame the loop and cool it.
5. Streak as at **C**.
6. Reflame the loop and cool it.
7. Streak as at **D**.
8. Label the plate and incubate it inverted

Streaking a Plate for Isolation





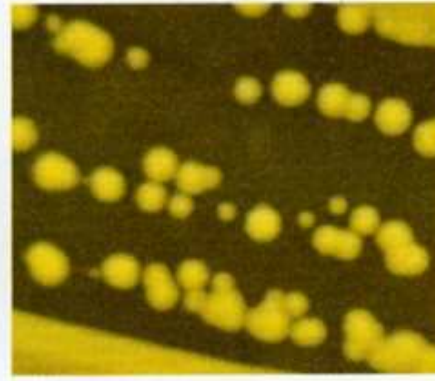
Aspects of bacterial colony morphology



A



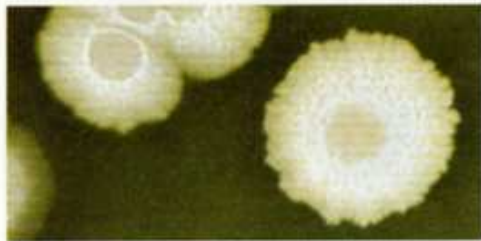
B



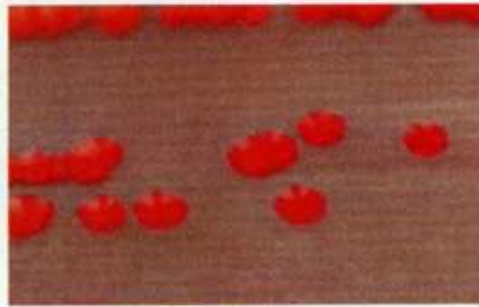
C



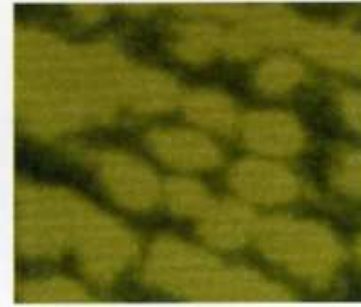
D



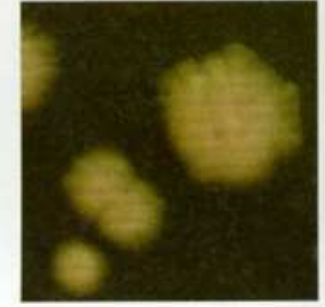
E



F



G



H

Aspects of bacterial colony morphology

Shape



Margin



Elevation



Size



Texture

Smooth or rough

Appearance

Glistening (shiny) or dull

Pigmentation

Nonpigmented (cream, tan, white)

Pigmented (purple, red, yellow)

Optical property

Opaque, translucent, transparent