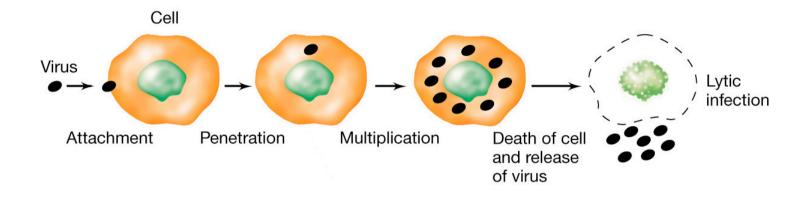
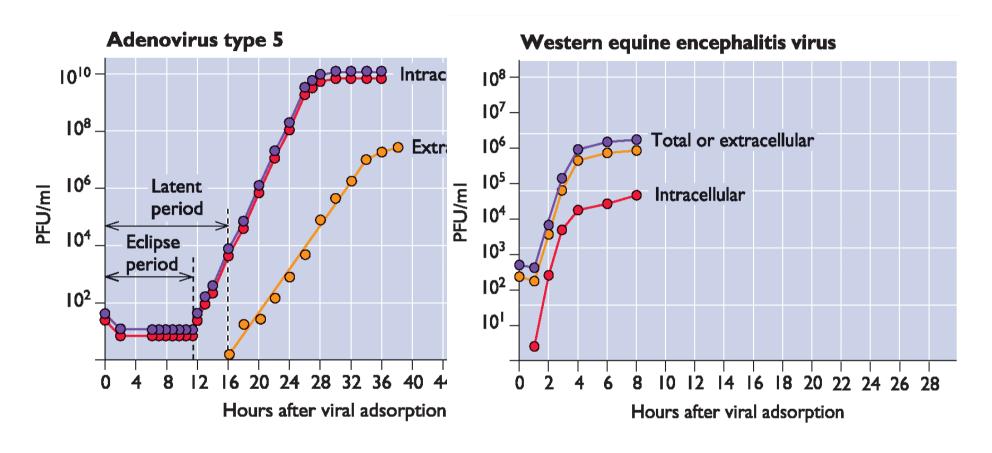
#### **VIROLOGY**

Viral replication



## Possible cytopathogenesis of cells infected with animal viruses

# The one-step growth curve is a fundamental feature of a virus



The time interval from infection to plateau represents the time required for a single cycle of growth.

The yield of virus at plateau shows the amount of virus produced per cell during a single round of infection .

#### The reproductive cycle of animal viruses

- Virus attachment to host cell
- Virus entry into cells
- •Transcription, translation and genome replication
- Assembly, exit and maturation of progeny virions

### **ATTACHMENT PENETRATION VIRAL SYNTHESIS TRANSCRIPTION UNCOATING** TRANSLATION REPLICATION ~~~~ **ASSEMBLY** (MATURATION) **EXIT**

# Viral replication virus attachment to host cells

#### Viral receptors and coreceptors

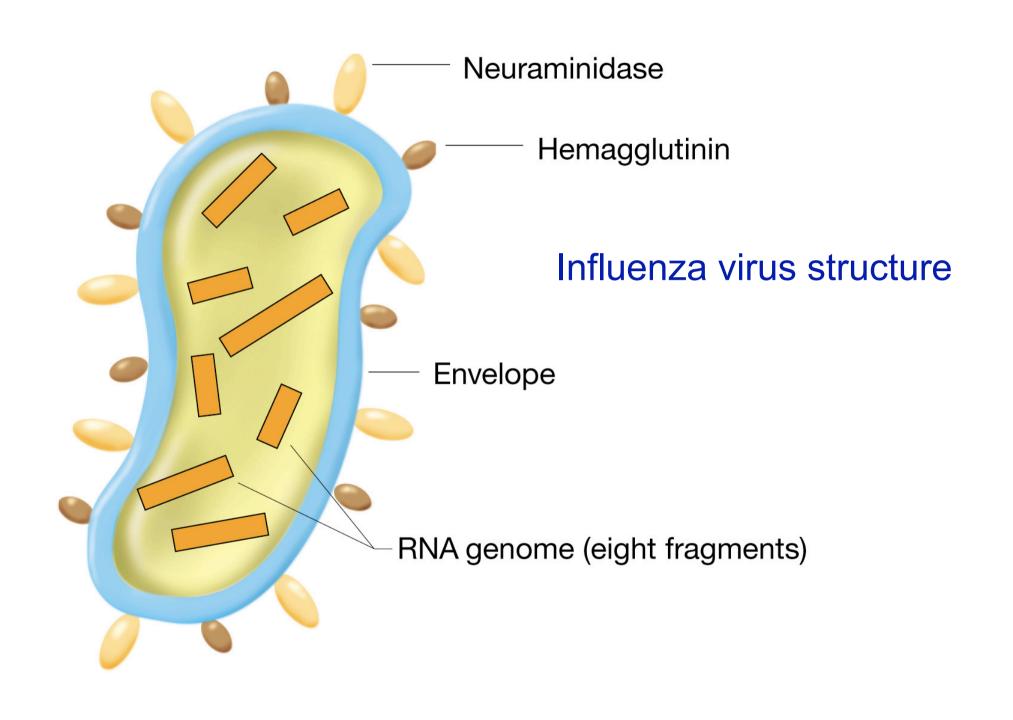
Table 4.1 Viral receptors and coreceptorsa

Table 4.1	Viral receptors and o	coreceptors <sup>a</sup>			
	Virus	Receptor	Type of molecule	Coreceptor	1
Picornavirida	е				
Foot-and-mouth disease virus (cell culture adapted)		Heparan sulfate	Glycosaminoglycan		
Foot-and-mouth disease virus		$\alpha_v \beta_3$ (vitronectin receptor)	Integrin		г
Encephalomyocarditis virus		Vcam-1	Ig-like		
		Sialylated glycophorin A (for hemag- glutination only)	Carbohydrate		L
Poliovirus	type 1 to 3	Pvr	Ig-like		
Coxsackieviruses A13, A18, A21		Icam-1	Ig-like		
Coxsackievirus A21		Decay-accelerating protein (CD55)	SCR-like (complement cascade)	Icam-1	
Coxsackievirus A9		$\alpha_{v}\beta_{3}$	Integrin		
Coxsackieviruses B1 to B6		Car (coxsackievirus-adenovirus receptor)	Ig-like		
Coxsackieviruses B1, B3, B5		CD55	SCR-like (complement cascade)	$\alpha_v^{}\beta_6^{}$ integrin	
Echoviruses 1 and 8		$\alpha_2 \beta_1$ integrin (Vla-2)	Integrin	$\beta_2$ microglobulin	
Echovirus 22		$\alpha_v \beta_3$ (vitronectin receptor)	Integrin		
Echoviruses 3, 6, 7, 11 to 13, 20, 21, 24, 29, 33		CD55	SCR-like (complement cascade)	$\beta_2$ microglobulin	
Enteroviru	ıs 70	CD55	SCR-like (complement cascade)		
Bovine en	terovirus	Sialic acid	Carbohydrate		
Hepatitis A virus		HAVCr-1	Ig-like, mucin-like		
Major group rhinoviruses (91 serotypes)		Icam-1	Ig-like		
Minor group (10 sero		Low-density lipoprotein receptor protein family	Signaling receptor		
Rhinoviru	s 87	Sialic acid	Carbohydrate		
Coronaviridae	2			"	
Mouse he	patitis virus	Bgp (biliary glycoprotein)	Ig-like		
Human coronavirus 229E		Aminopeptidase N	Protease		
Transmissible gastroenteritis virus		* *	Protease		į
Human coronavirus OC43		Sialic acid	Carbohydrate		
Bovine coronavirus		Sialic acid	Carbohydrate		
Togaviridae			-		
Semliki Forest virus		Major histocompatibility class I molecule	Ig-like		
Sindbis virus		High-affinity laminin receptor	Integrin		
Shidois virus		Heparan sulfate	Glycosaminoglycan		
Dengue vi	rue	Heparan sulfate	Glycosaminoglycan		
		neparan sunate	Giycosaniniogiycan		
Rhabdovirida					
Rabies vir	us	Nicotinic acetylcholine receptor	Neurotransmitter receptor		
		Neural cell adhesion molecule CD56	Ig-like		
		Low-affinity nerve growth factor receptor	Tnf receptor protein superfamily		
Paramyxoviri	dae				
Measles vi	irus	Membrane cofactor protein, CD46	Complement-regulating protein		i
Sendai virus		Sialic acid	Carbohydrate		
		Asialoglycoprotein receptor Gp-2	Transport protein (receptor- mediated endocytosis)		
Orthomyxovii	ridae				
	A and B viruses	Sialic acids (N-acetyl neuraminic acid)	Carbohydrate		
Influenza C virus		Sialic acids (9- <i>O</i> -acetyl neuraminic acid)	Carbohydrate		
Arenaviridae			. 2		
Lymphocytic choriomeningitis virus		$\alpha\text{-}Dystroglycan$	Laminin receptor		
Lassa viru	9	α-Dystroglycan	Laminin receptor		
Lassa VIII	<i>3</i>	u-Dysnogrycan	Lammint receptor		<del>,</del>

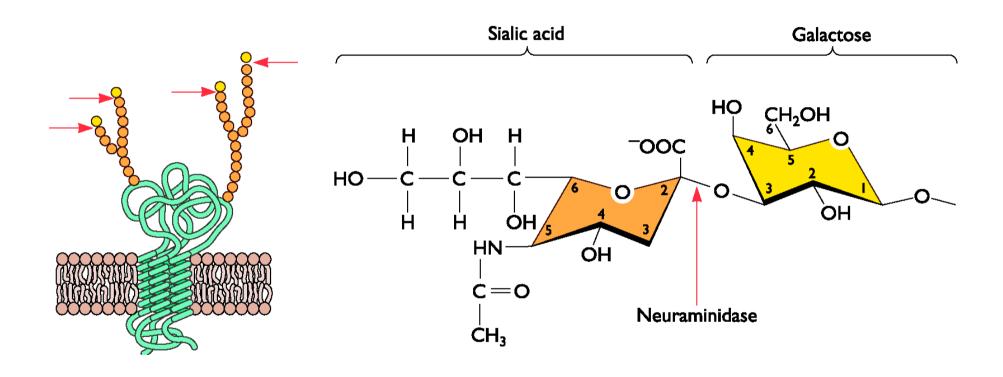
Table 4.1 Viral receptors and coreceptors<sup>a</sup> (continued)

Table 4.1 Viral receptors and	coreceptors <sup>a</sup> (continued)					
Virus	Receptor	Type of molecule	Coreceptor			
Reoviridae	•	••	·			
Reovirus	Sialic acids	Carbohydrate				
Group A porcine rotavirus	Sialic acids	Carbohydrate				
Retroviridae						
Human immunodeficiency	CD4	Ig-like	Chemokine receptors			
virus type 1	CD4	ig-like	(Ccr5, Cxcr4, Ccr3)			
**	Galactosylceramide	Glycolipid				
Human immunodeficiency	CD4	Ig-like	Chemokine receptors			
virus type 2						
	Cxcr4	7-transmembrane superfamily				
Simian immunodeficiency	CD4	Ig-like	Chemokine receptors			
virus	Cl1	C-4: 4 4 b				
Gibbon ape leukemia virus	Glvr1	Sodium-dependent phosphate transport protein				
Feline leukemia virus B	Glvr1	Sodium-dependent phosphate				
Temie realema (nao 2	G.,,,,	transport protein				
Amphotropic murine leukemia	Ram-1	Sodium-dependent phosphate				
virus		transport protein				
Ecotropic murine leukemia	Cat	Cationic amino acid transport				
virus	The same of the sa	protein				
Subgroup A avian leukosis and sarcoma virus	Tva	Low-density lipoprotein receptor protein family				
Subgroup B and D avian	Carl	Tnf receptor family protein				
leukosis and sarcoma viruses	Sur 1	superfamily				
Bovine leukemia virus	BLVRcp 1	Unknown				
Feline immunodeficiency virus	Cxcr4	7-transmembrane superfamily				
Visna virus	Major histocompatibility complex class II molecule	Ig-like				
Parvoviridae						
Bovine parvovirus	Sialic acids	Carbohydrate				
Adeno-associated virus type 2	Heparan sulfate	Glycosaminoglycan	α β, integrin			
Papovaviridae	<b>F</b>		-VI-3 D			
Simian virus 40	Major histocompatibility class I molecule	Ig-like				
	wajor instocompatibility class i molecule	лд-п <b>к</b> с				
Adenoviridae		* 10				
Adenovirus subgroups A, C, D, E, F	Car	Ig-like	$\alpha_{_{_{f v}}}$ integrins			
Adenovirus type 5	Major histocompatibility class II molecule	Ig-like	α, integrins			
(subgroup C)	valor instocompanionery causs it morecure	ag mac	w micginis			
Adenovirus type 2	$\alpha_{M}\beta_{2}$	Integrin	$\alpha_v$ integrins			
(subgroup C)						
Adenovirus type 9	$\alpha_{_{_{f v}}}$ integrins	Integrin				
(subgroup D)						
Herpesviridae						
Herpes simplex type 1	Heparan sulfate	Glycosaminoglycan	HveA, Prrl			
Herpes simplex type 2	Heparan sulfate	Glycosaminoglycan	HveA, Prr1, Prr2			
Pseudorabies virus	Heparan sulfate	Glycosaminoglycan	Pvr, Prr1, Prr2			
Bovine herpesvirus 1	Heparan sulfate	Glycosaminoglycan	Pvr, Prr1			
Human herpesvirus 7	CD4	Ig-like				
Epstein-Barr virus	Complement receptor Cr2 (CD21)	SCR-like (complement cascade)				
Human cytomegalovirus	Heparan sulfate	Glycosaminoglycan	Aminopeptidase N (CD13)			
Poxviridae						
Vaccinia virus	Heparan sulfate	Glycosaminoglycan				
vaccina virus	Epidermal growth factor receptor	Signaling receptor				
·	Epidermai growm factor receptor	Digitamik receptor				

The name of the receptor and the type of molecule are listed for selected viruses. When coreceptors have been identified, they are listed; a blank in the coreceptor column indicates that none have been identified to date. Abbreviations: Vcam, vascular cell adhesion molecule; Prrl, Prr2, Pvr-related proteins 1 and 2; SCR, short consensus repeat; Ig, immunoglobulin; Tnf, tumor necrosis factor; Car1, cytopathic avian leukosis and sarcoma virus receptor; Car, coxsackievirus and adenovirus receptor; HveA, herpesvirus entry mediator.

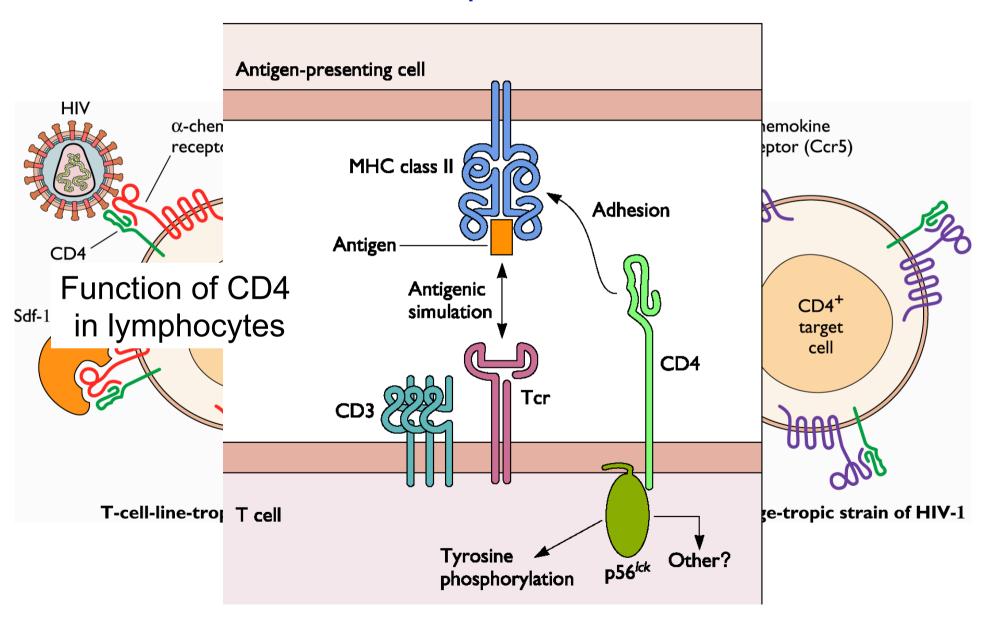


#### Sialic acid receptors for influenza virus

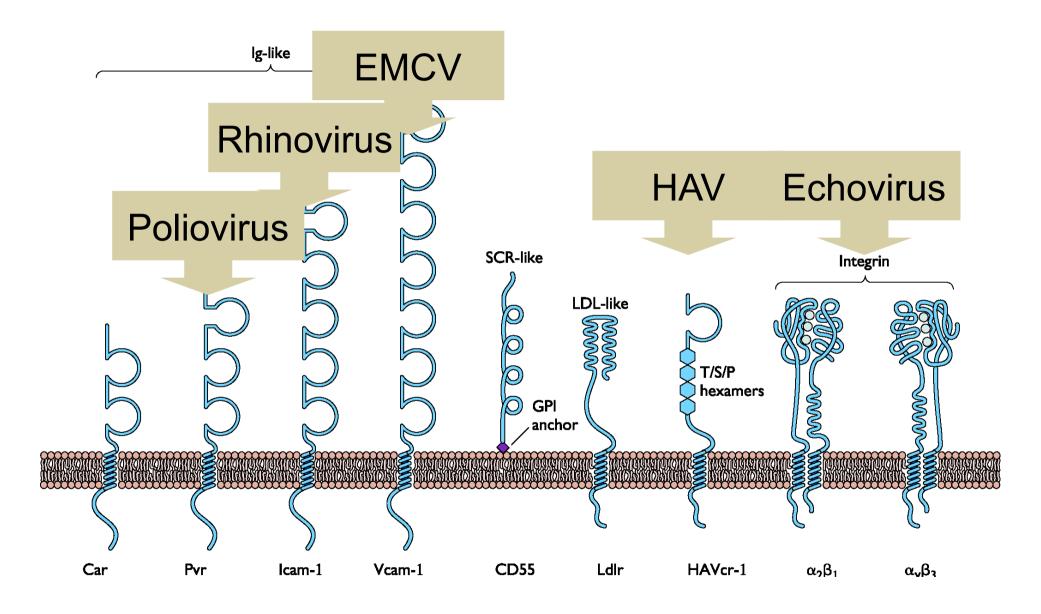


The interaction of influenza virus with sialic acid moieties is mediated by the viral surface glycoprotein hemagglutinin (HA)

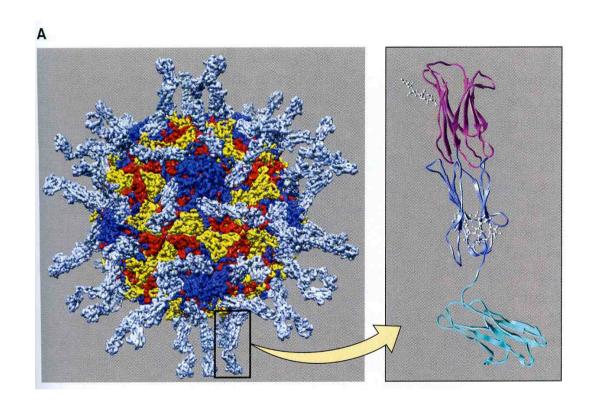
#### Receptor and coreceptors for macrophage/monocyteand T-cell- tropic strains of HIV-1

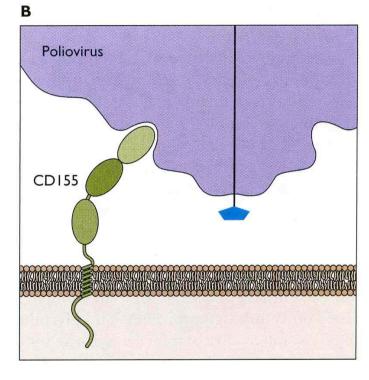


#### Cell receptors for picornaviruses

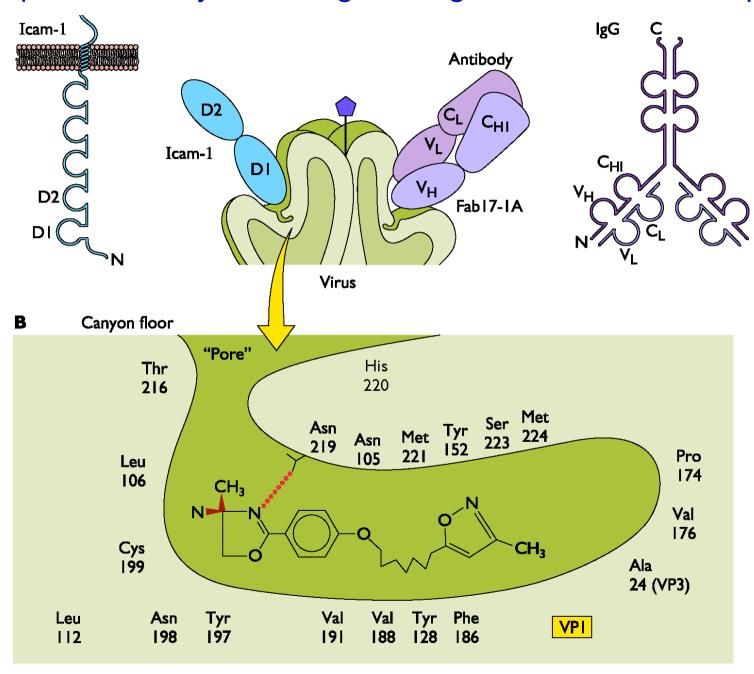


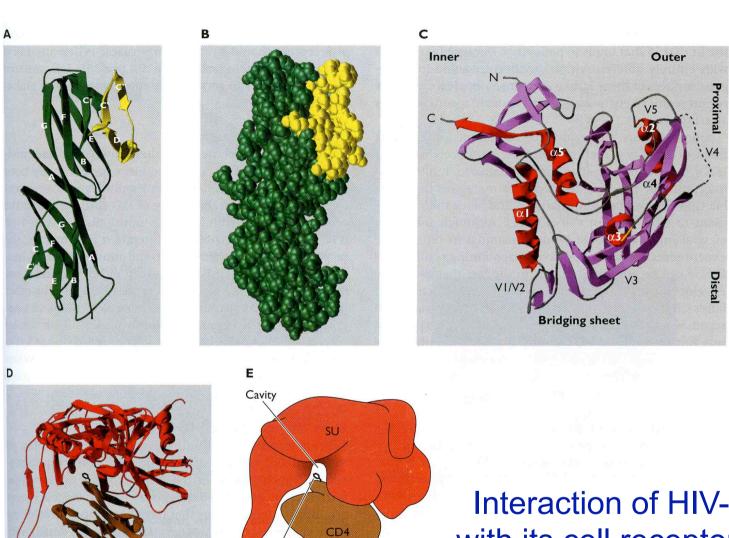
#### Poliovirus-receptor interactions





#### Receptor, antibody, and drug binding to the rhinovirus capsid

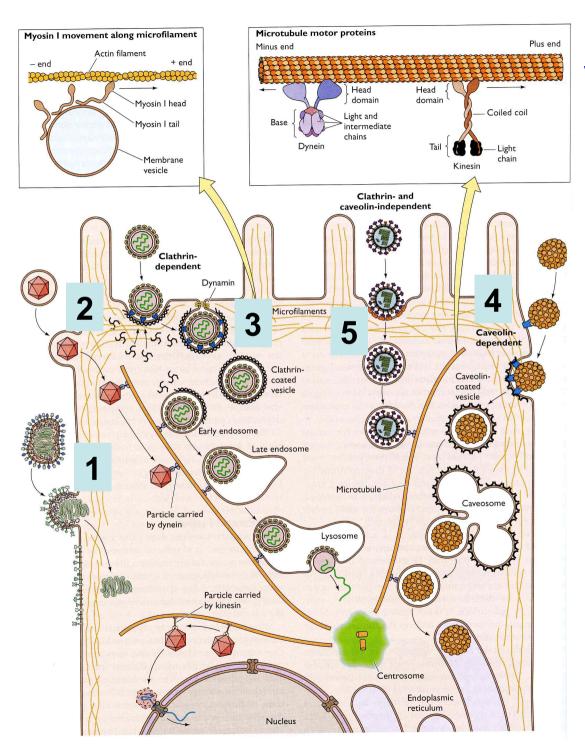




Phe43

Interaction of HIV-1 SU with its cell receptor, CD4

# Viral replication virus entry into host cells

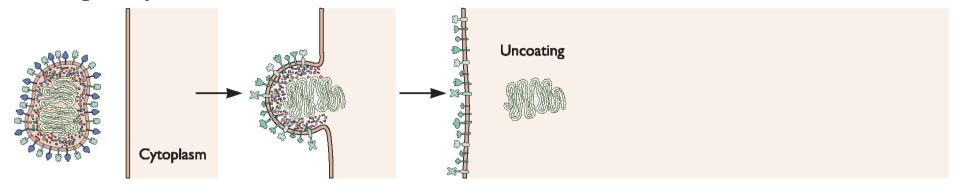


#### Virus entry strategies

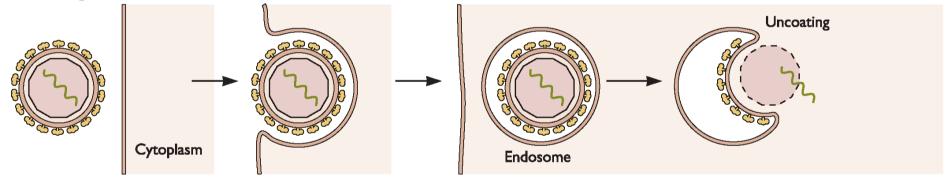
- 1. Entry and uncoating at the plasma membrane
- 2. Entry at the **plasma membrane** and uncoating at the **nuclear membrane**
- 3. Entry by **clathrin-dependent** endocytosis
- 4. Entry by **caveolin-dependent** endocytosis (raft-mediated)
- 5. Entry clathrin- and caveolinindependent endocytosis

#### Three entry and uncoating strategies

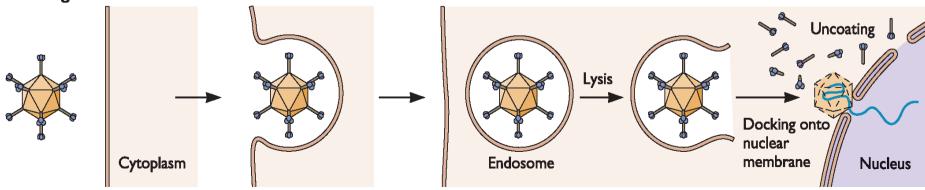
#### Uncoating at the plasma membrane

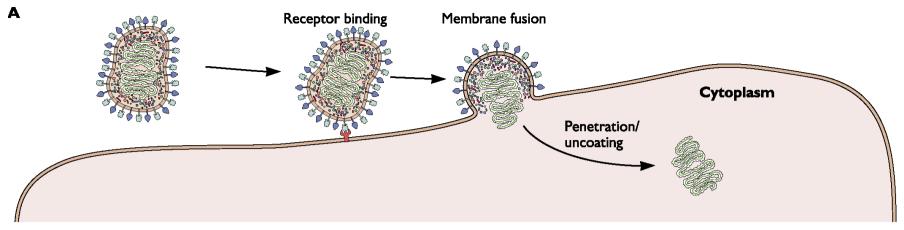


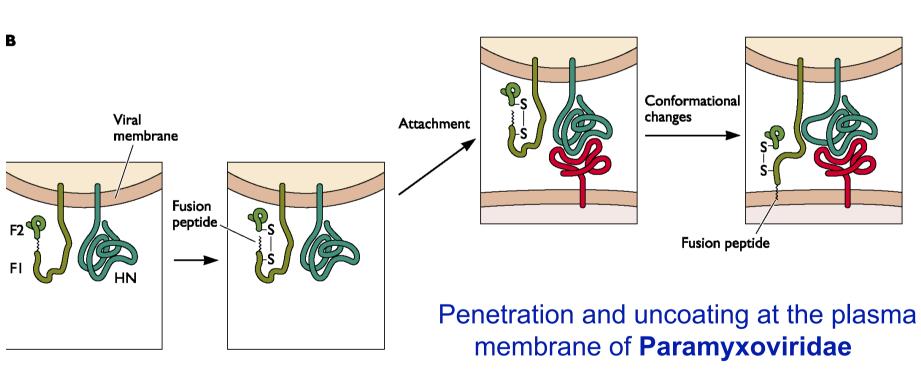
#### Uncoating within endosomes



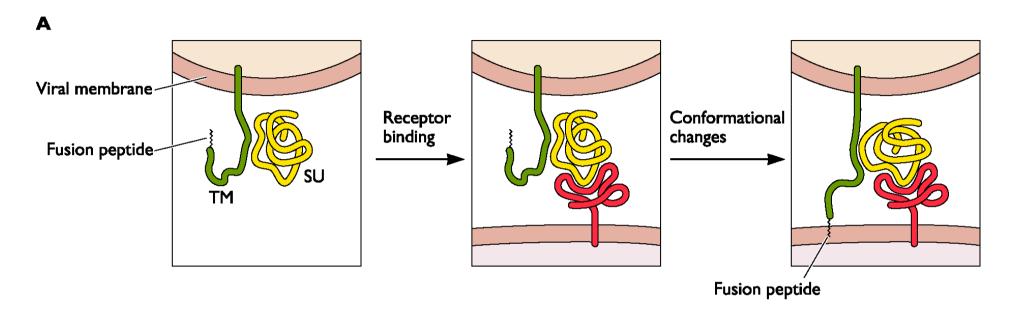
#### Uncoating at the nuclear membrane







#### Mechanism of retroviral fusion with the plasma membrane

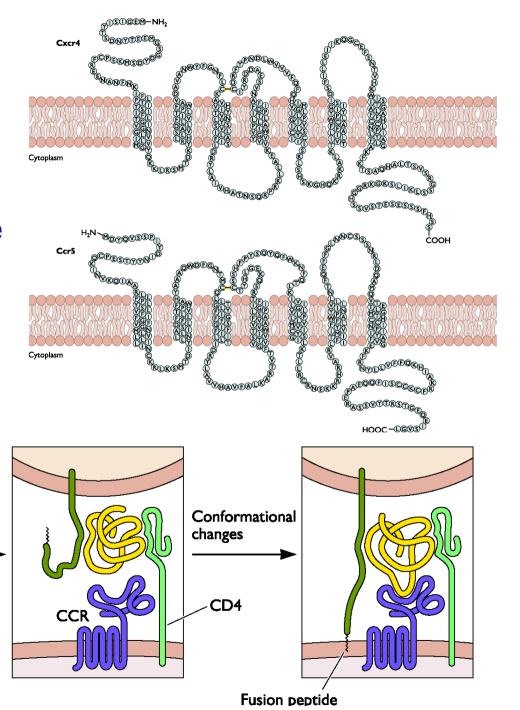


# Mechanism of HIV-1 fusion with the plasma membrane

Receptor binding

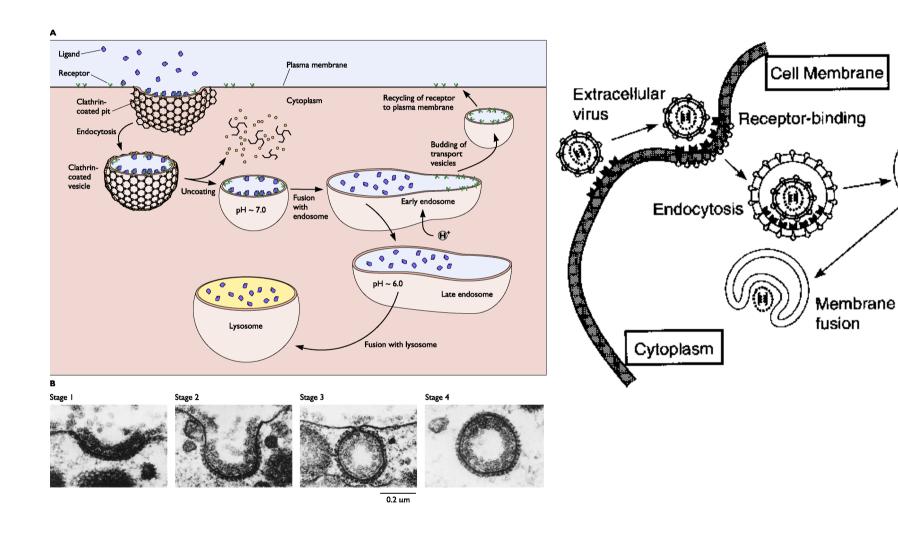
Viral membrane

Fusion peptide-

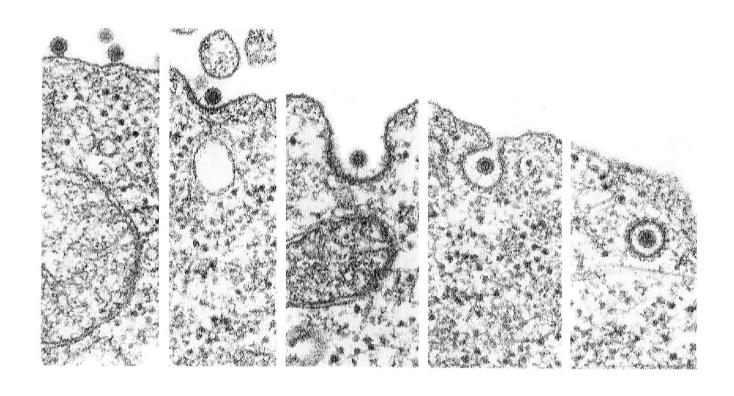


#### Mechanism of uncoating within endosomes

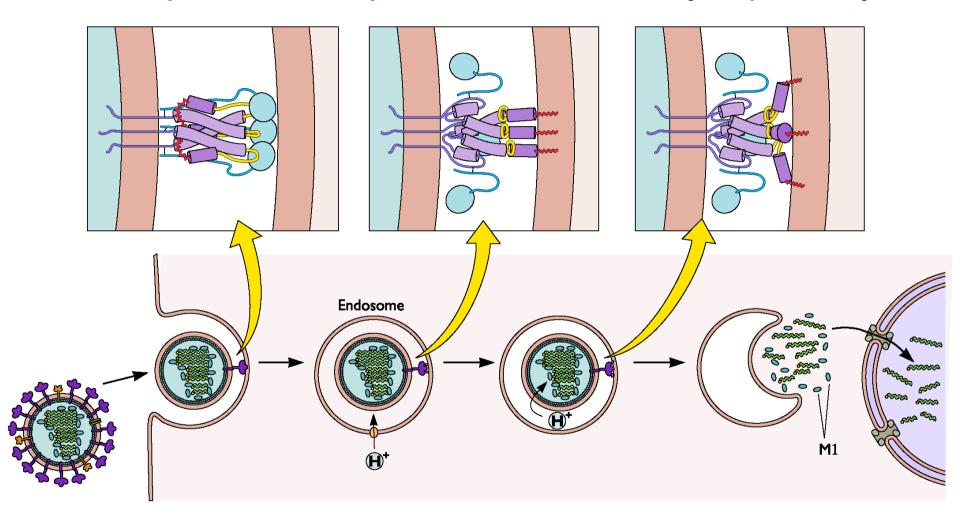
Acidification of vesicles



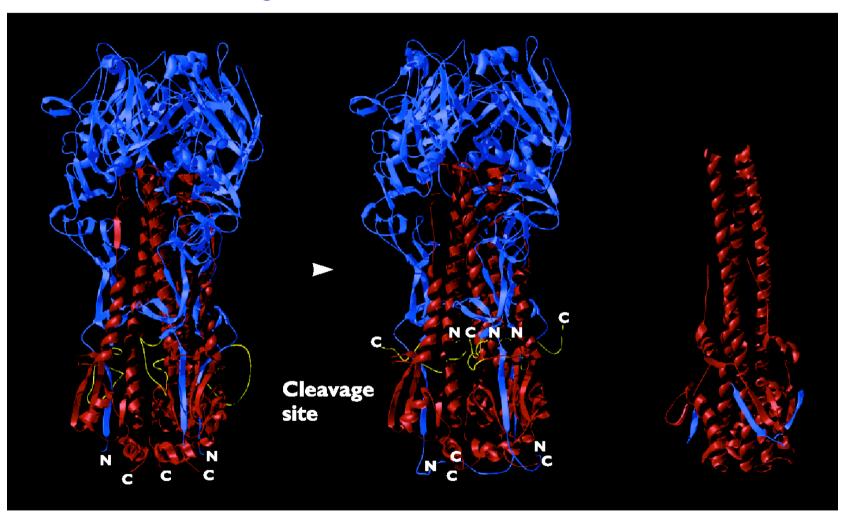
#### Virus entry via receptor-mediated endocytosis



#### Influenza virus: an example of virus entry via the clathrindependent receptor-mediated endocytic pathway



# Cleavage- and low-pH-induced structural changes in the Influenza virus HA

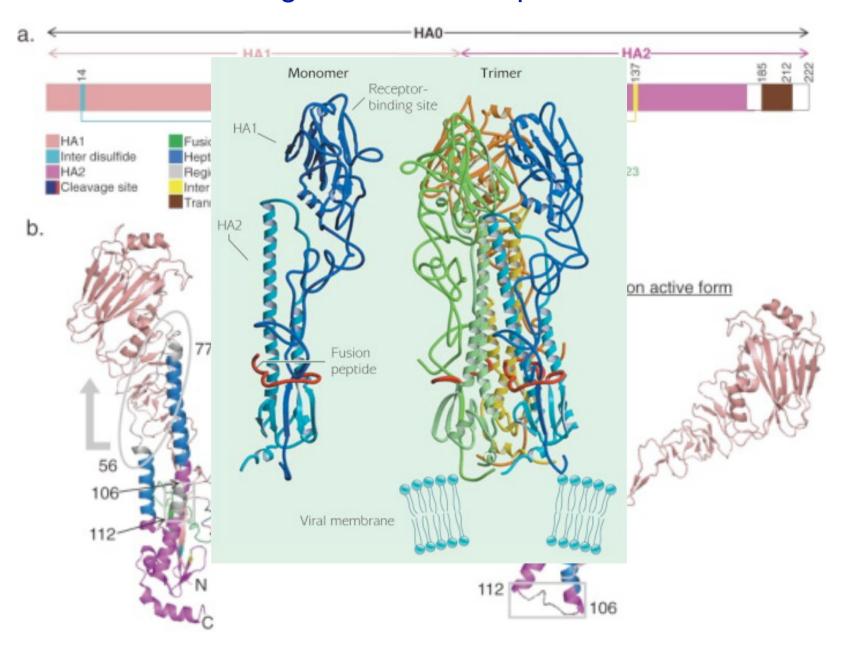


**Uncleaved HAO precursor** 

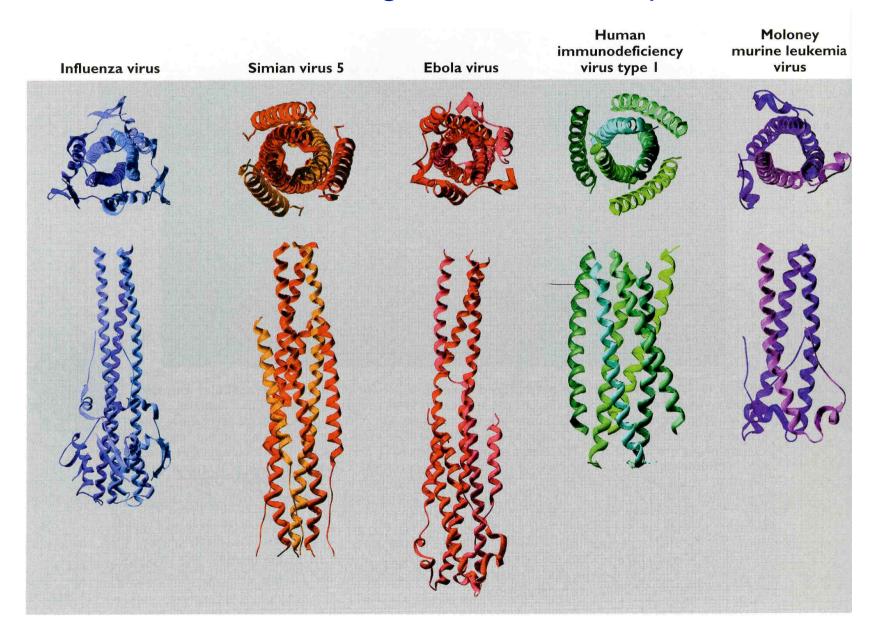
Structure of the HA trimer at neutral pH

Structure of the low pH trimer (only HA2 is shown)

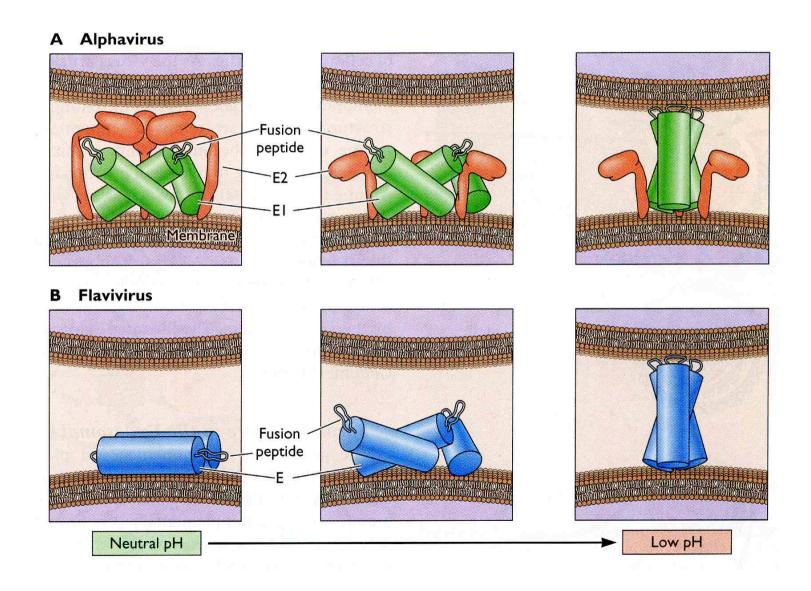
#### Conformational changes of HA at the pH of membrane fusion

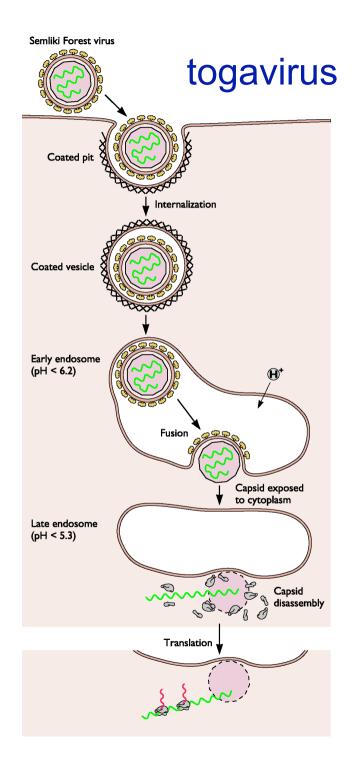


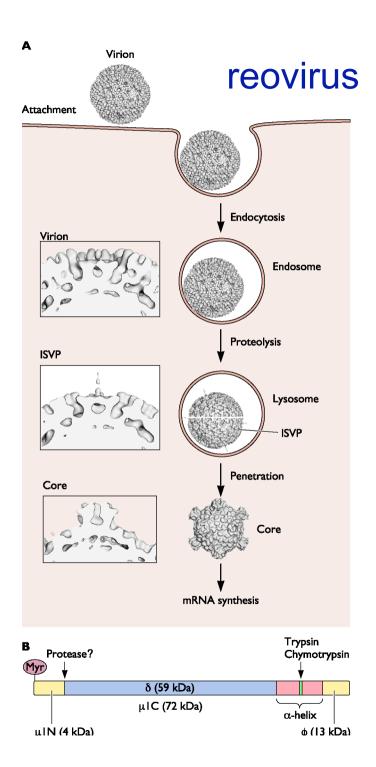
#### Similarities among five viral fusion protein



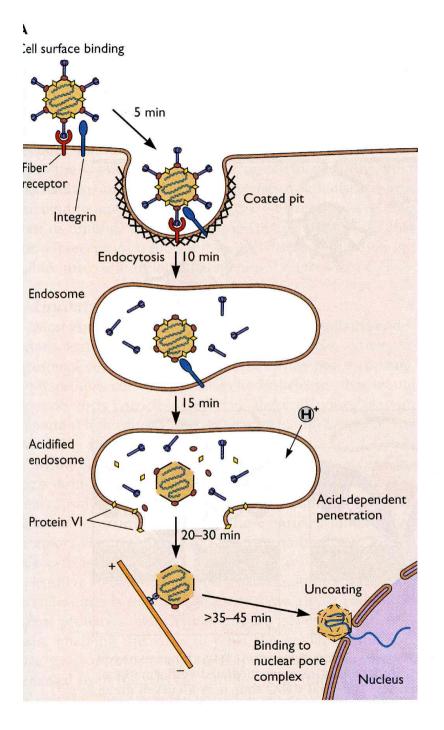
# Model for low-pH-induced movement of alphavirus and flavivirus glycoproteins

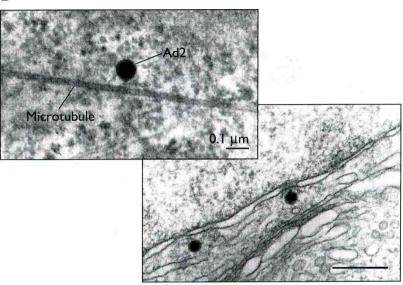






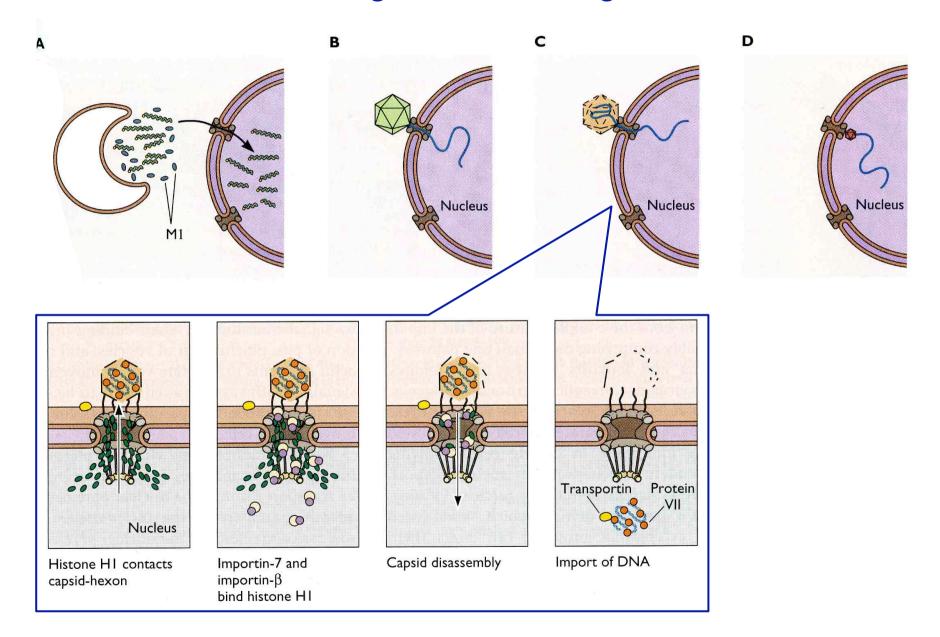




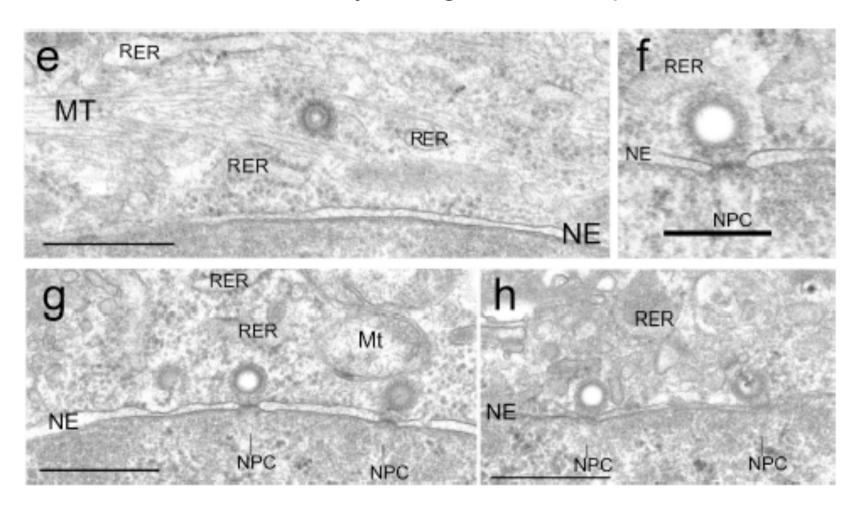


Adenovirus: an example of virus entry via the clathrin-dependent receptor-mediated endocytic pathway and uncoating at the nuclear membrane

#### Different strategies for entering the nucleus

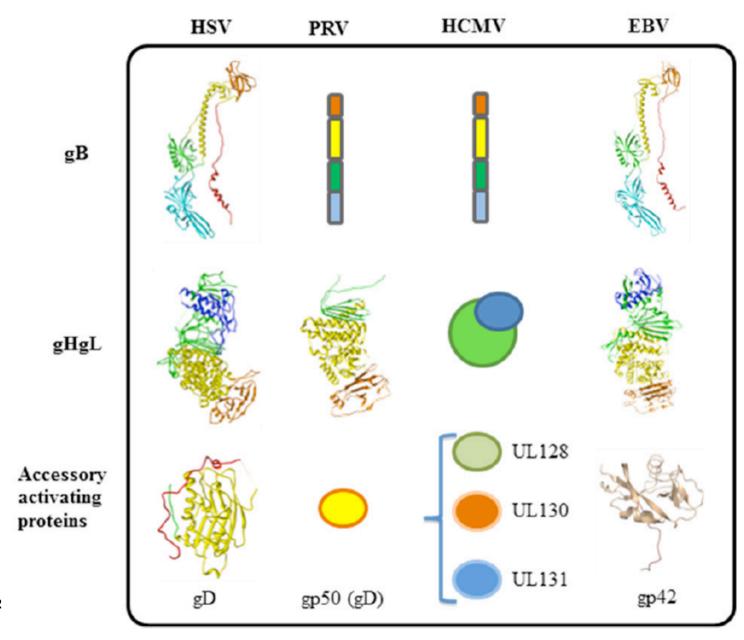


# The microtubule network mediates nuclear targeting of Human Cytomegalovirus capsids



# Viral replication virus entry into host cells: Herpesvirus entry

#### Fusion machinery of Herpesviruses



Eisenberg et al., 2012

#### The basic steps in Herpes simplex virus entry

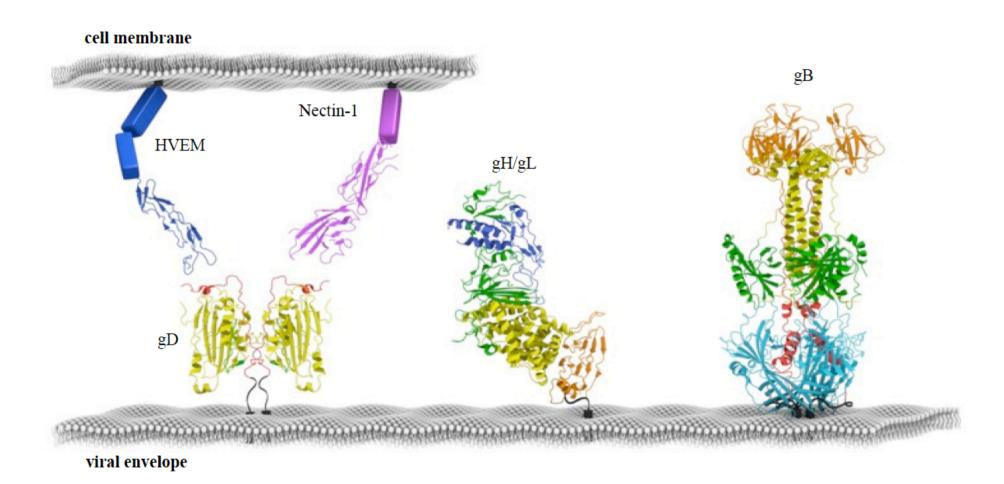
Herpes simplex virus enter cells via direct fusion with the plasma membrane at neutral pH, or by endocytosis.

The fusion is a process that consists of three basic steps carried out by the virion glycoproteins:

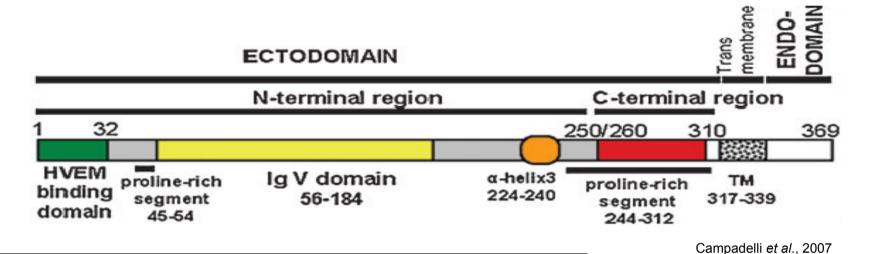
- 1) recognition of a cellular receptor by a viral glycoprotein
- 2) triggering of fusion
- 3) fusion execution

These steps are carried out by four essential virion glycoproteins: **gB**, **gD**, **gH** and **gL**.

#### Fusion machinery of HSV-1



#### Structure and function of HSV gD glycoprotein



•369 aa

organized in three domains:

1) ecto-domain: aa 1-317

N-terminal: aa 1-260

core: aa 56-184;

 $\alpha$ -helix, aa 224-240

C-terminal: aa 261-310

gD hairpin

2) trans-membrane domain: aa 318-339

3) endo-domain: aa 340-369

gD functions:

1) receptor recognition

2) triggering of fusion

#### Functions of HSV gD: receptor recognition

#### The three natural gD receptors:

#### 1. herpesvirus entry mediator (HVME):

tumor necrosis factor receptor family; in T-lymphocytes or lymphoyd organ.

HVEM binding-site: aa 1-32 (contact residues between aa 7-15 and 24-32).

#### 2. nectin 1:

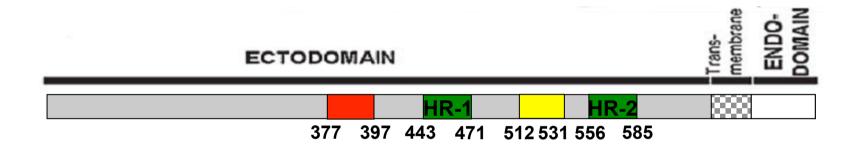
intercellular adhesion molecules family; in sensory neurons, muco-epithelia or epithelia cells. nectin-1 binding-site: critical aa residues (aa 34, 38, 215 and aa 222-223).

#### 3. O-sulphated HS (heparan sulfate):

modified heparan sulfate by enzymes in neuronal and endothelial cells, corneal fibroblasts.

#### Structure and function of HSV gH

HSV-1 gH exibits structural and functional features typical of class I viral fusion gp

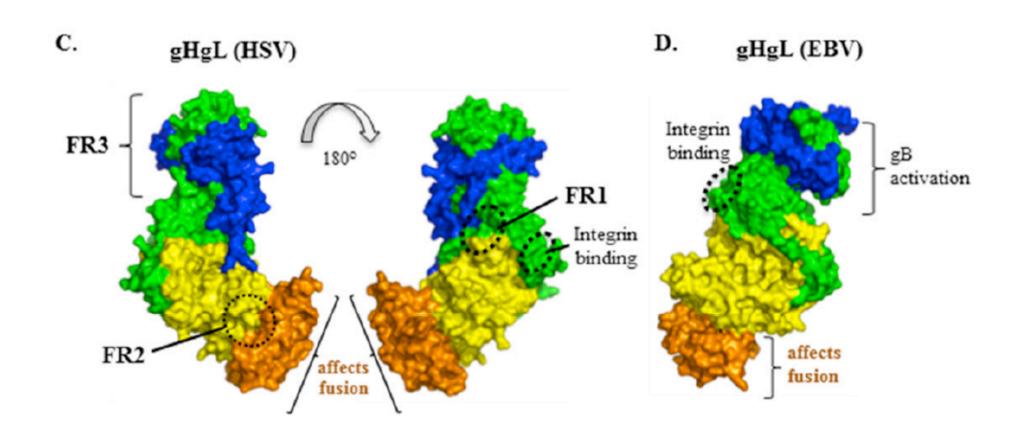


- 838 aa
- organized in three domains:
- 1) ecto-domain heptad repeats: HR-1 (aa 443-471) HR-2 (aa 556-585);
- 2) trans-membrane domain
- 3) endo-domain

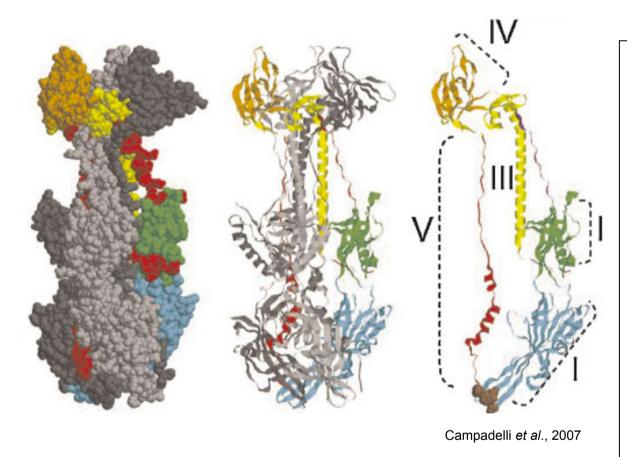
#### gH functions:

- 1) interactions with gD and gB
- 2) triggering of fusion

#### Structure and function of HSV gH



#### Structure and function of HSV gB: an effector of membrane fusion



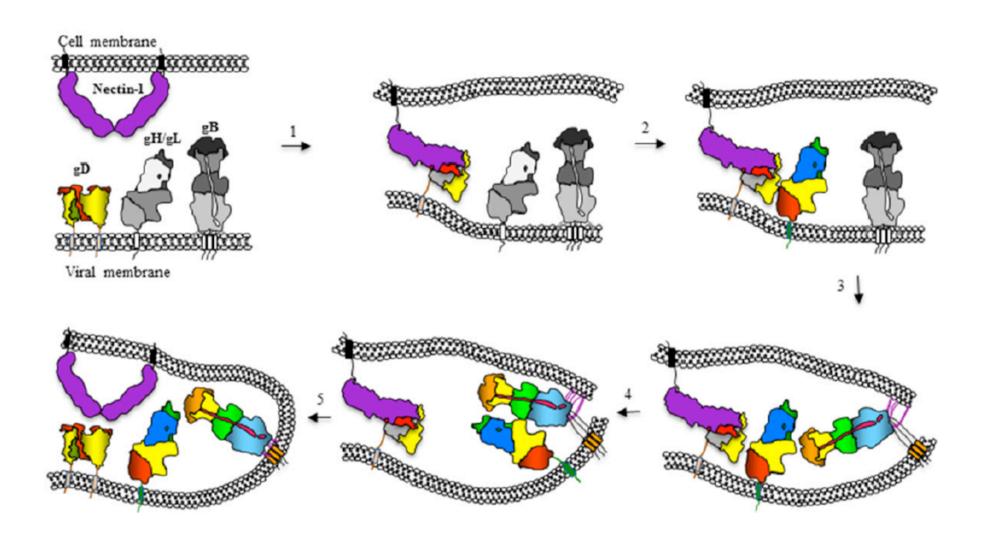
- 904 aa
- trimer with a coiledcoil core
- organized in three domains:
  - 1) ecto-domain: 696 aa α-helix III, HR-1 (aa 92-112) HR-2 (618-631)
  - 2) trans-membrane domain:69 aa
  - 3) endo-domain: 109 aa

#### gH functions:

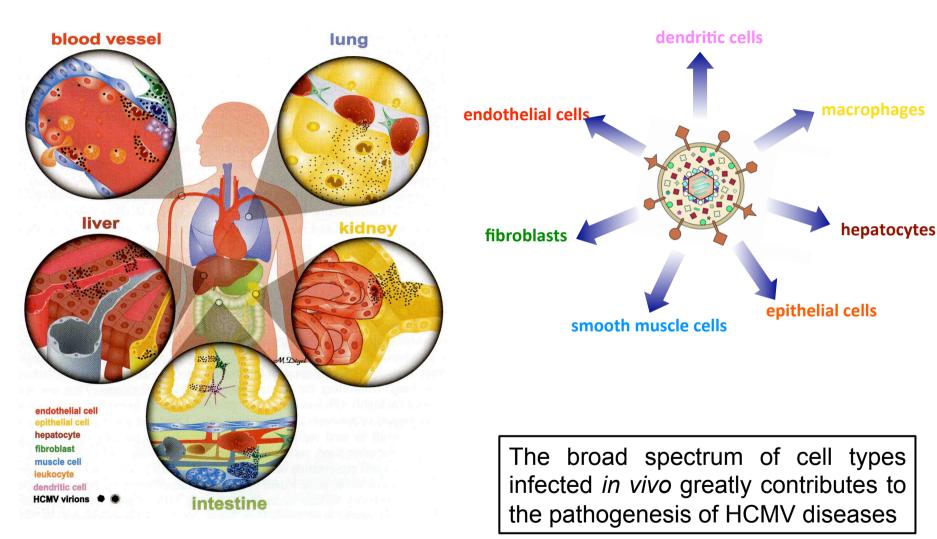
- 1) Binding to HSPG
- 2) Fusion execution

**HSV-1 gH** exibits structural and functional features typical of Class III viral fusion gp

#### Working model for HSV entry into cells

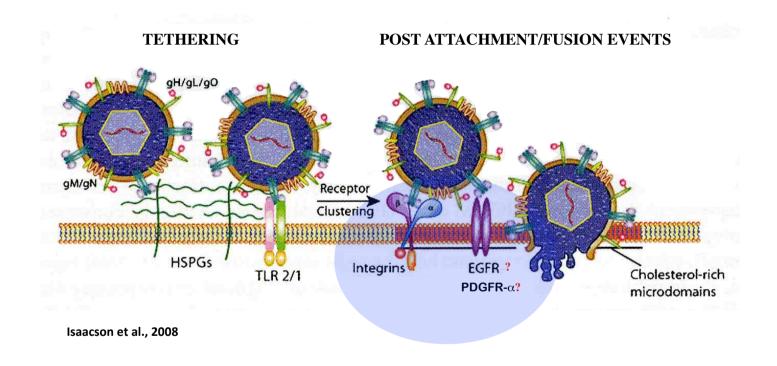


#### The broad cell tropism of HCMV



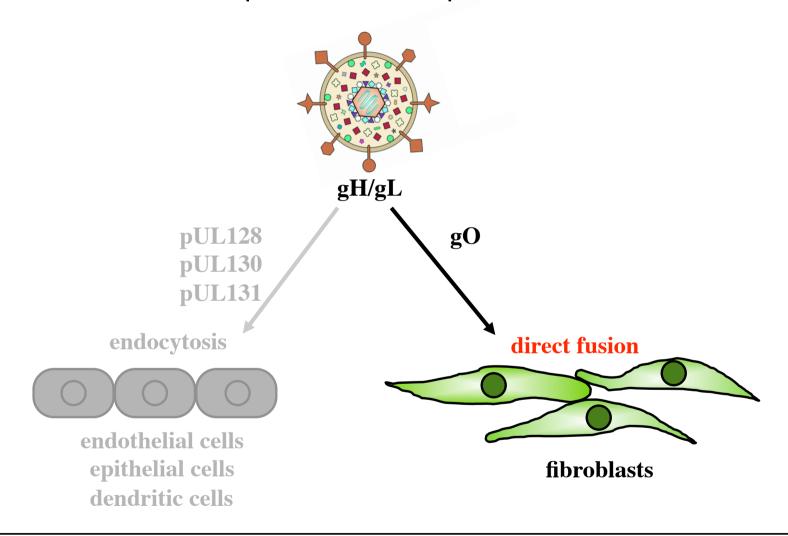
Sinzger et al., 2008

#### The broad cell tropism of HCMV: different receptor utilization



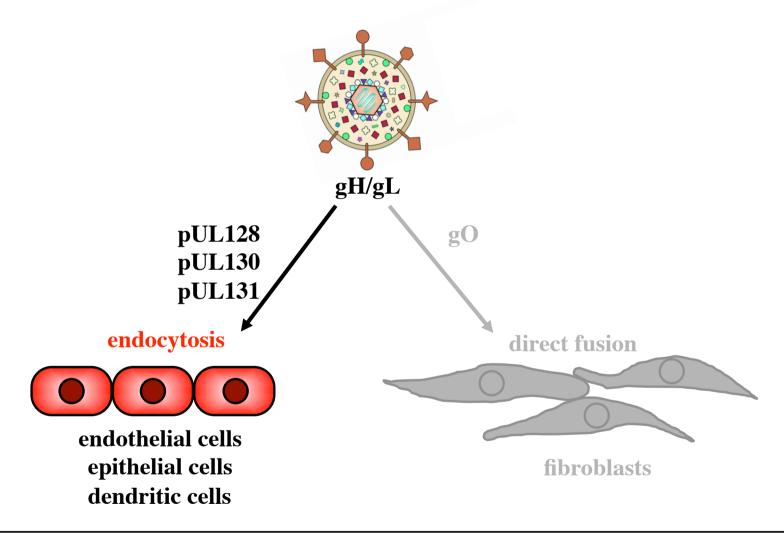
Presence of multiple and/or ubiquitously expressed cellular receptors

## The broad cell tropism of HCMV: different envelope glycoproteins and tropism factors requirements



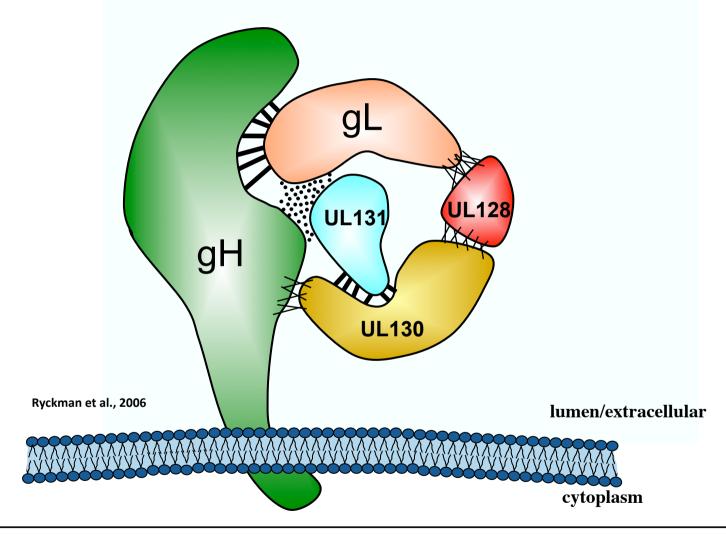
Presence of modular envelope complexes that mediate viral entry in different cell types by different pathways

## The broad cell tropism of HCMV: different envelope glycoproteins and tropism factors requirements



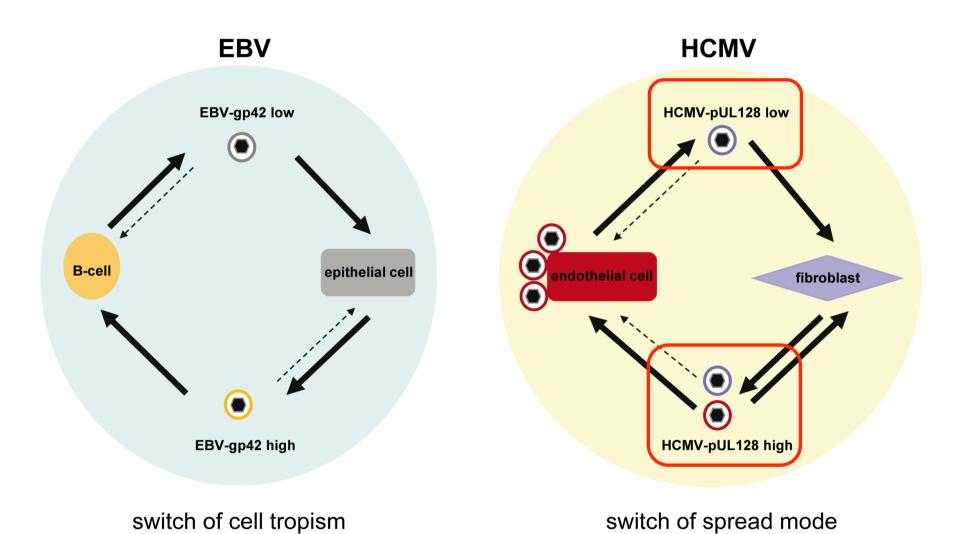
Presence of modular envelope complexes that mediate viral entry in different cell types

#### The broad cell tropism of HCMV: the role of pUL tropism factors



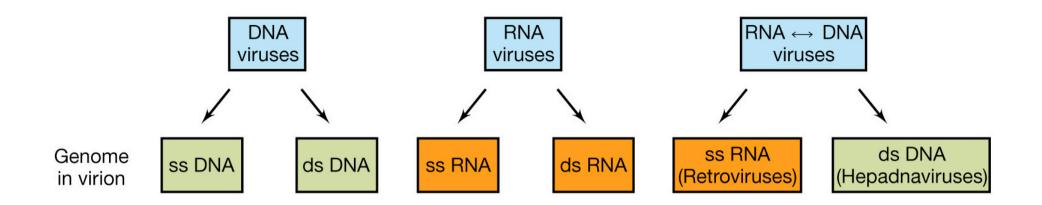
pUL (UL128, UL130 and UL131) proteins assemble onto a gH/gL scaffold to form a virion complex that mediates entry in epithelial and endothelial cells.

### The presence of different Herpesvirus envelope complexes may switch route of infection in vivo



Scrivano et al., 2011

# Viral replication transcription, translation and genome replication



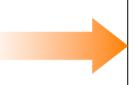
#### Genomes of DNA viruses

- •unimolecular
- ds or ss
- •5-240 kb (1.2 Mb NCLDV)
- •linear or circular

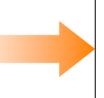
#### Genomes of RNA viruses

- unimolecular or segmented
- •ss or ds
- •1,7-30 kb
- linear or circular
- •(+ )or (-) polarity

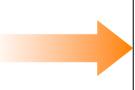
#### Synthesis of viral macromolecules



The diversity of viral transcription strategies



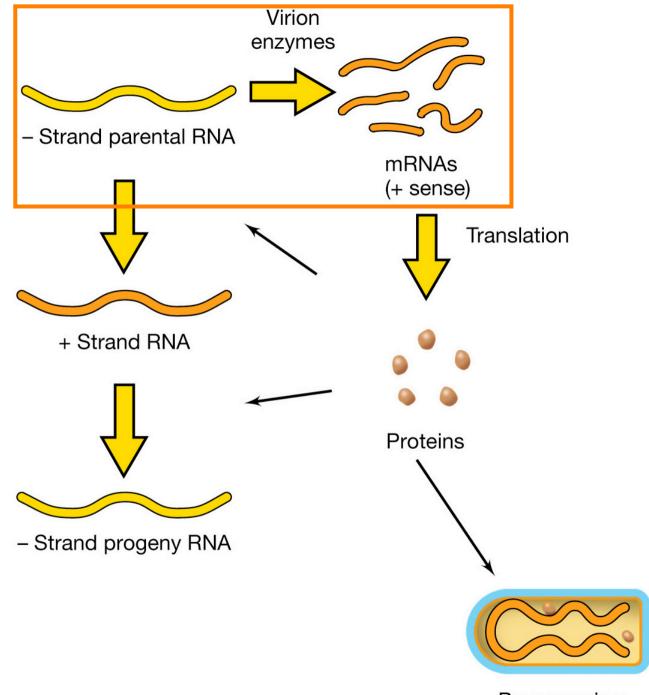
The diversity of viral translation strategies



The diversity of viral genome replication strategies

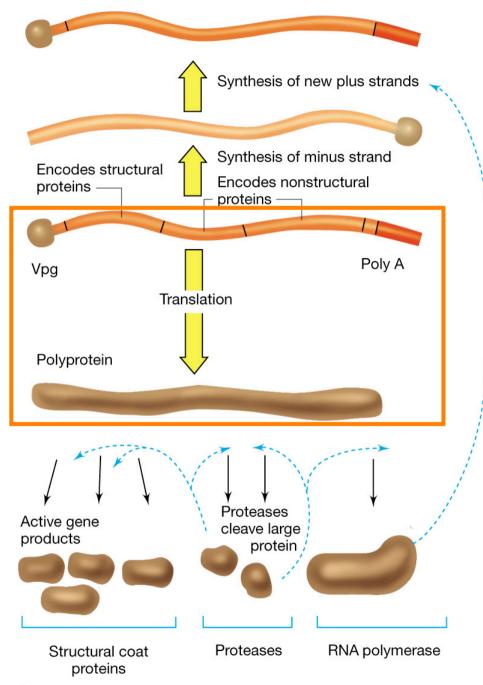
The diversity of viral transcription and translation strategies:

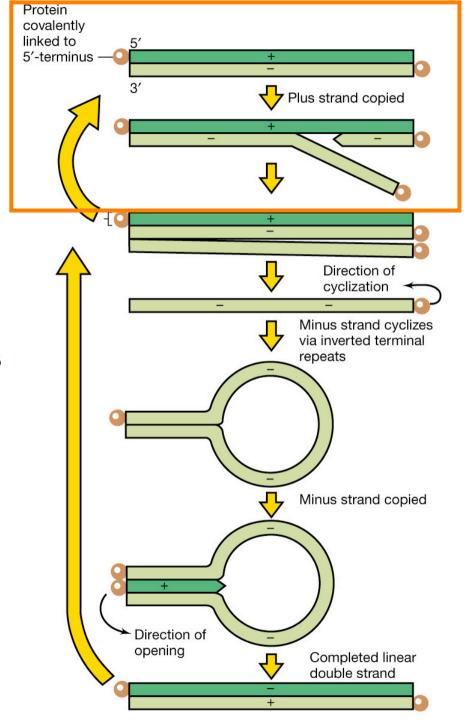
Rhabdoviruses
(-) ssRNA



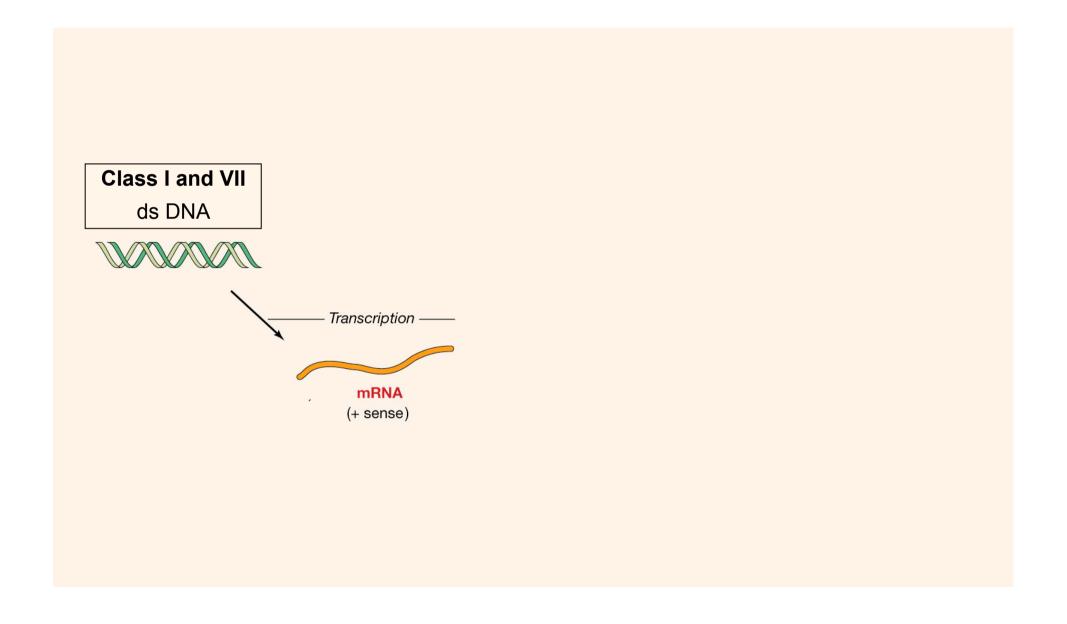
Progeny virus

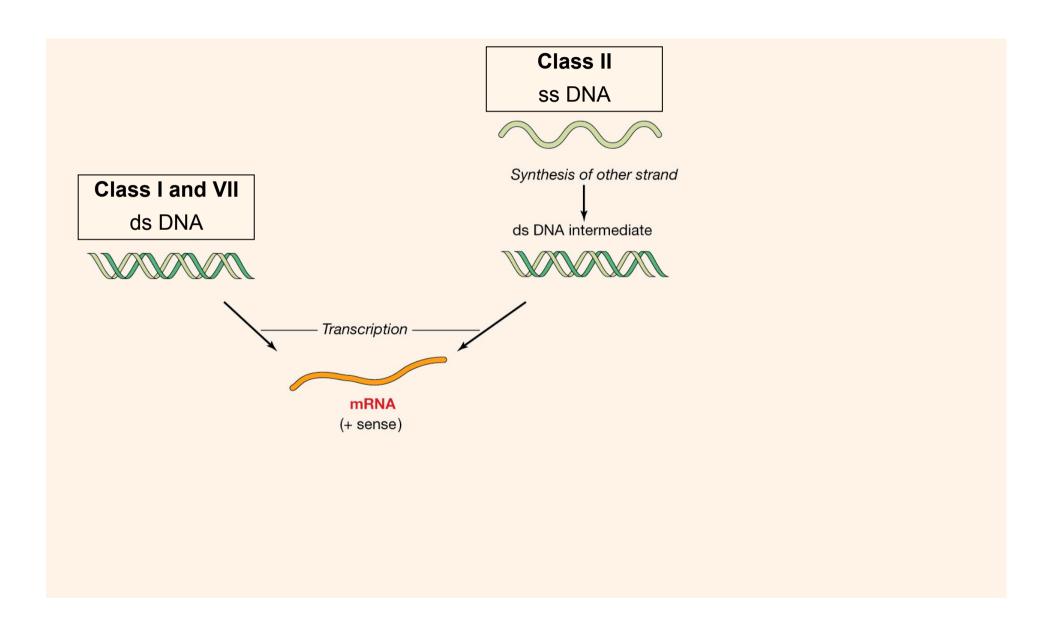
# The diversity of viral translation strategies: Picornaviruses polyprotein synthesis

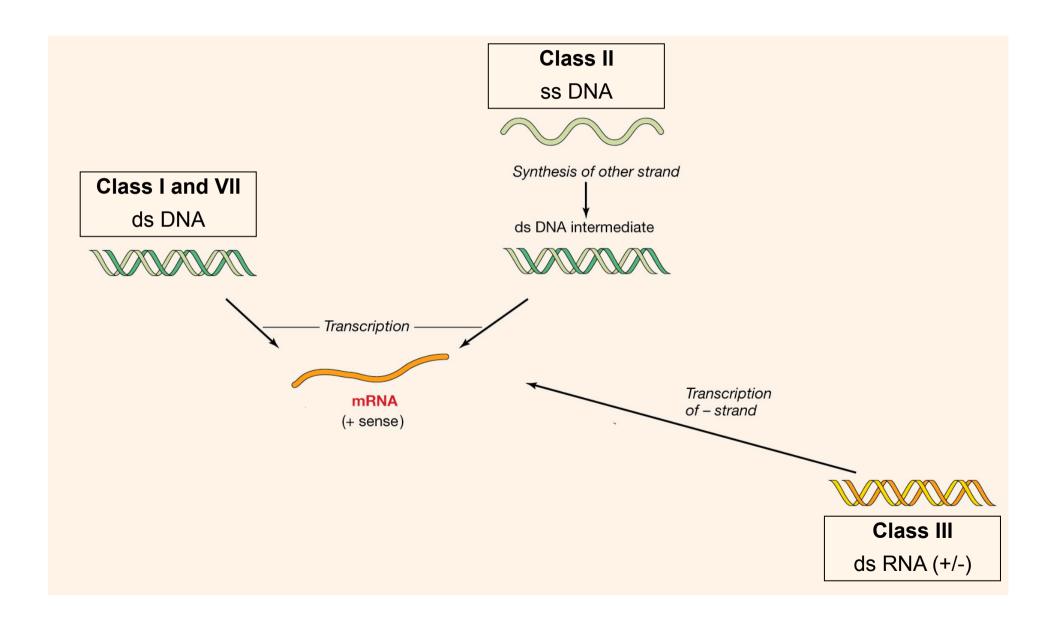


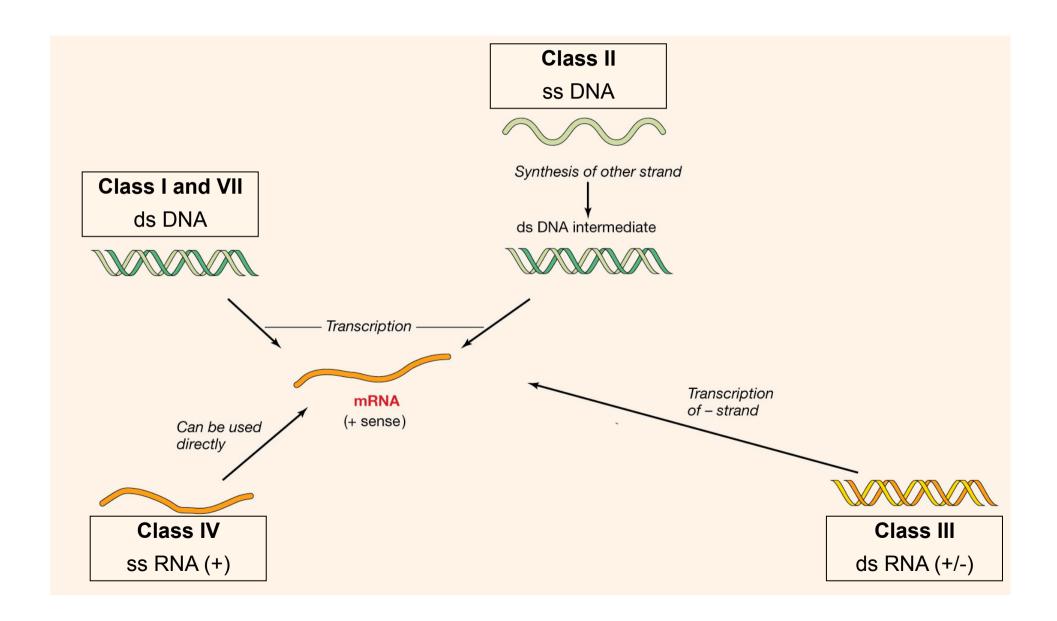


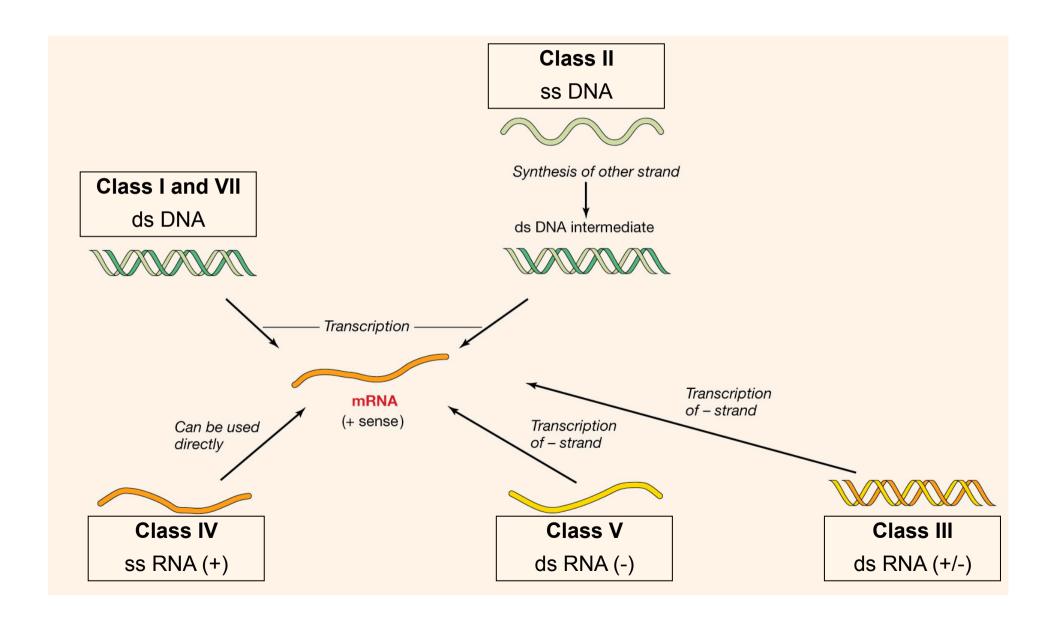
The diversity of viral genome replication strategies: adenovirus DNA replication

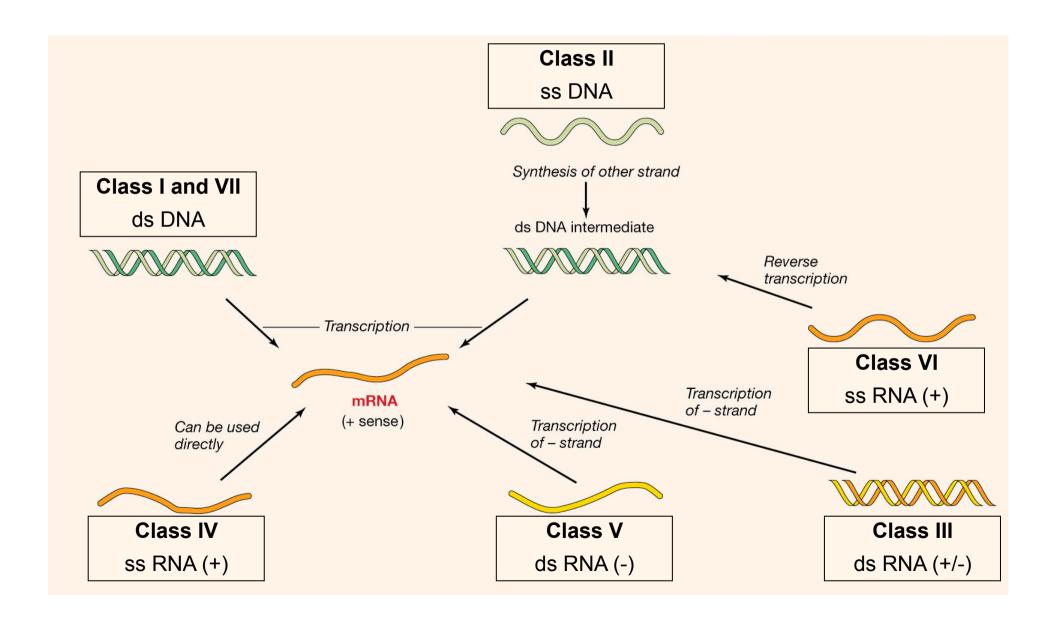


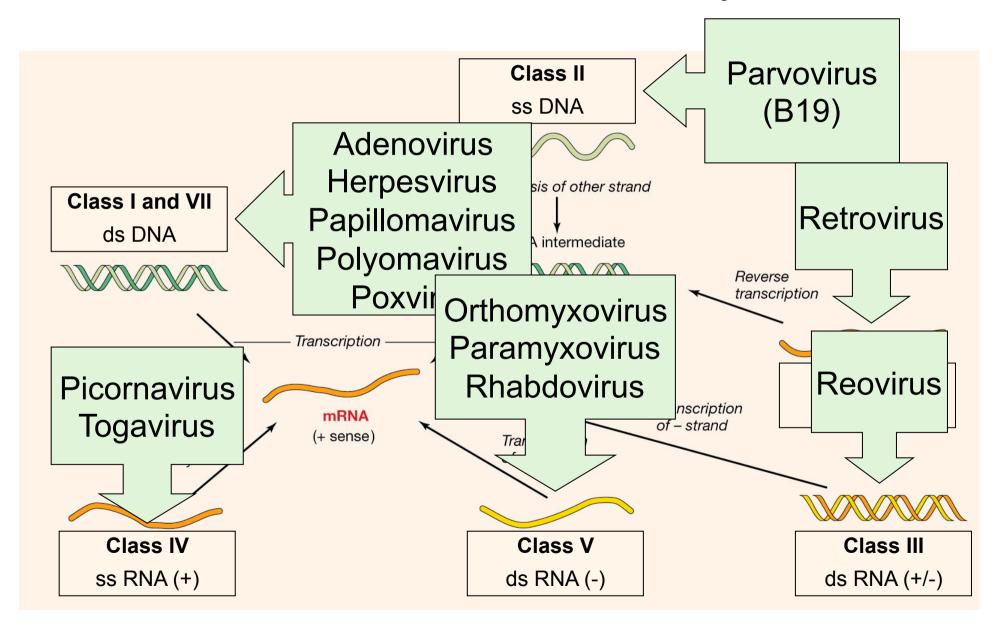






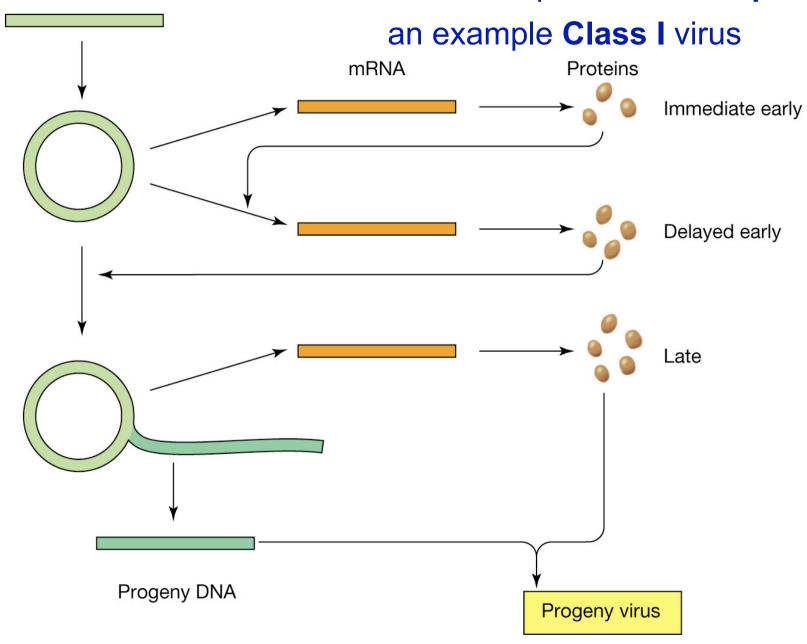




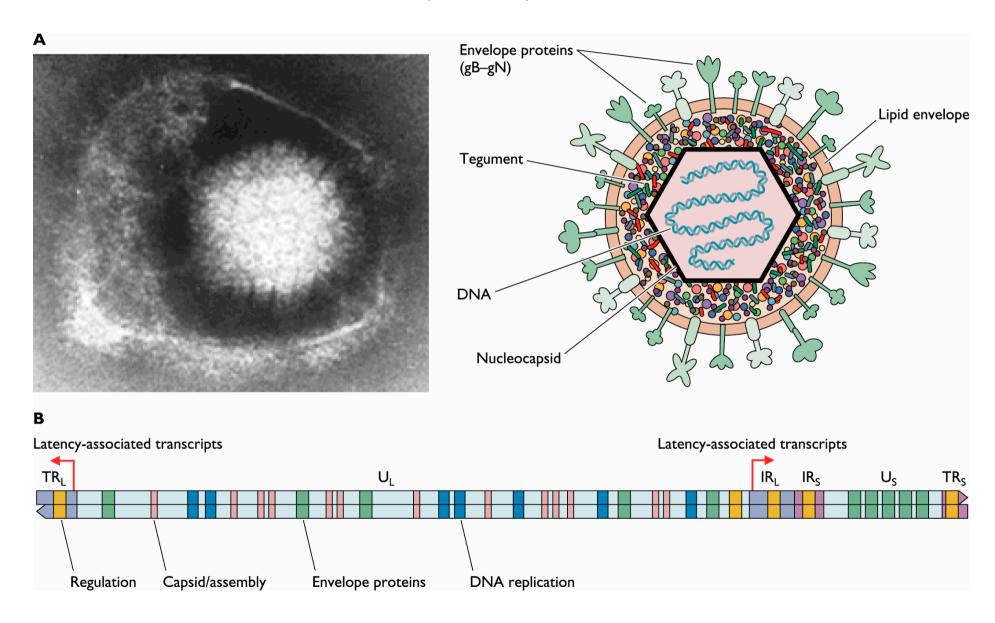


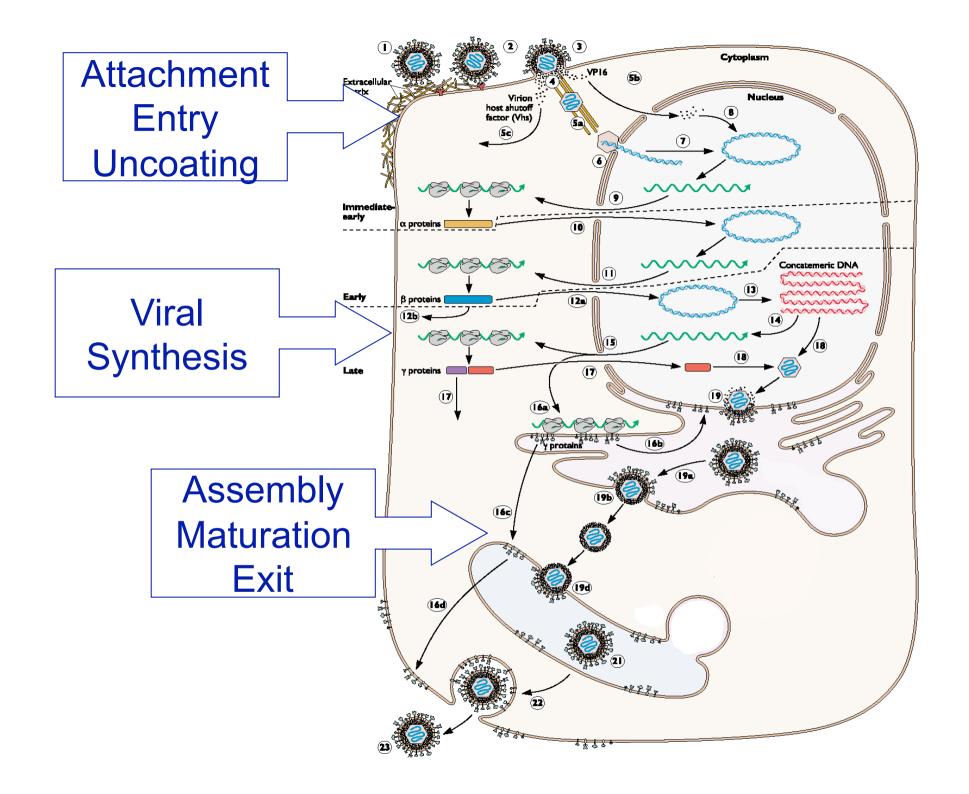
# Viral replication transcription, translation and genome replication of DNA viruses

#### Flow of events in multiplication of **Herpeviruses**:

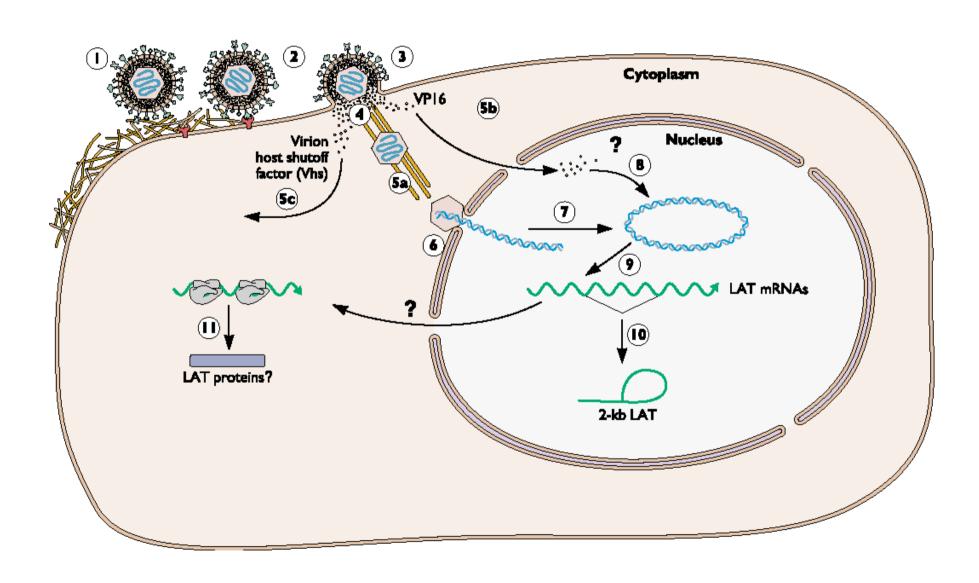


# Structure and genome organization of Herpes simplex 1 virus (HSV-1)

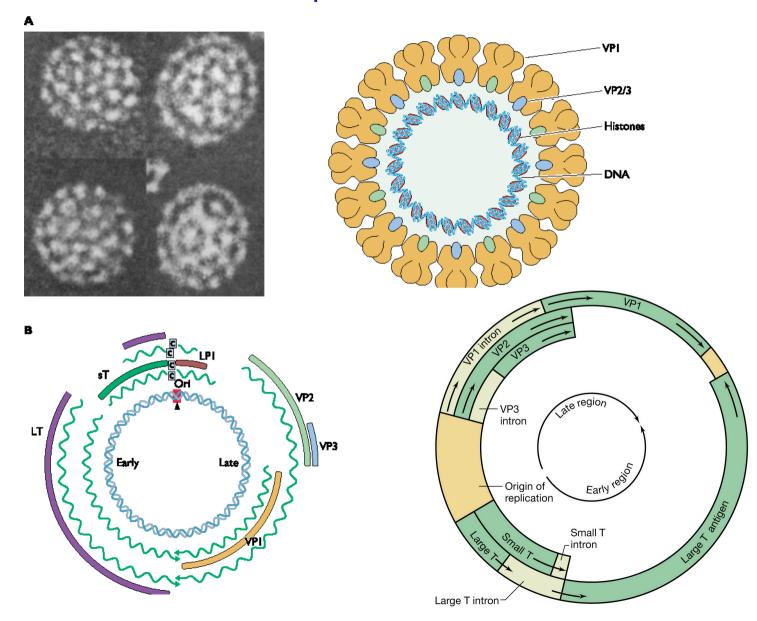


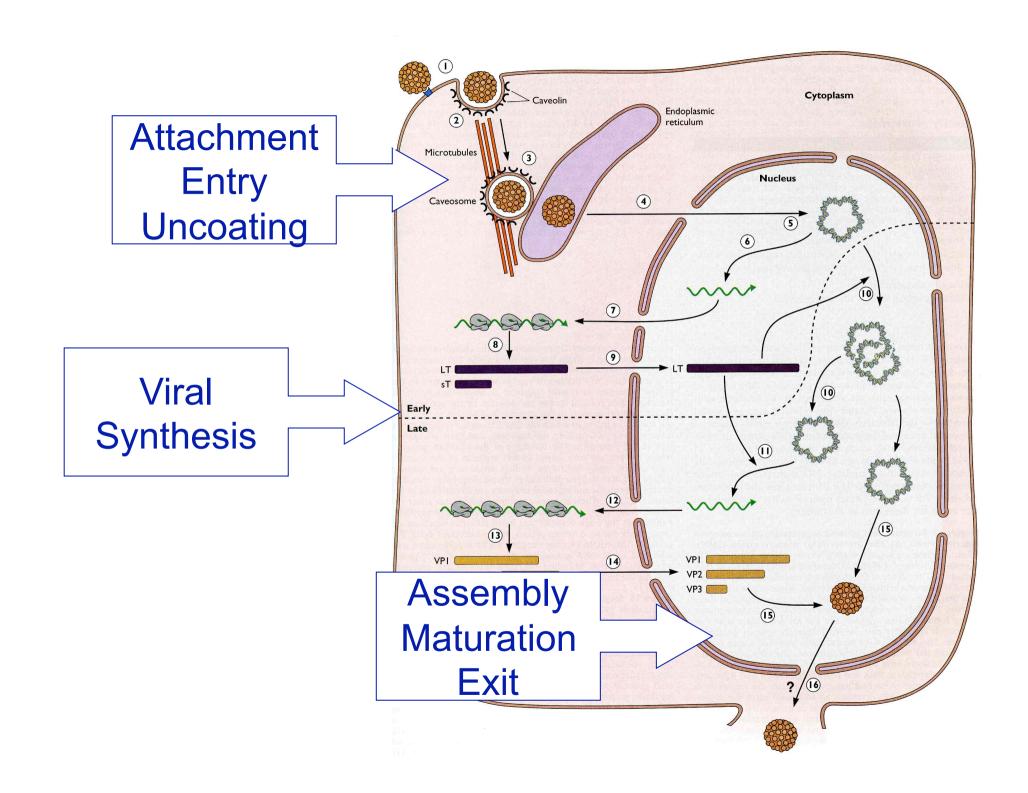


#### Herpes simplex virus latent infection in neurons

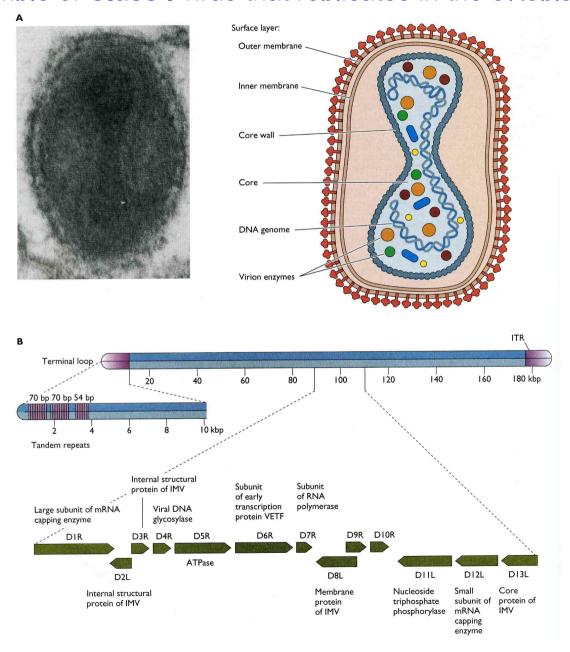


# Structure and genome organization of the **Polyomavirus**Simian Virus 40: an example of Class I tumoral virus



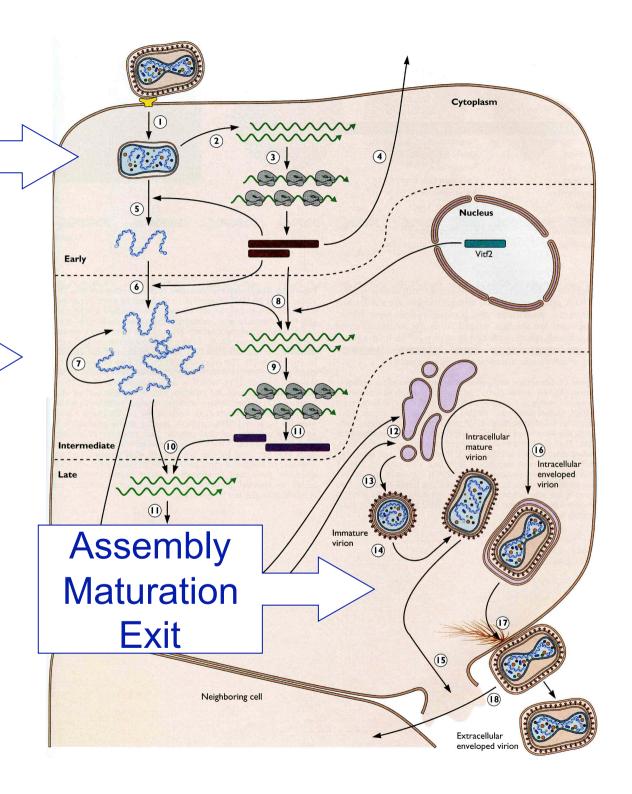


### Structure and genome organization of the **Poxvirus** Vaccinia virus: an example of **Class I** virus that replicates in the cvtoplasm

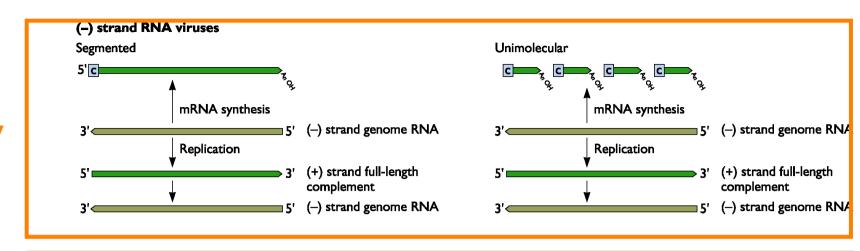


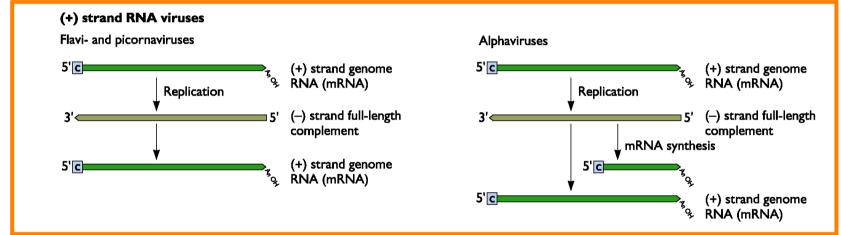
Attachment Entry Uncoating

Viral Synthesis



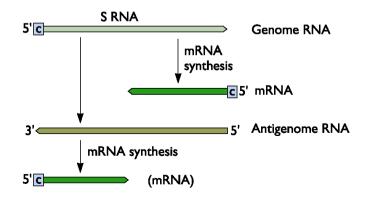
# Viral replication transcription, translation and genome replication of RNA viruses

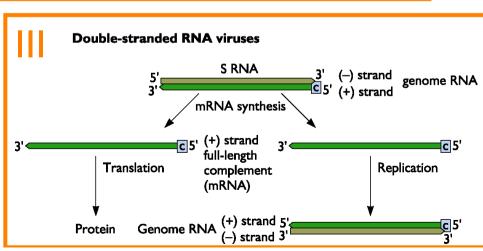




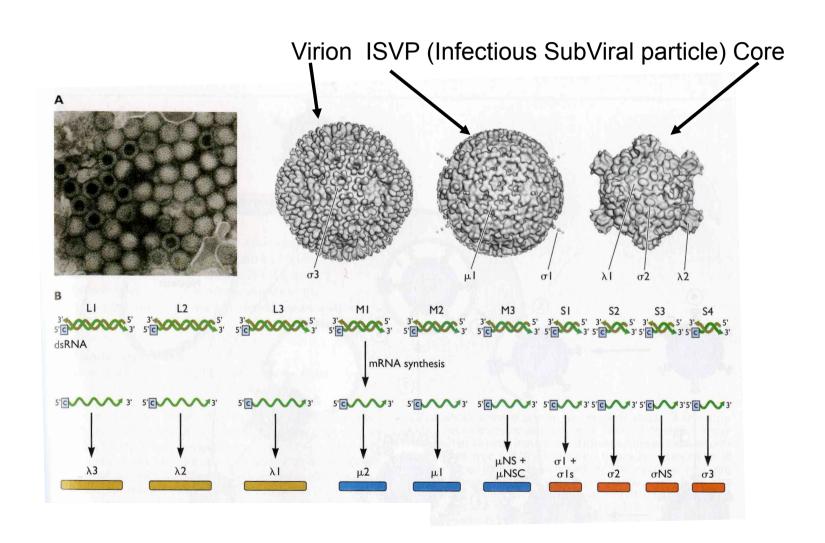


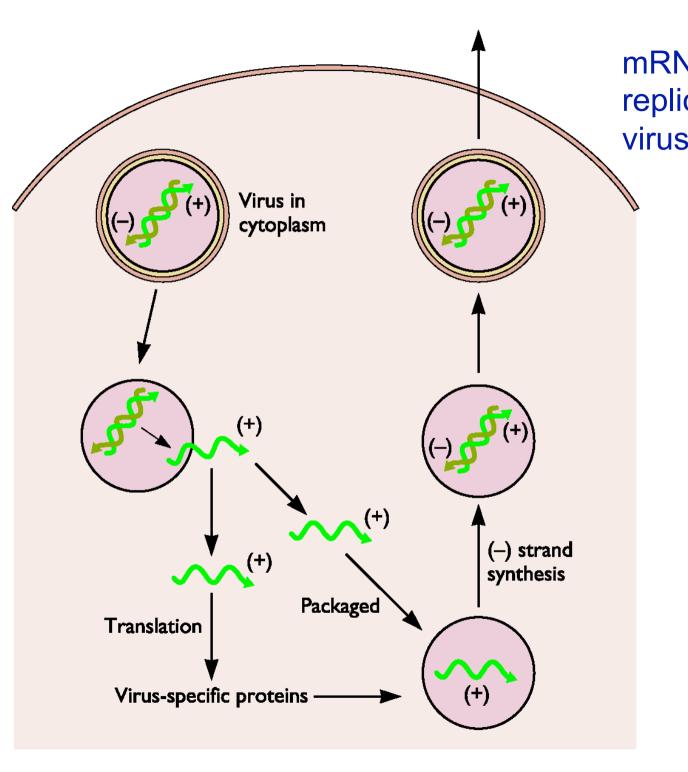
IV





#### Structure and genomic organization of a Reovirus

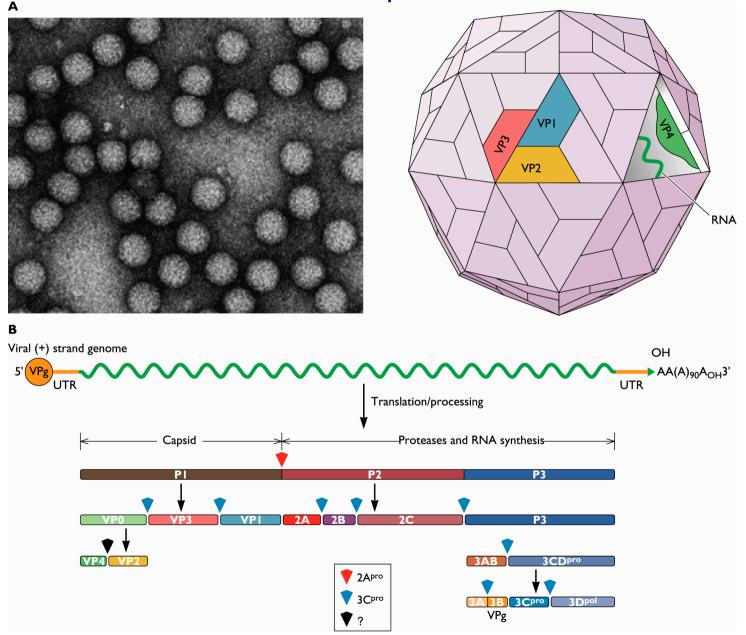




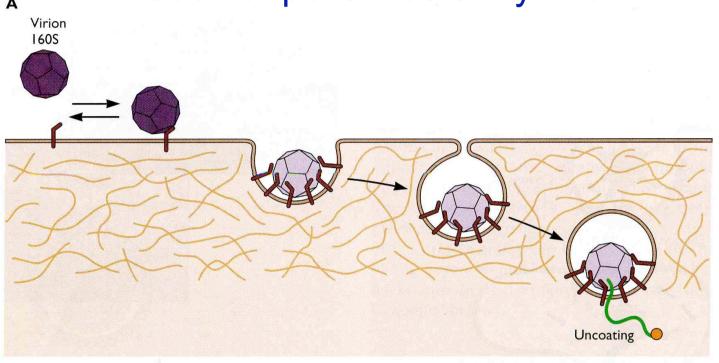
mRNA synthesis and replication of **Class III** virus: the **Reoviruses** 

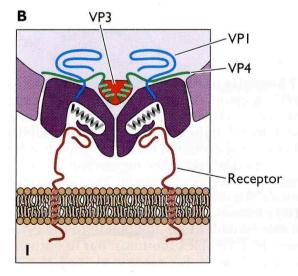
## Reproductive cycle of a Reovirus Nucleus ISVP Core Virion Subviral particles Cytoplasm

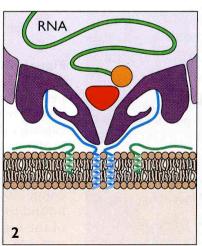
Structure and genome organization of the **Picornavirus**Poliovirus: an example of **Class IV** virus

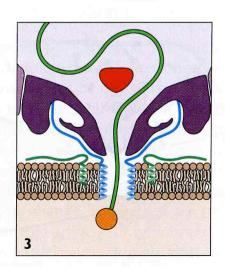


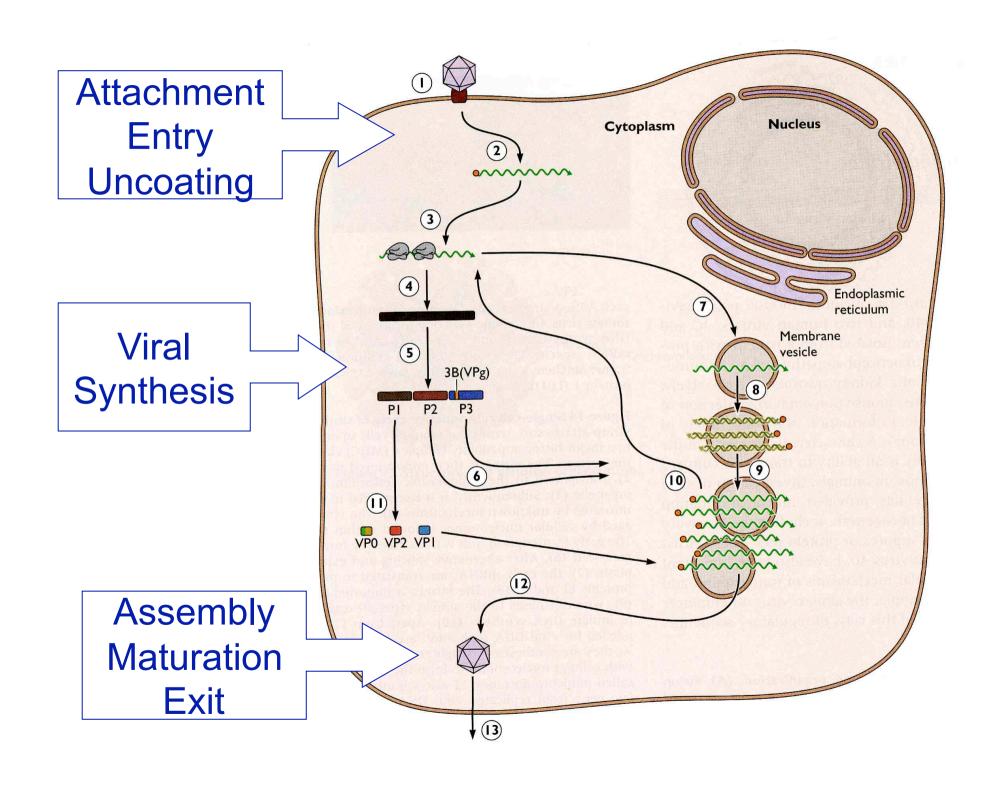
### Model for poliovirus entry into



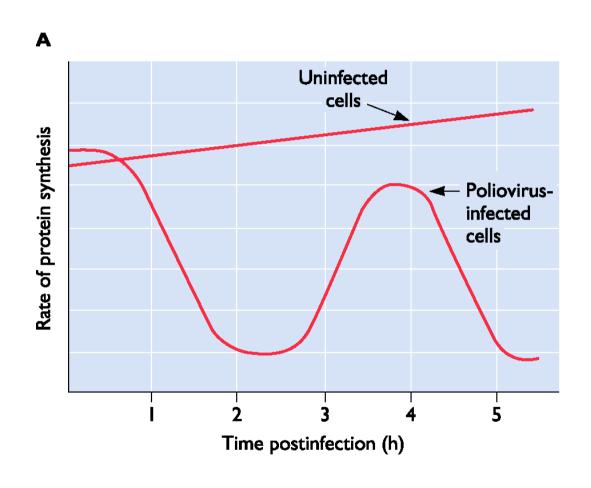


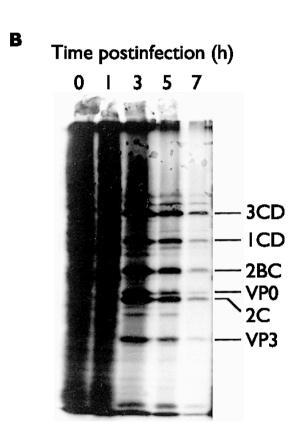




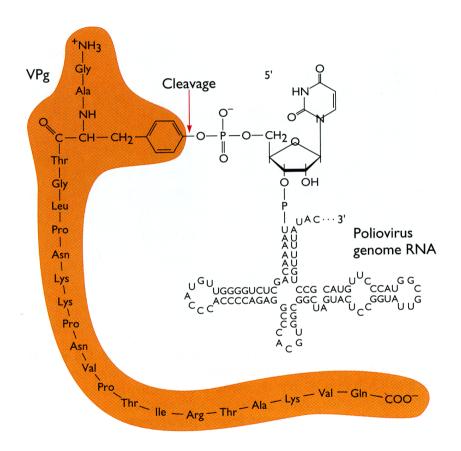


### Inhibition of translation in poliovirus-infected cells

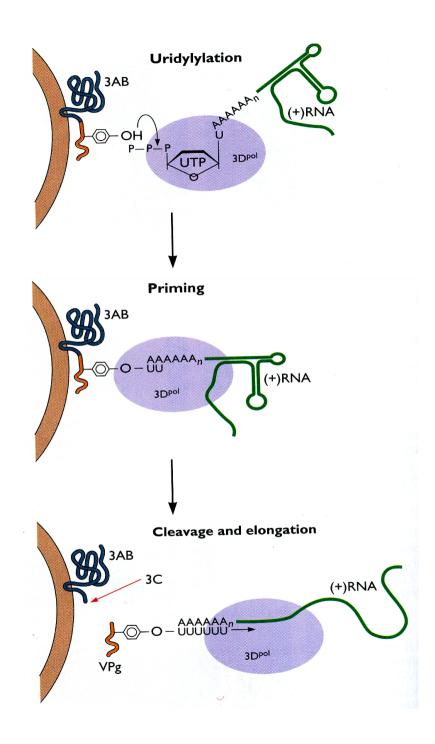




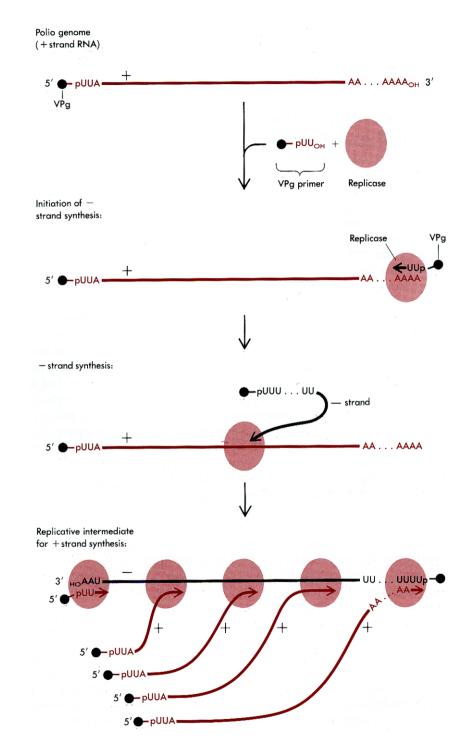
### Poliovirus RNA replication



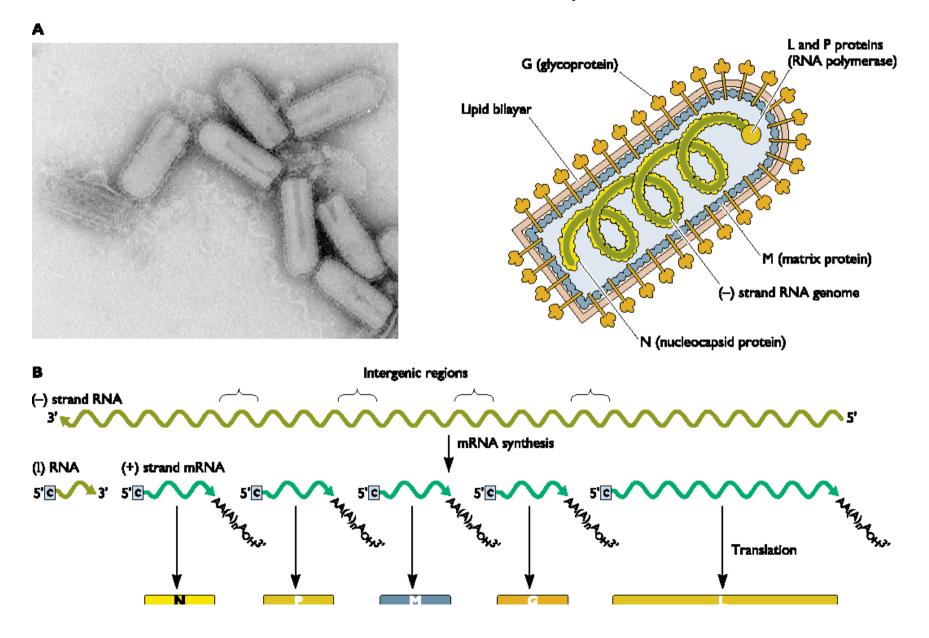
Linkage of VPg to polioviral genomic RNA



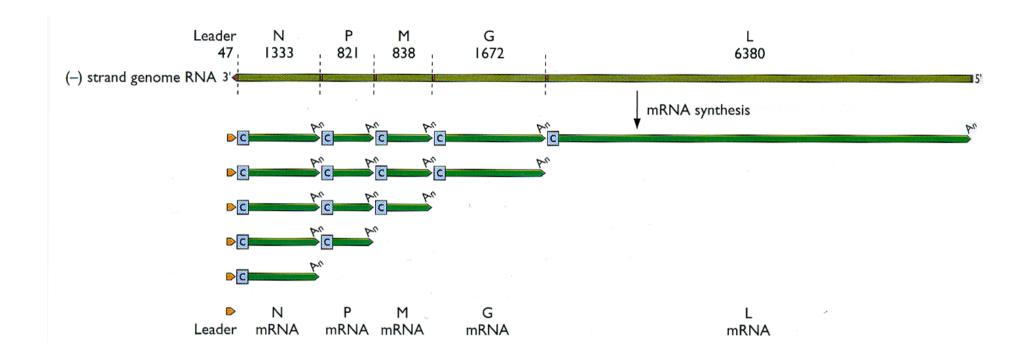
### Poliovirus RNA replication

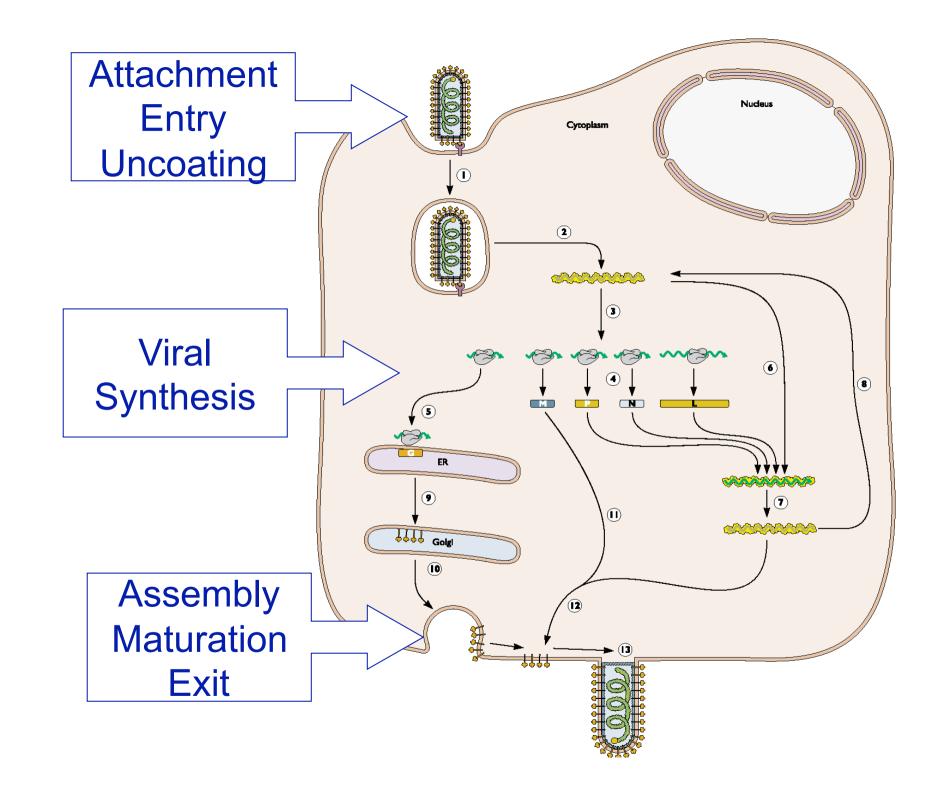


### Structure and genome organization of the **Rhabdovirus** Vesicular Stomatitis Virus: an example of Class V virus

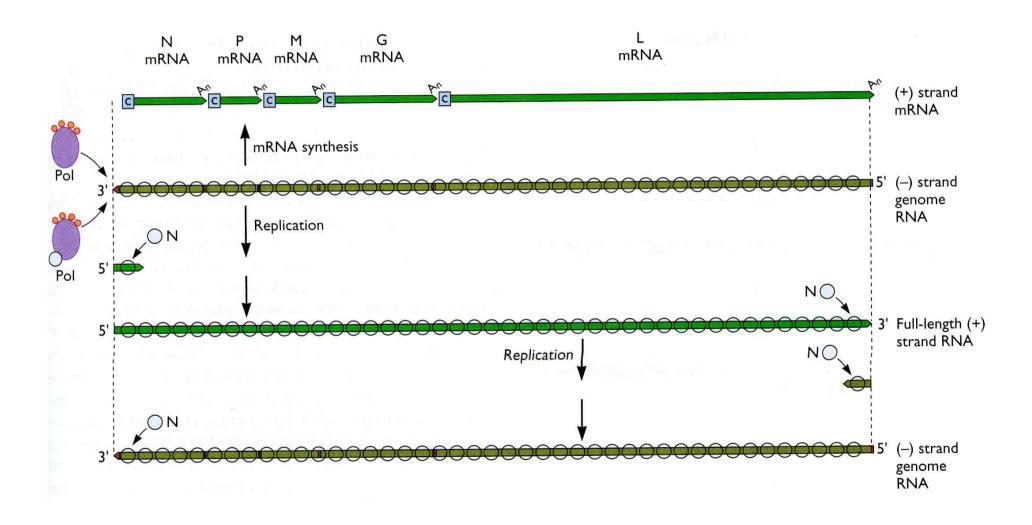


### Vesicular stomatitis virus mRNA map



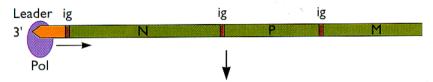


### Transcription and replication of the VSV genome

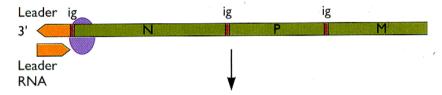


#### VSV mRNA synthesis and function of RNA pol at an intergenic region

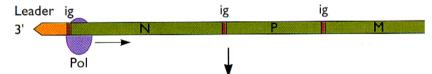
#### Initiation at 3' end of VSV genome RNA



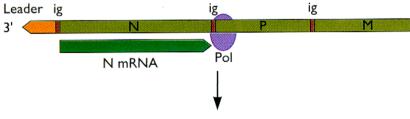
#### Synthesize leader and terminate at intergenic region (ig)



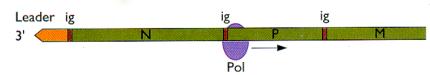
#### Reinitiate at 3' end of N gene

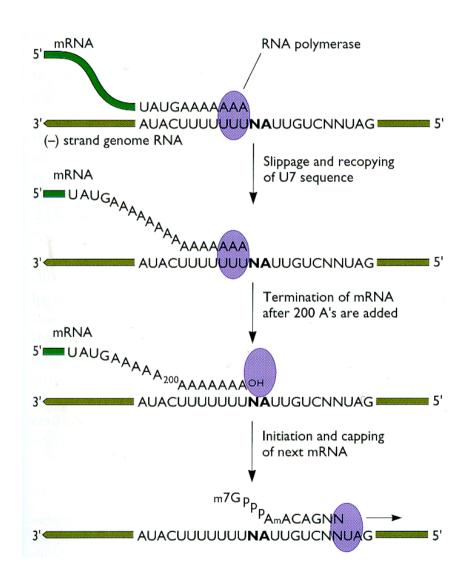


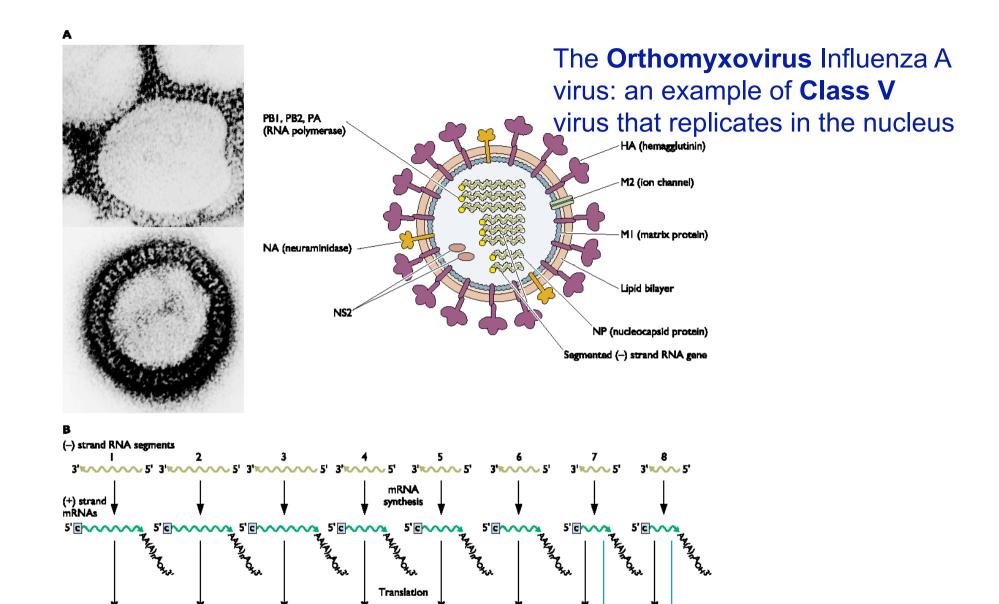
#### Synthesize N gene and terminate at intergenic region (ig)



#### Reinitiate at 3' end of P gene





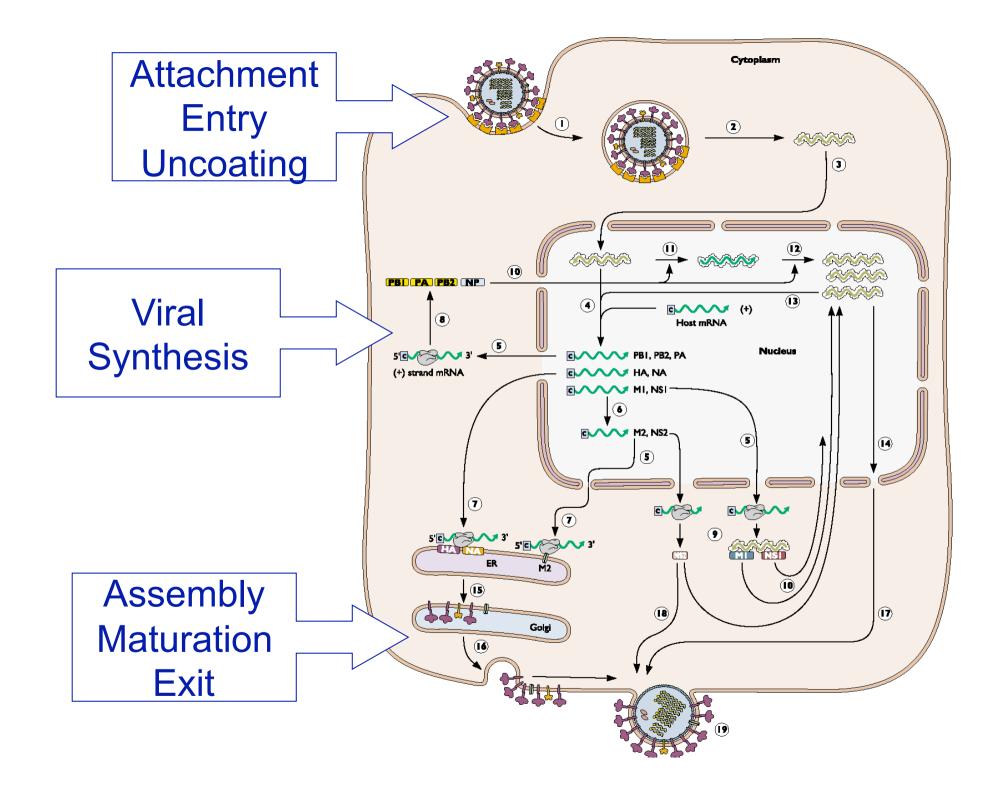


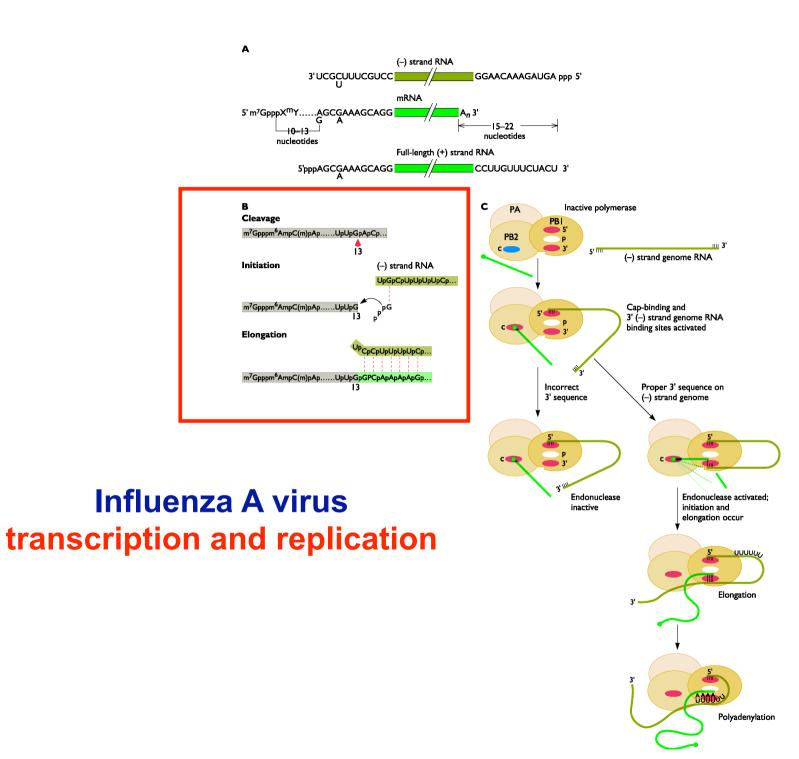
MI

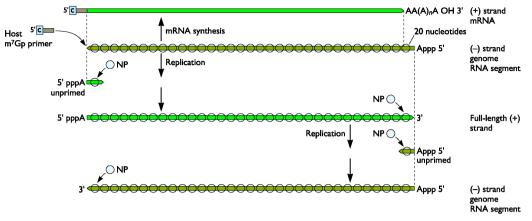
Translation,

NSI

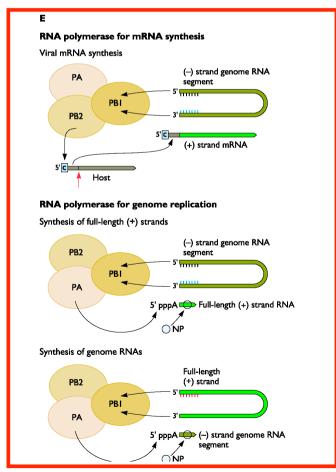
Splicing

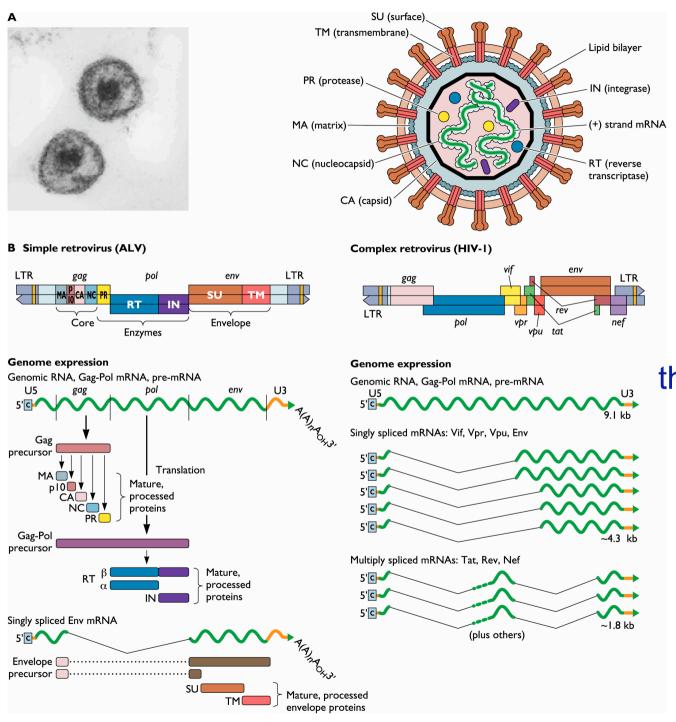




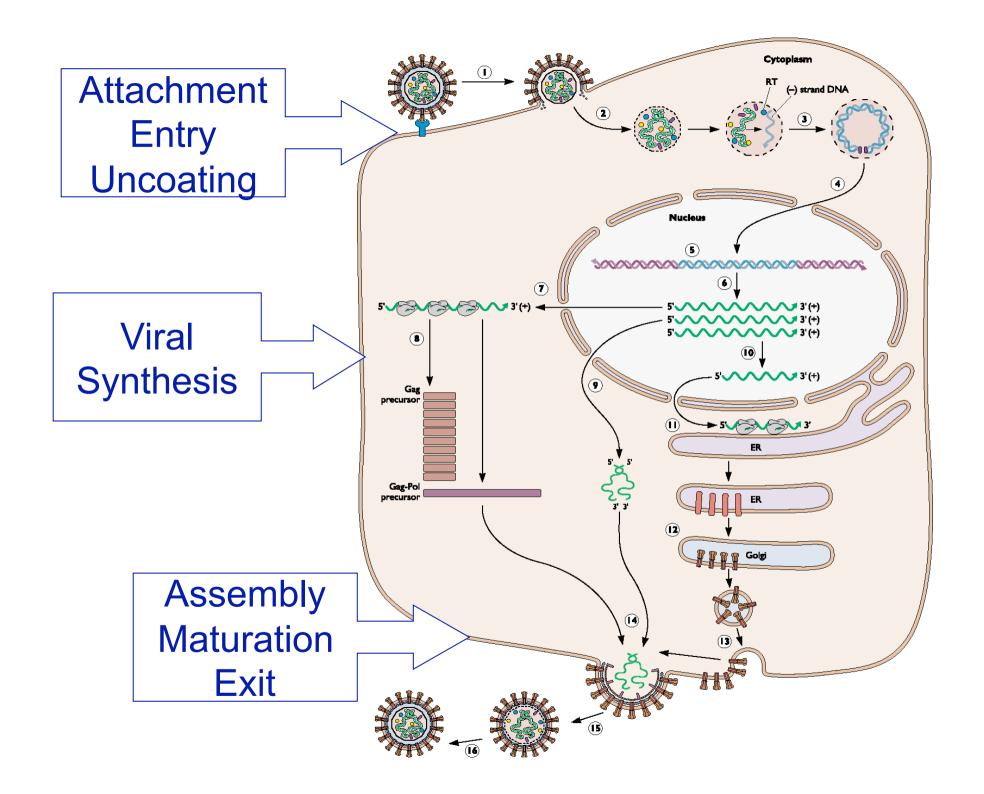


### Influenza A virus transcription and replication

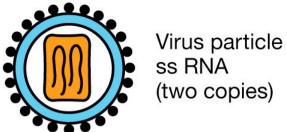


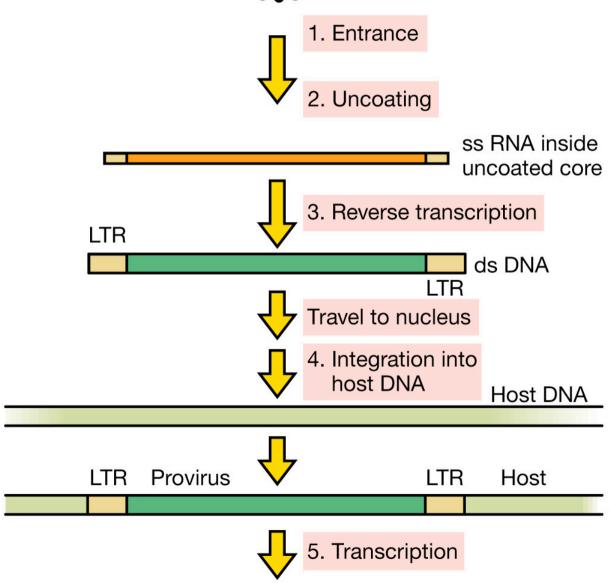


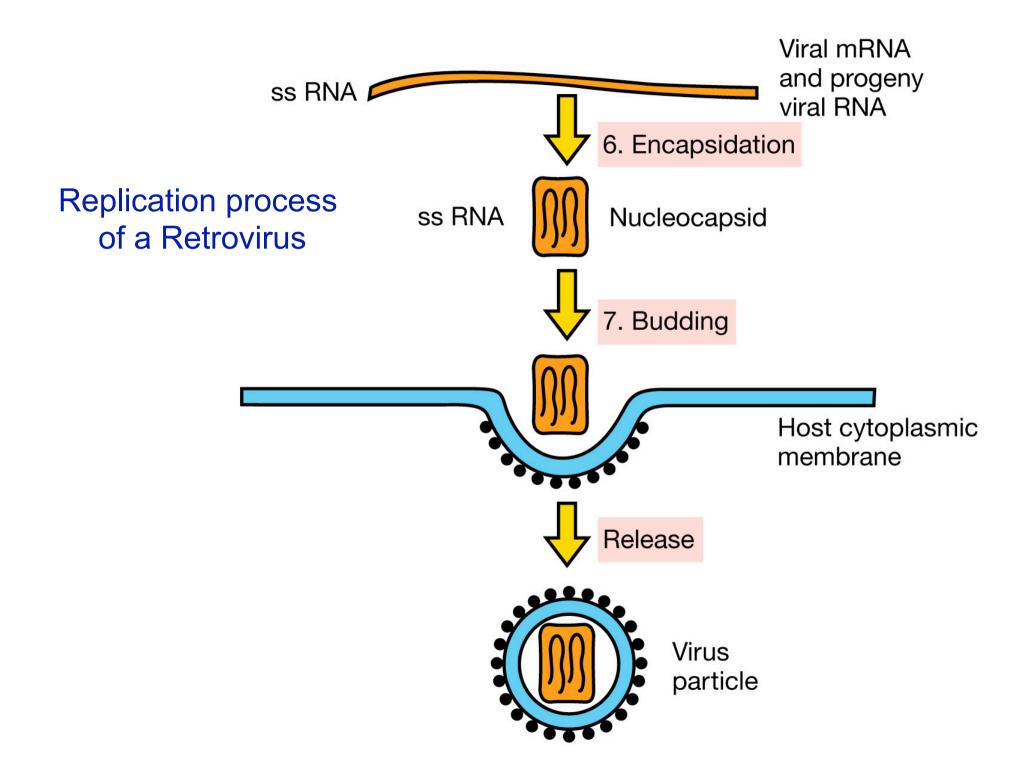
### Class VI virus: the Retroviruses



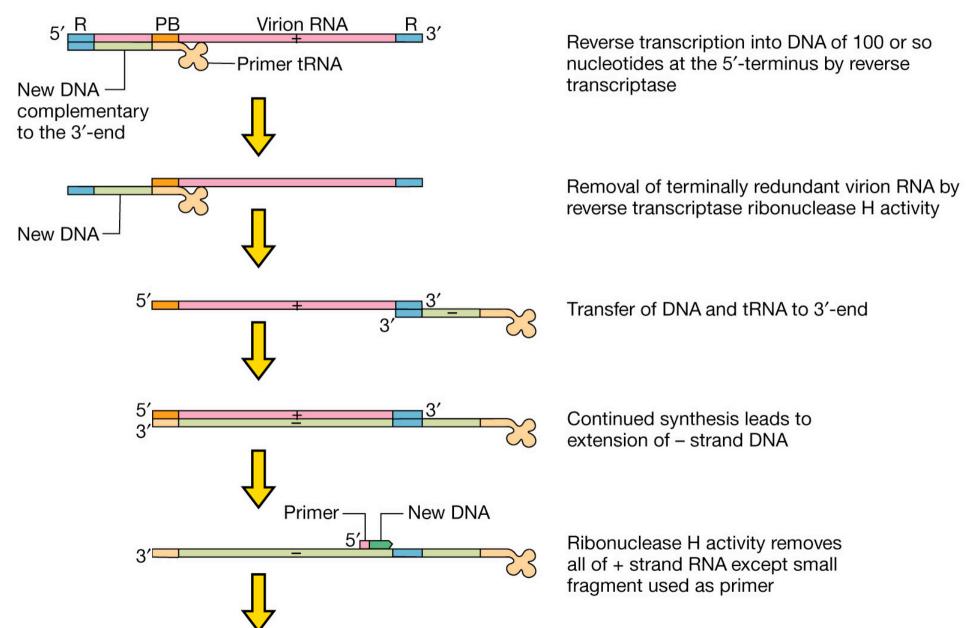
### Replication process of a Retrovirus

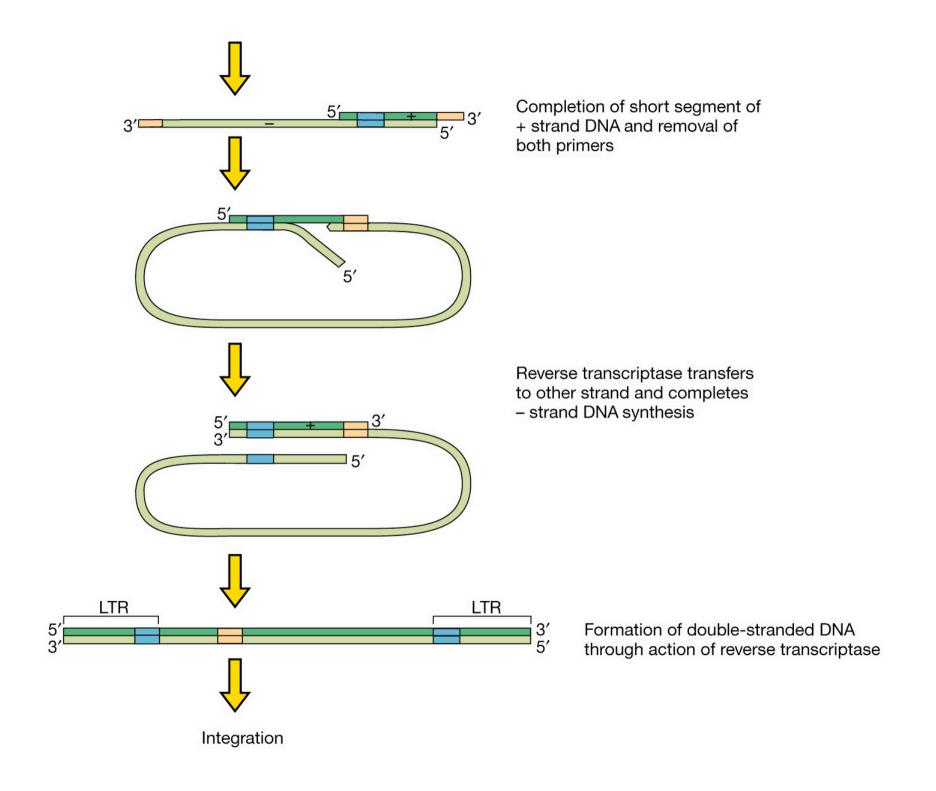




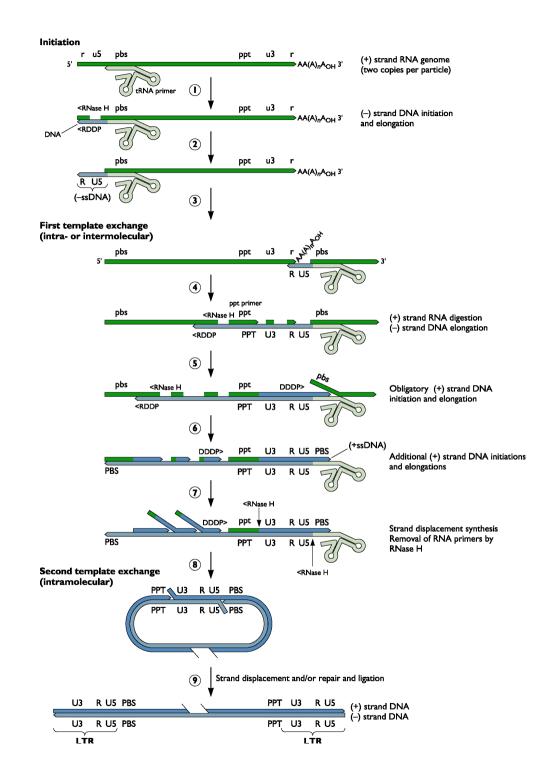


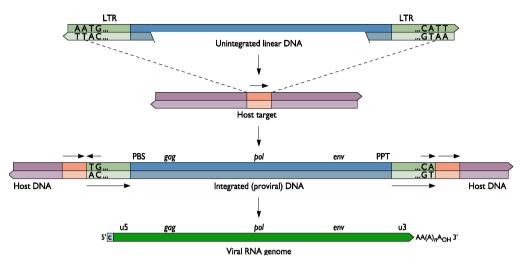
### Overall steps in the formation of double-stranded DNA from Retrovirus single-stranded RNA



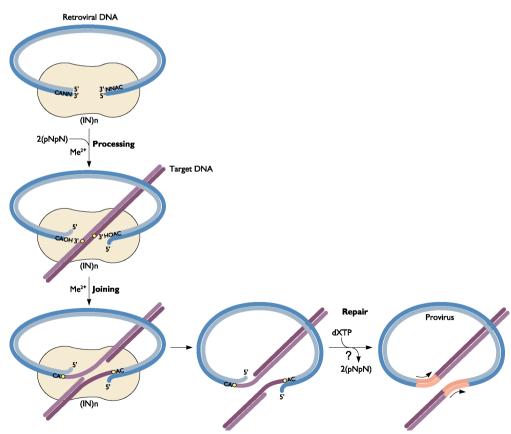


# Reverse transcription process

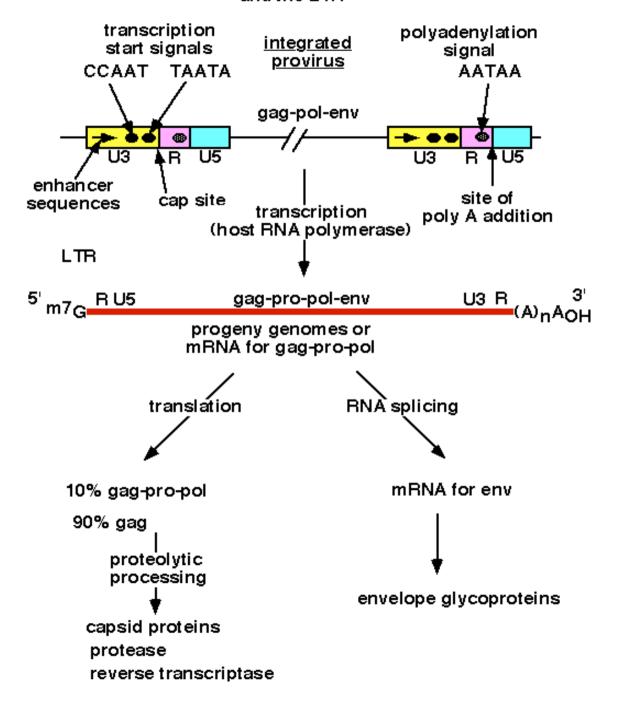


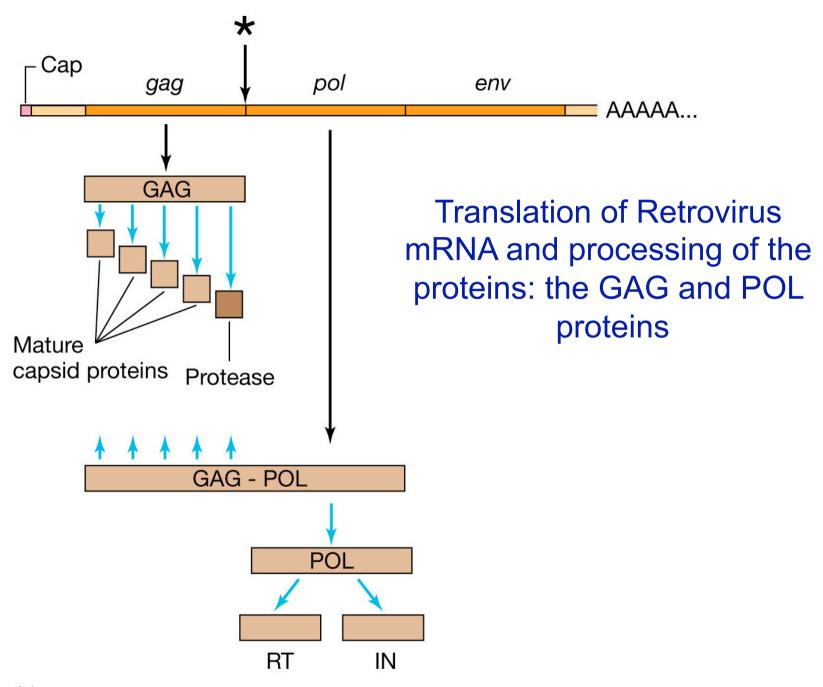


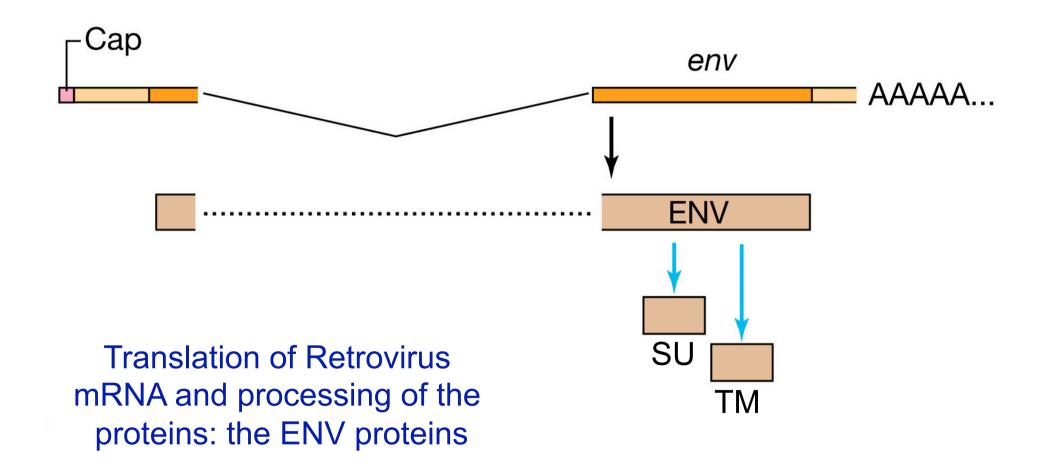
### Retroviral DNA integration



#### Retrovirus Gene Expression and the LTR





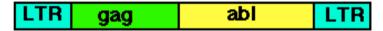


### Most Transforming Retrovirus are Defective and Cannot Replicate without Helper Virus

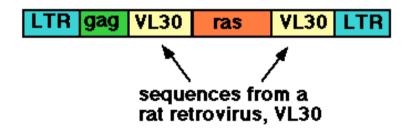
Rous sarcoma virus (a non-defective, transforming avian virus)



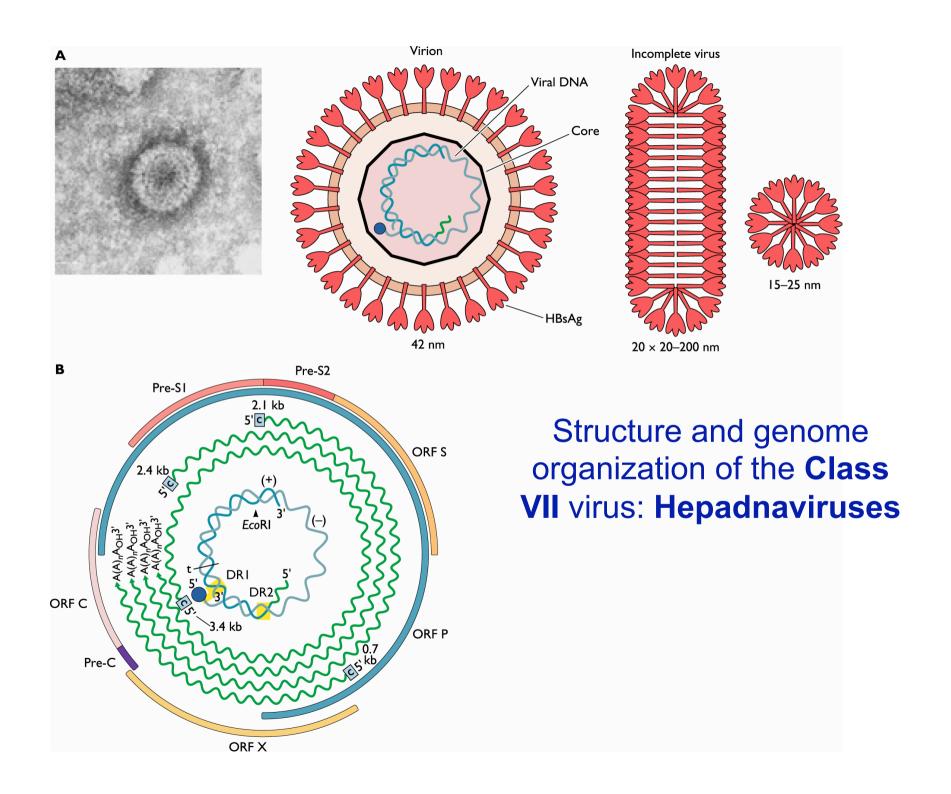
Abelson murine Leukemia virus (defective)

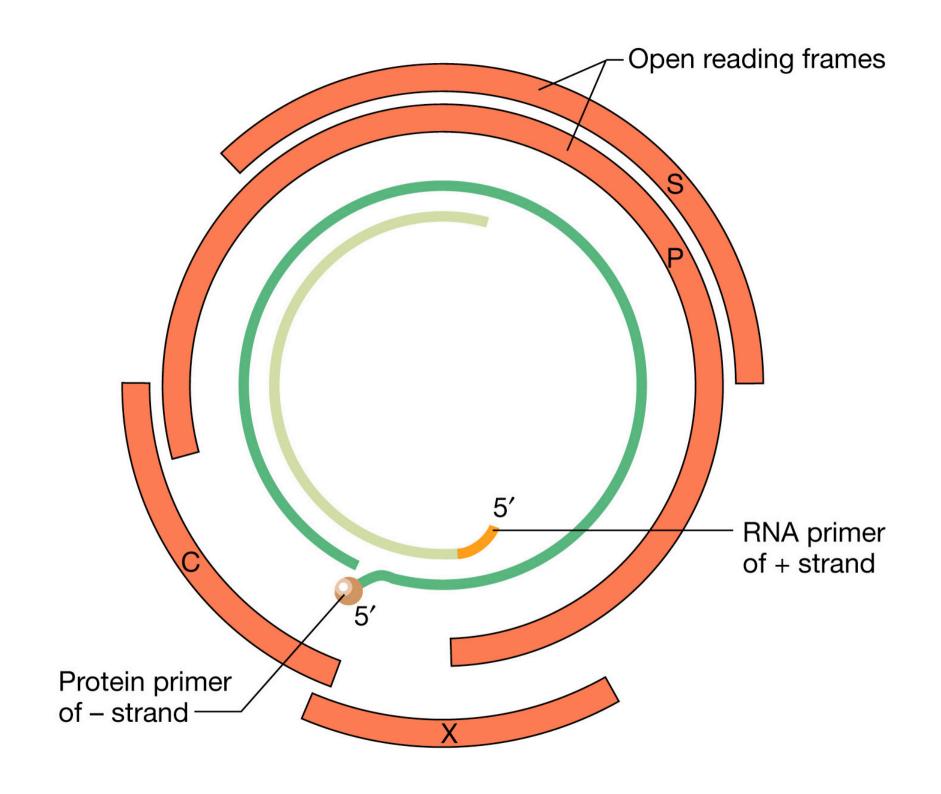


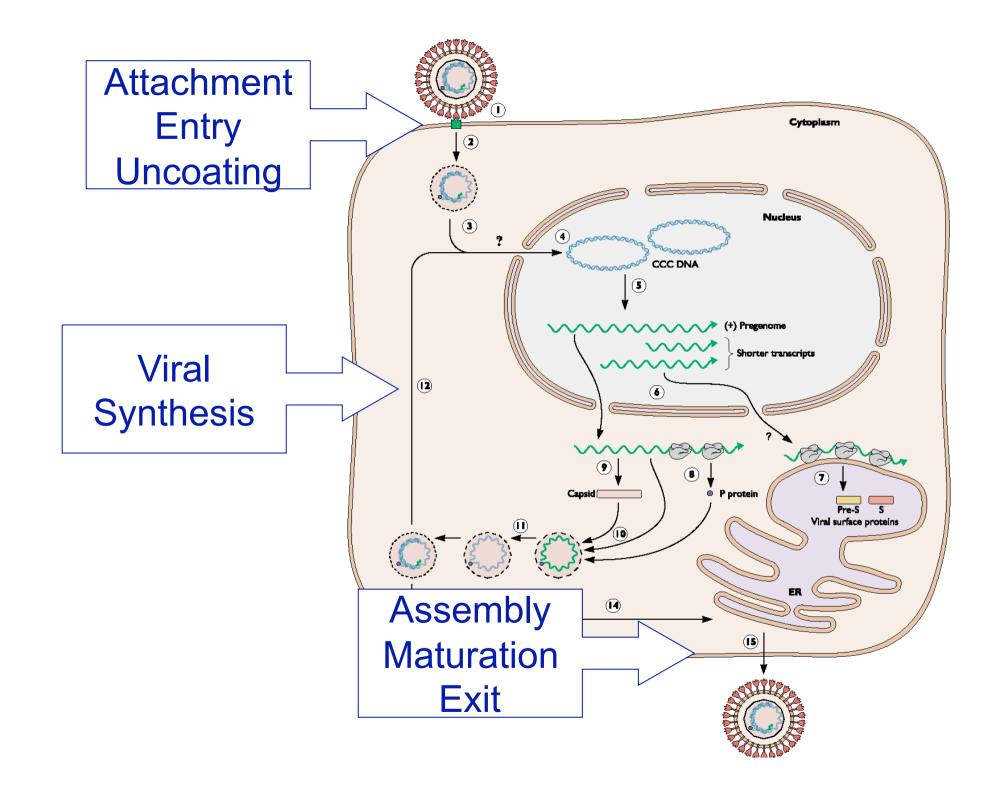
Harvey sarcoma virus (a defective murine virus)



src, ab/and rasare v-onc sequences which were picked up (probably as processed trancripts) from c-onc sequences in the host.



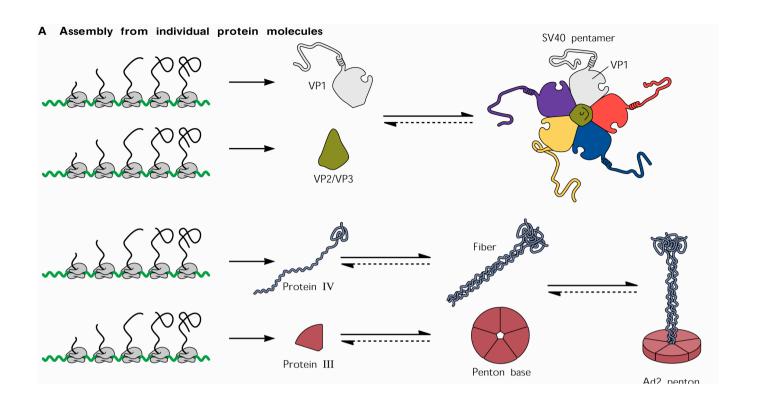




# Viral replication assembly, exit and maturation of progeny virions

# Hypothetical pathway of virion assembly and release

Formation of individual structural units of the protein shell from one or several viral proteins

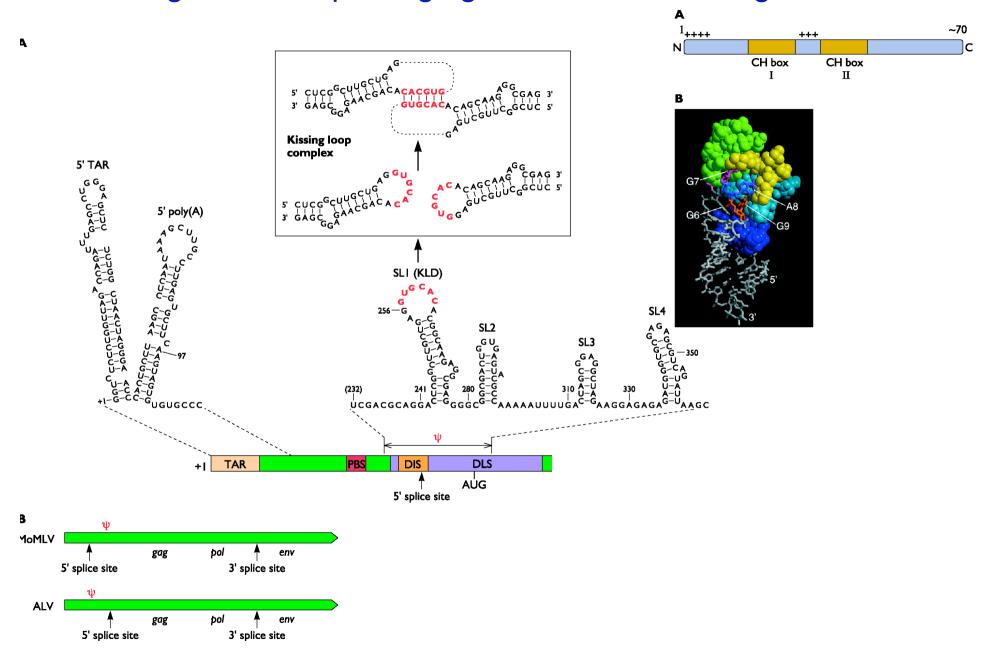


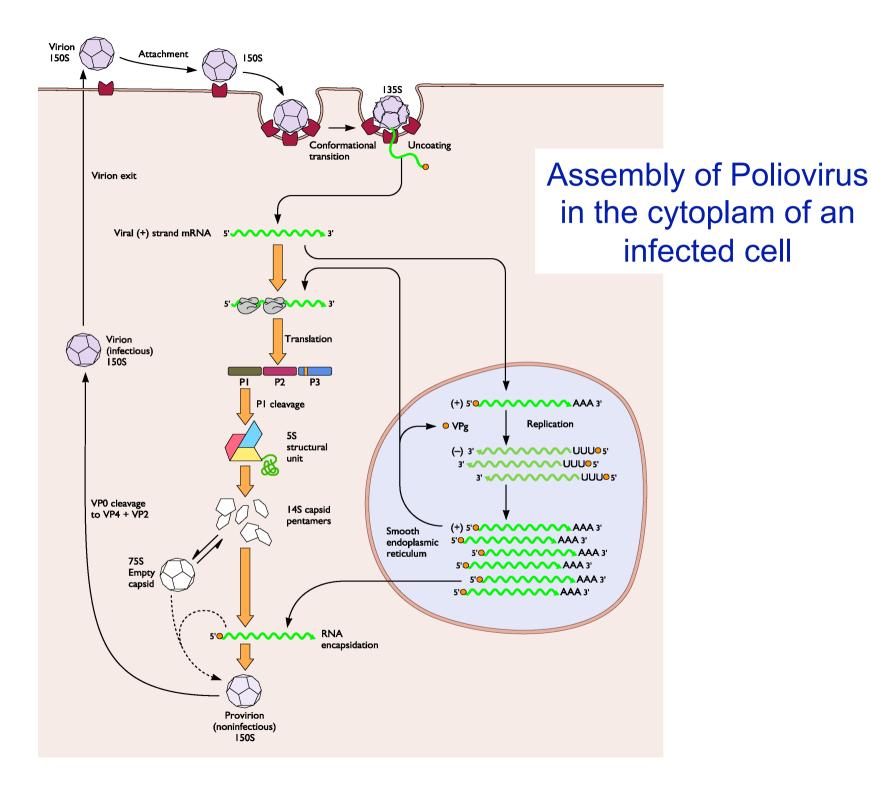
# DR2 pac2 DRI pacl DR2 ep I ер 2

### Recognition and packaging of the nucleid acid genome

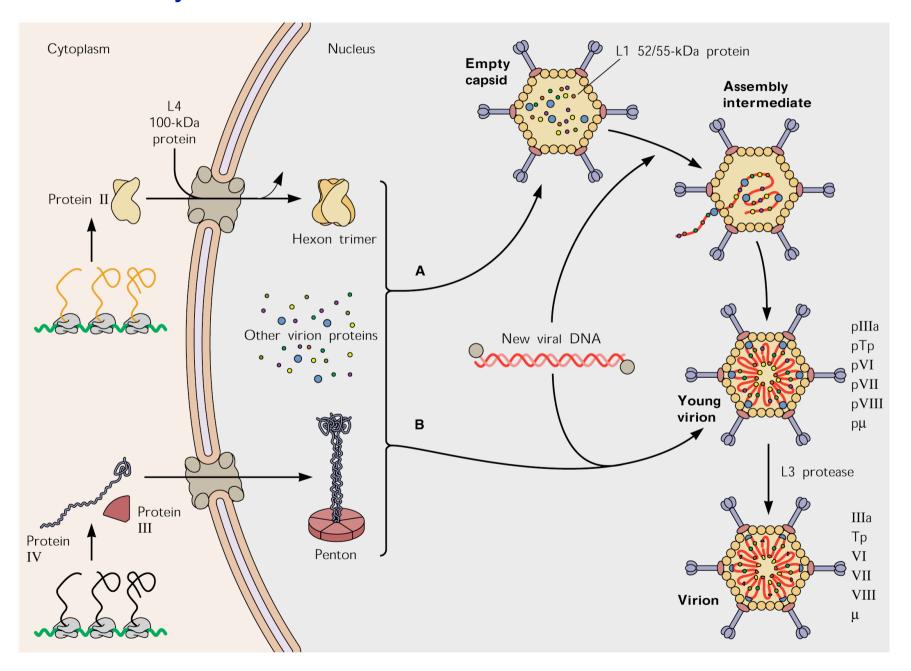


#### Recognition and packaging of the nucleid acid genome

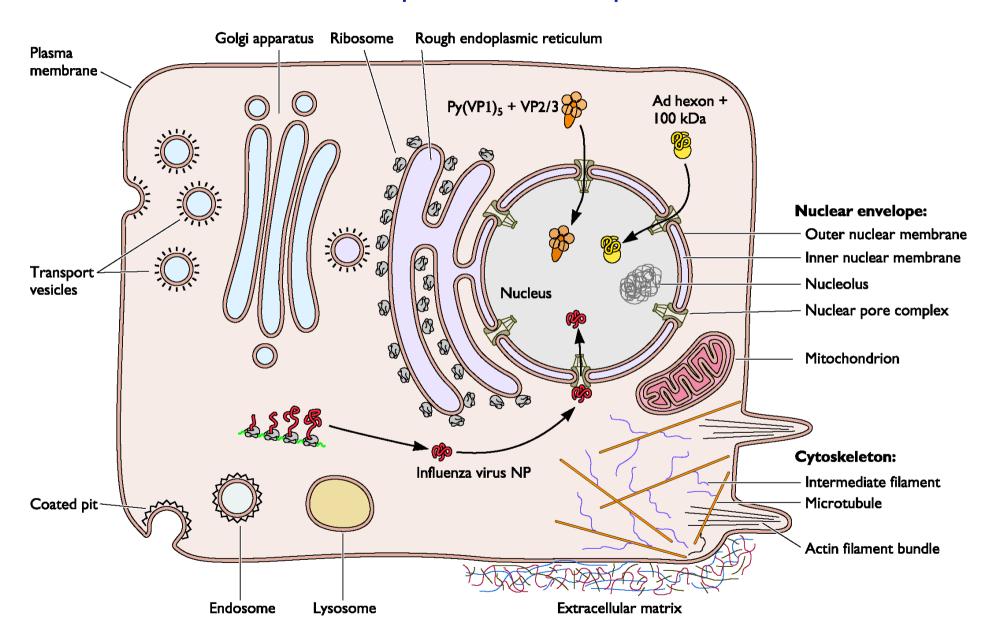




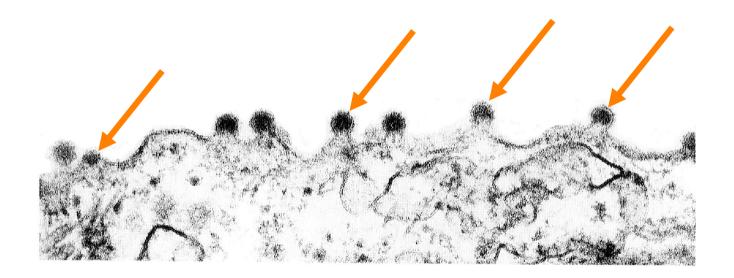
#### Assembly of Adenovirus in the nucleus of an infected cell

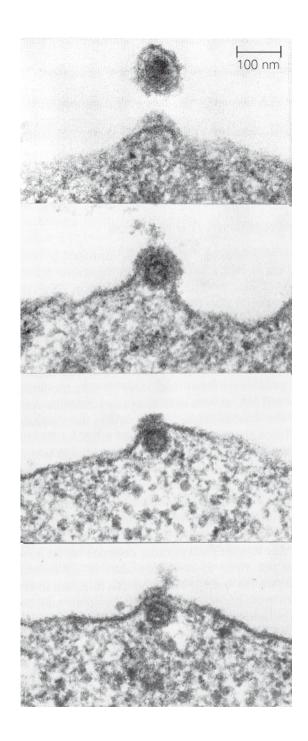


#### Localization of viral proteins to the plasma membrane

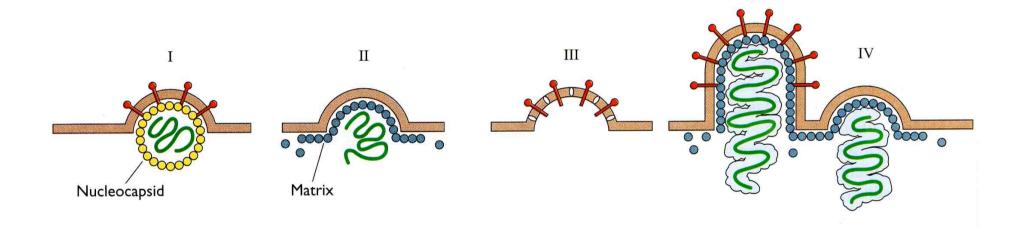


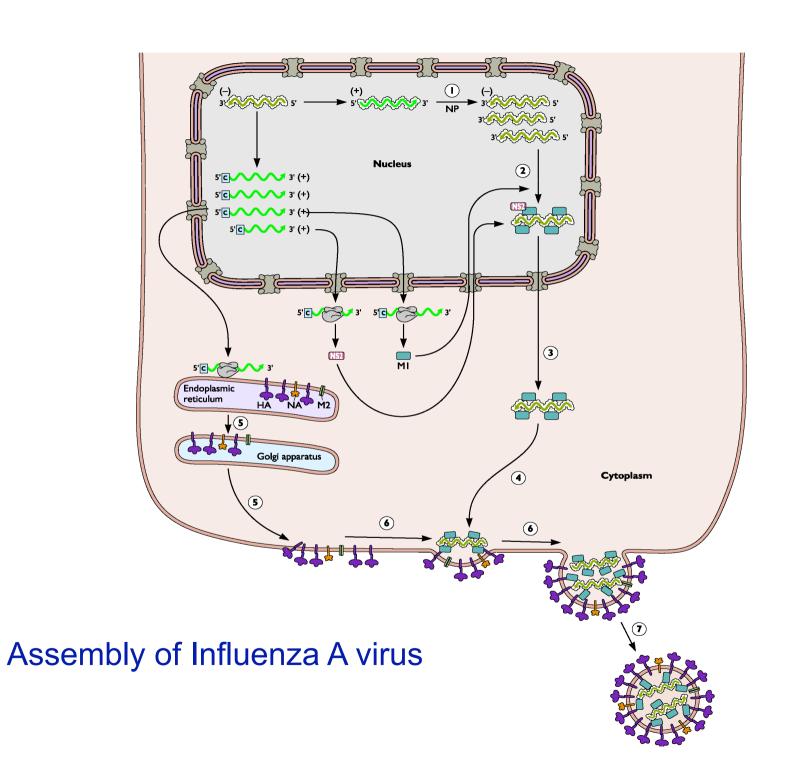
#### Mechanism of budding of enveloped viruses



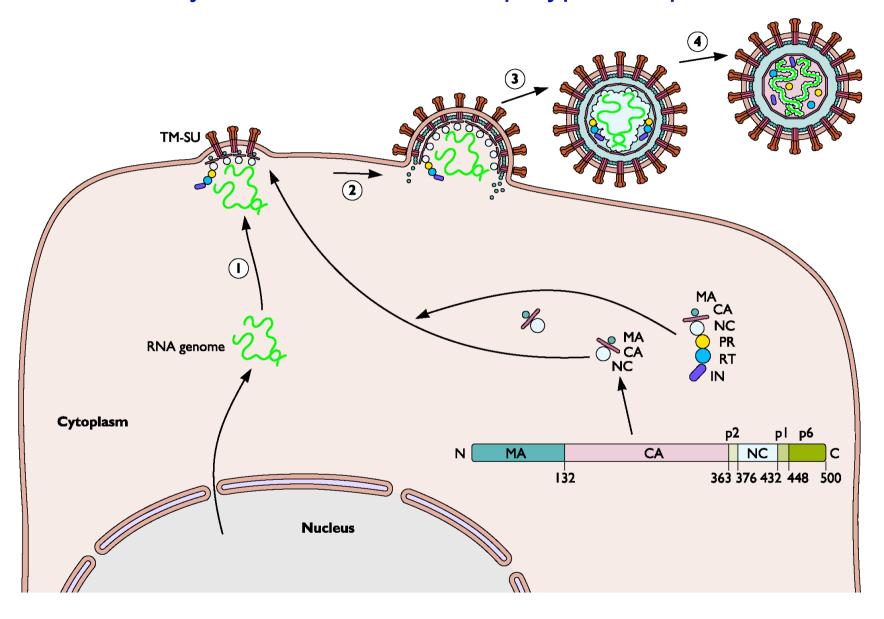


# Interaction of viral proteins responsible for budding at the plasma membrane

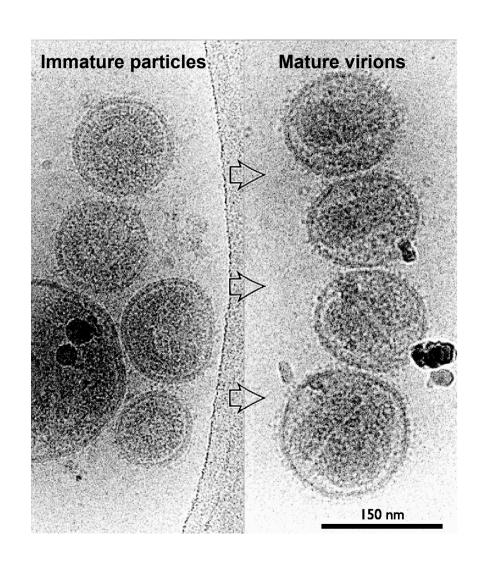


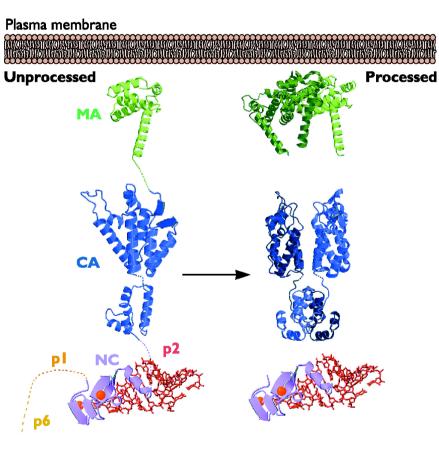


#### Assembly of a Retrovirus from polyprotein precursors

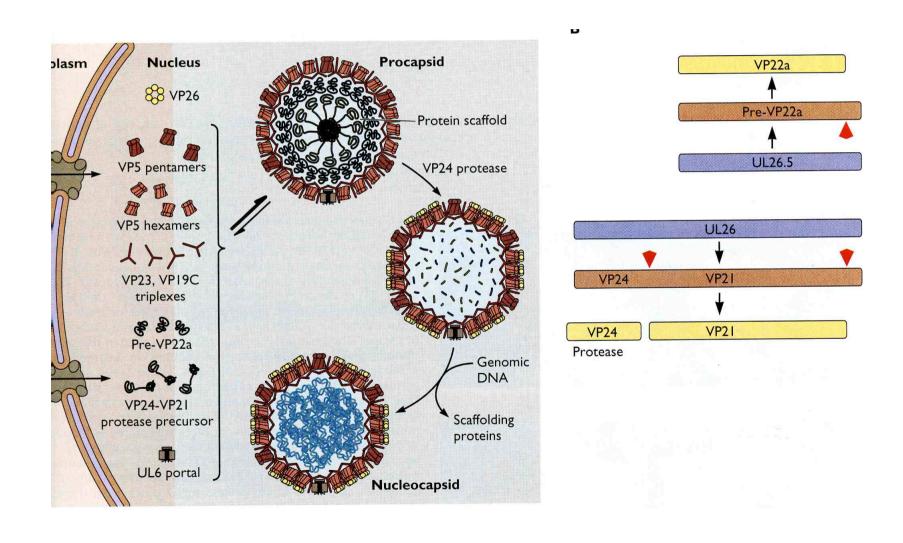


# Morphological rearrangement of the HIV-1 particle upon proteolytic processing of the Gag polyprotein

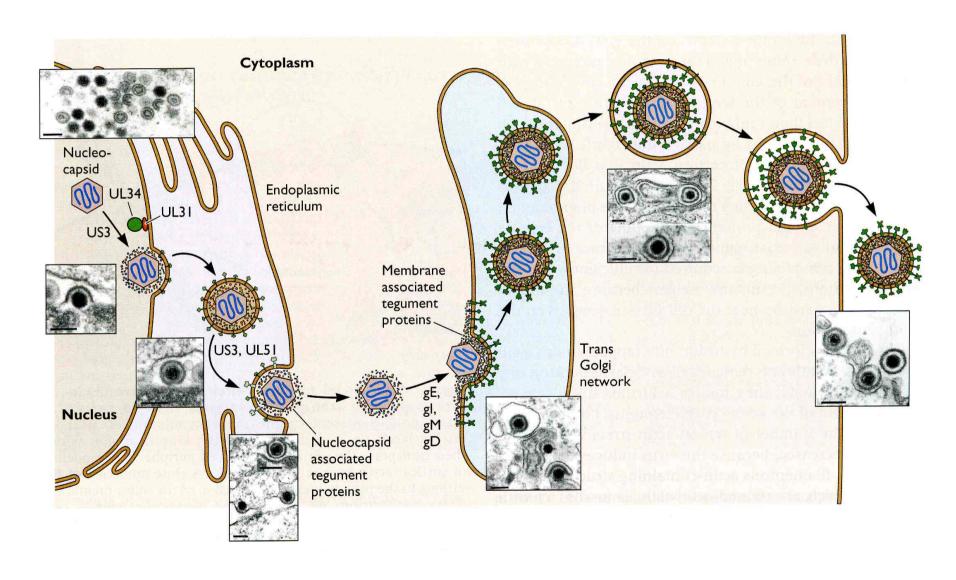


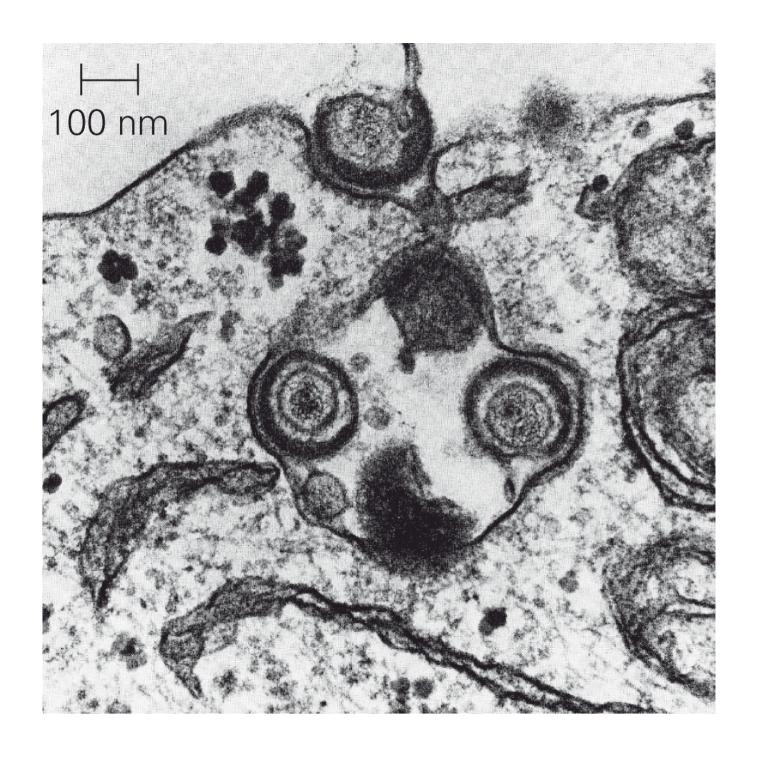


## Assembly of HSV-1 nucleocapsids and the pathway proposed for the virus exit from an infected cell

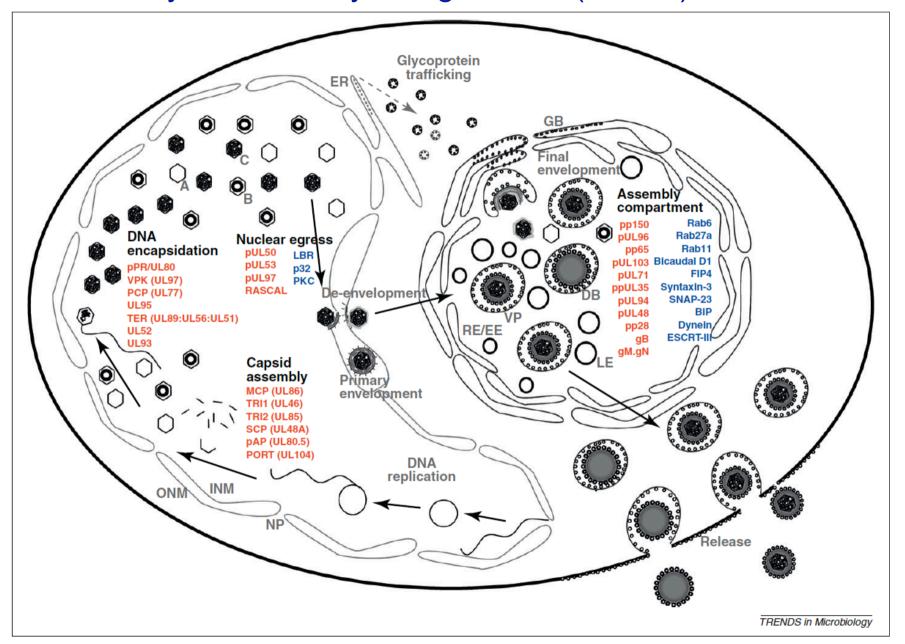


# Assembly of HSV-1 nucleocapsids and the pathway proposed for the virus exit from an infected cell

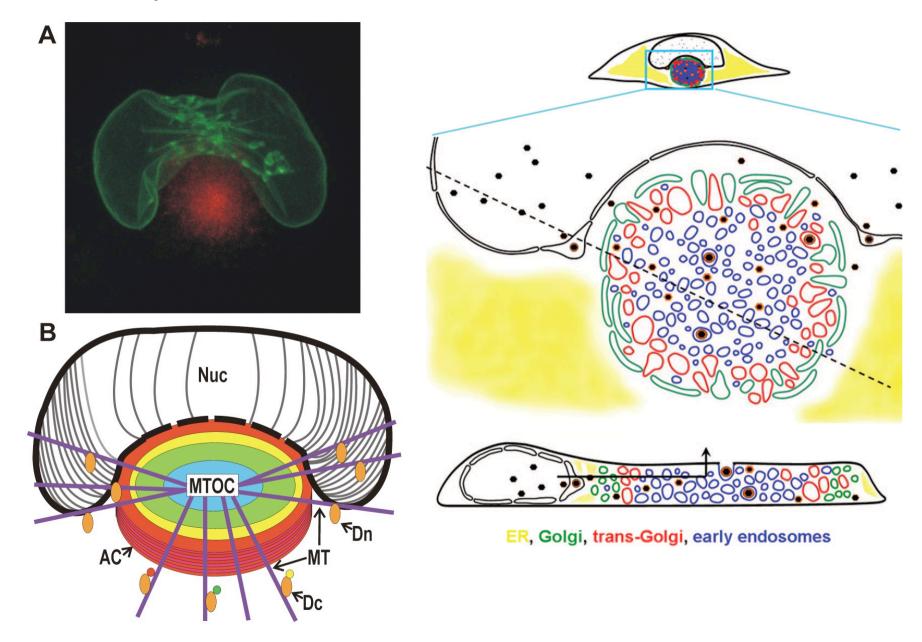




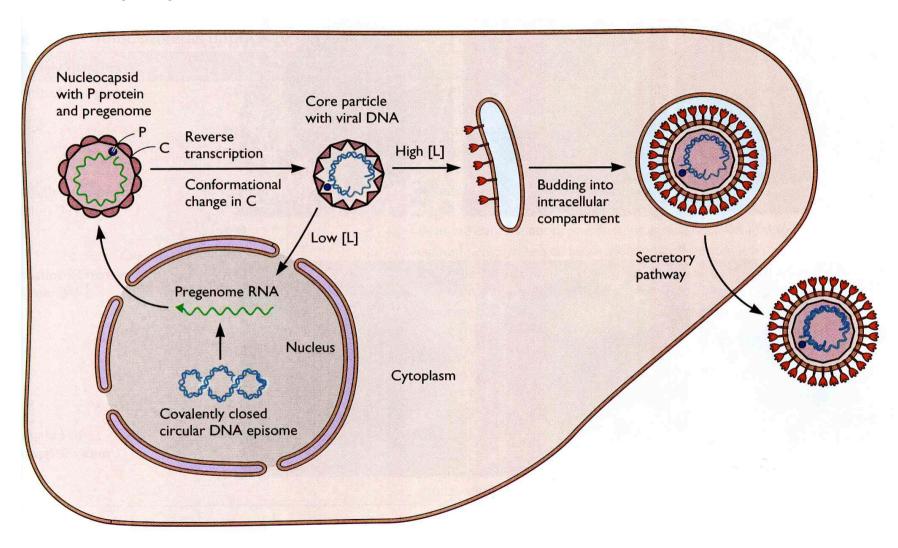
#### Summary of human cytomegalovirus (HCMV) maturation.



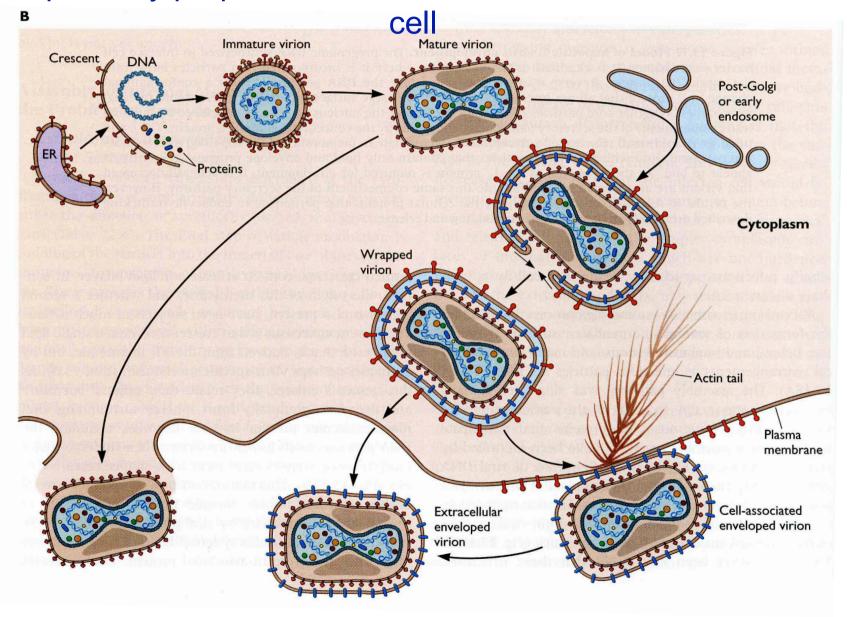
### Microscopic and diagrammatic representations of the assembly compartment and nucleus in an HCMV-infected cell.



### Assembly of HBV nucleocapsids and the pathway proposed for the virus exit from an infected cell



# Assembly of Vaccinia virus nucleocapsids and the pathway proposed for the virus exit from an infected



#### BOX 13.11 BACKGROUND Extracellular and cell-to-cell spread

Many viruses spread from one host cell to another as extracellular virions released from an infected cell (A). Such extracellular dissemination is necessary to infect another naive host. Some viruses, notably alphaherpesviruses and some retroviruses, can also spread from cell to cell without passage through the extracellular environment (B) and can therefore spread by both mechanisms (C).

