STEM CELLS

Definition? Where from? What's for? Where are we?

Definition?

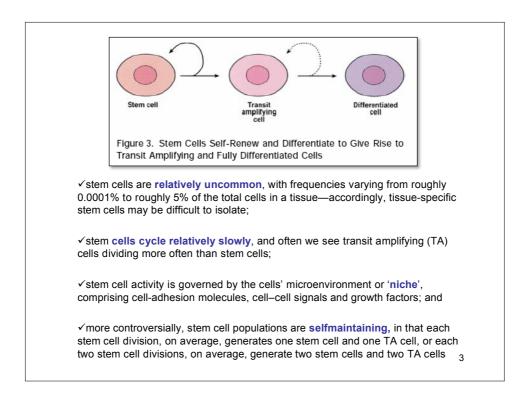
It is now well accepted that a stem cell must fulfill three criteria:

- 1. First, it must be capable of self-renewal, i.e., undergoing symmetric or asymmetric divisions through which the stem cell population is maintained.
- 2. A single cell must be capable of multilineage differentiation.
- 3. In vivo functional reconstitution of a given tissue.

The definition of 'stem cell' is essentially **functional**: "rather than referring to a discrete cellular entity, a stem cell most accurately refers to a biological function that can be induced in many distinct types of cells, even differentiated cells"

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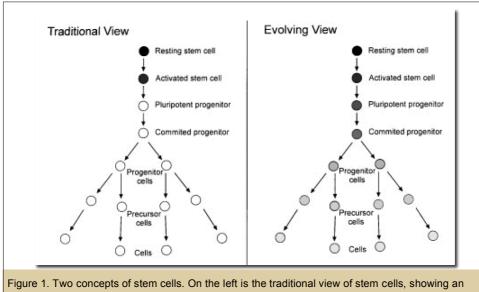
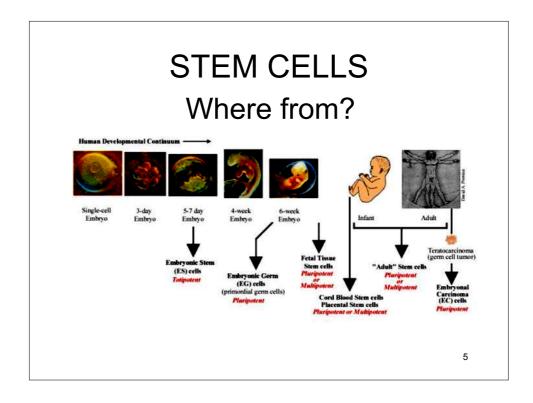
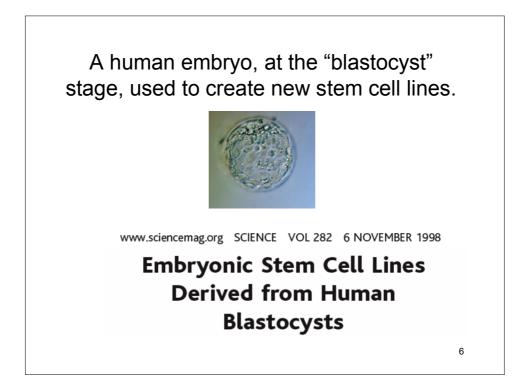
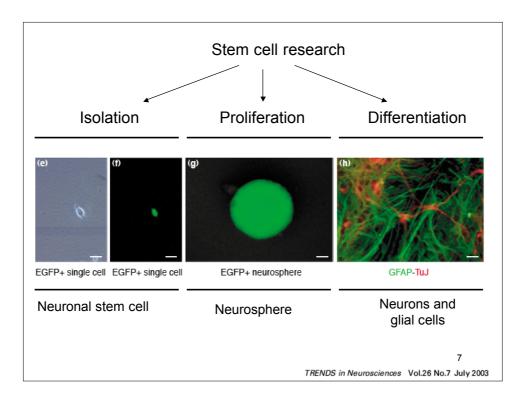


Figure 1. Two concepts of stem cells. On the feft is the traditional view of stem cells, showing an irreversible loss of potency in maturing stem cells. On the right is an evolving view postulated by Blau et al. whereby stemness is a biological function that progressively degenerates over time but remains potentially recruitable within even differentiated cells in particular contexts. Redrawn with substantial modification from Fig. 7 in Blau HM, Brazelton TR, Weimann JM. 2001. The evolving concept of a stem cell: Entity or function? Cell 105:829–841, with permission of Elsevier.







Neural Stem Cells - The ins and outs of Neurospheres

Today, the neurosphere assay is the most common way of isolating and expanding neural stem cells in vitro. So what are "neurospheres"? Neural stem cells can be isolated by taking brain tissue from, for example, embryonic day 14.5 cortex, followed by mechanical dissociation and then plating on culture dishes containing serum-free defined media with EGF. After about 6 days free floating colonies of nestin positive cells can be seen, with each colony containing approximately 200-500 cells (Figure 1). They can be expanded either by cutting the neurospheres into 4-8 pieces or by dissociation into single cells followed by reculturing. If neurospheres are plated on laminin, the EGF withdrawn and



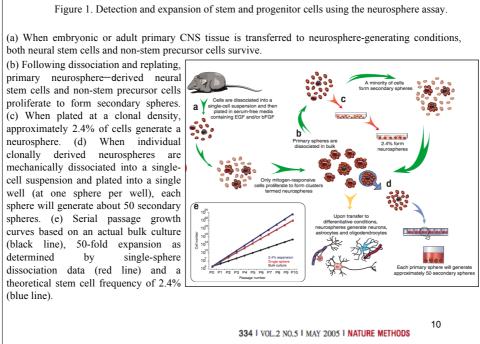
Serum added, they can be seen to differentiate into the three main cell types of the brain, astrocytes, oligodendrocytes and neurons. In particular cells at the periphery of the neurosphere begin to migrate and differentiate while cells at the center remain undifferentiated. By selecting the plating matrix and growth factors the proportion of the three cell types can be controlled (1). For example, NB3 has recently been shown the promote oligodendrocyte formation from mouse brain neurosperes (2).

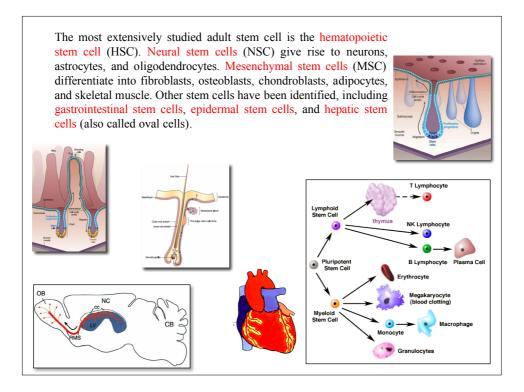
(1) Nature Biotechnology19, (2001) 475-479. (2) J. Biol. Chem: 279, (2004) 25858-25865.⁸

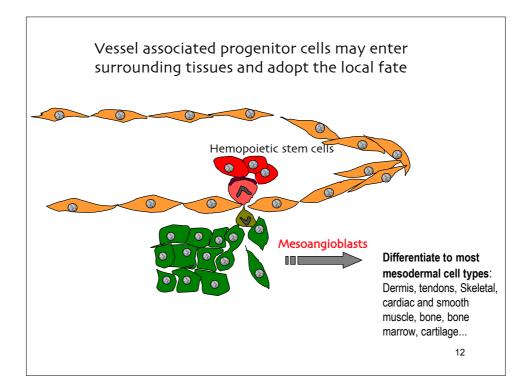
Neurospheres are formed because daughter neural stem cells remain attached to their mothers through many rounds of cell division. So are all the cells in a neurosphere the same? This is an important question that has only been partially answered. Neurospheres are comprised of a heterogenous mix of neural stem cells, neural progenitors, differentiated cells and extracellular matrix proteins (7). Clonal analysis where single cells are isolated and characterised has to be carried out to investigate the exact make-up of neurospheres. Culturing neurospheres from single isolated cells is one way of doing clonal analysis, however, growing cells at very low density may lead to selection of unique populations that are not representative. We will have to identify novel ways of following neural stem cell lineages. Nevertheless, it is clear that the neurosphere assay is a very powerful way to propagate neural stem cells and model neurodevelopment.

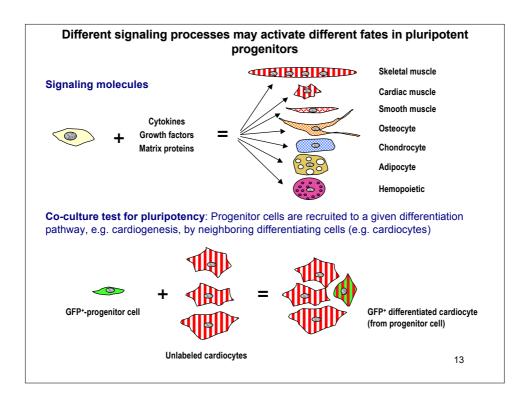
Looking to the future, it will be very important to understand the relationship between the cells of the colony and exactly how many different cell types there are in any one particular neurosphere. The composition of neurospheres isolated from different regions of the brain or under different growth conditions will also be important to establish.

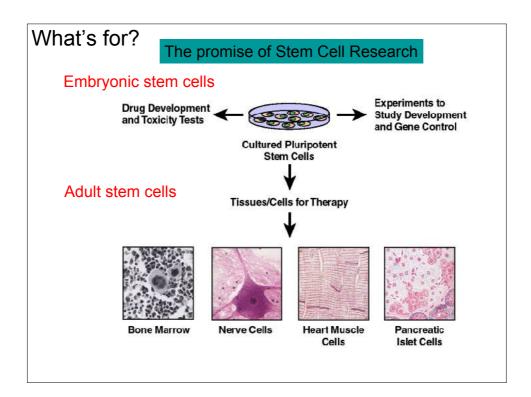
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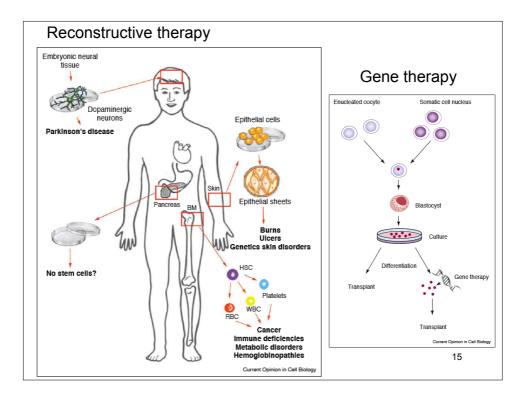


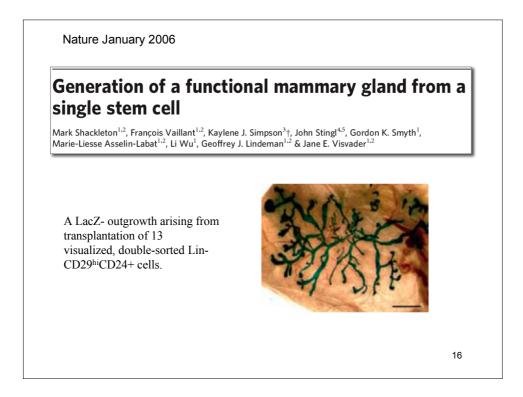


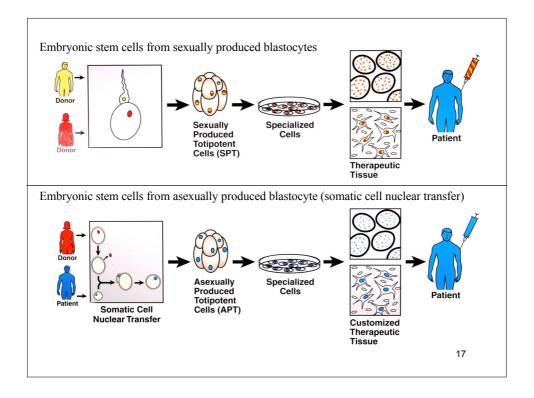


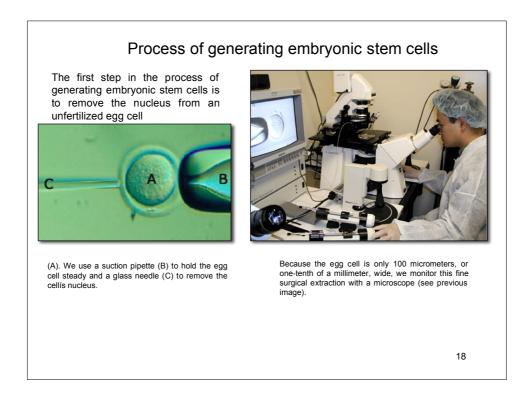


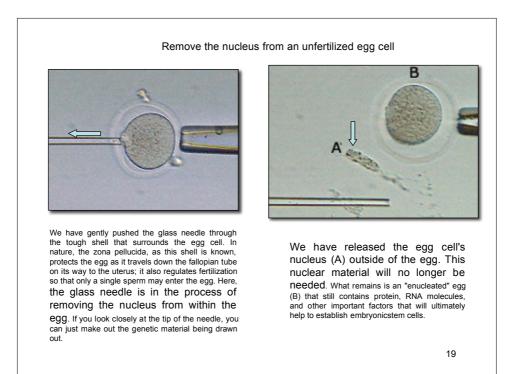


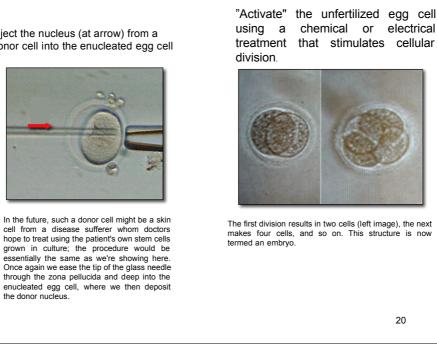






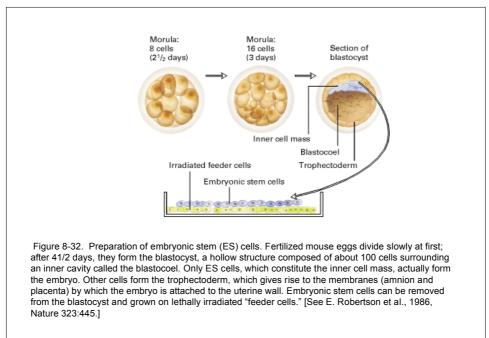




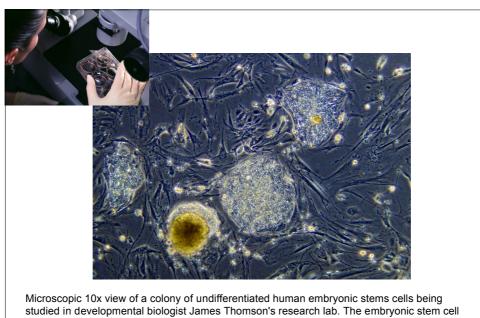


Inject the nucleus (at arrow) from a donor cell into the enucleated egg cell

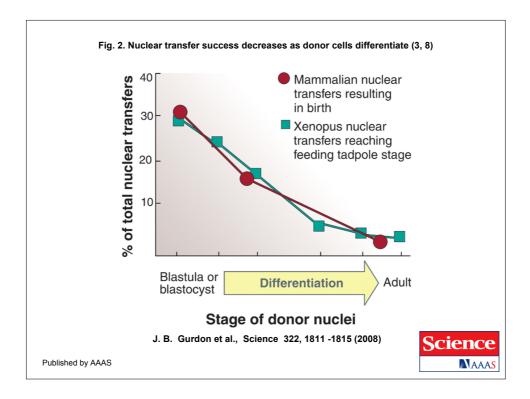
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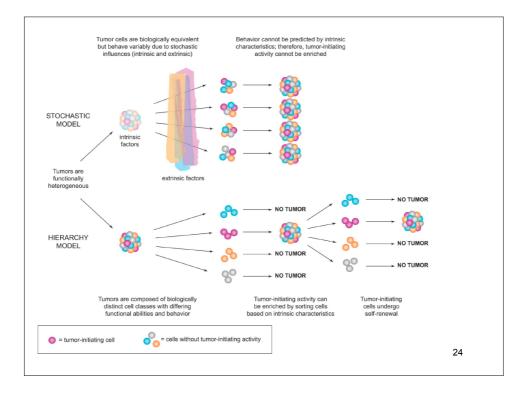


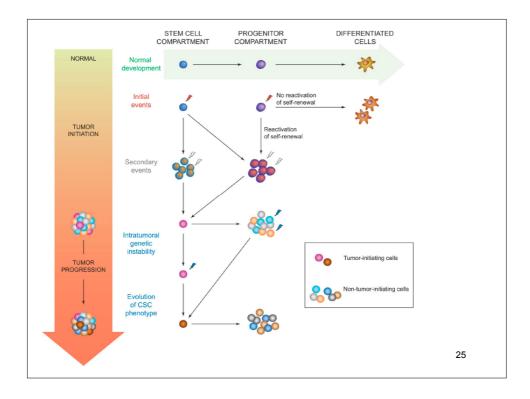


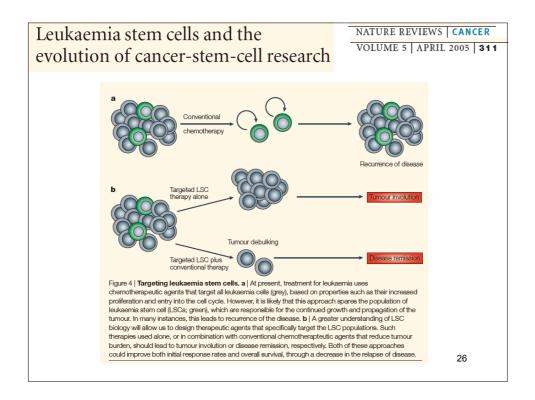


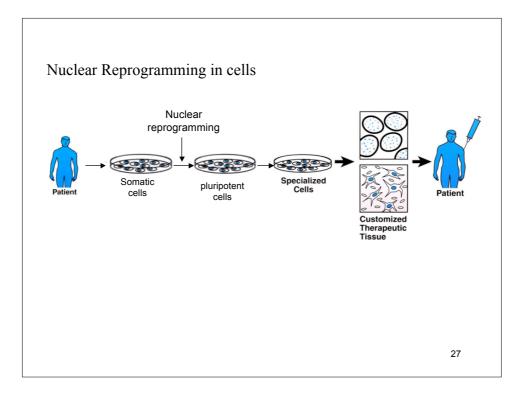
Microscopic 10x view of a colony of undifferentiated human embryonic stems cells being studied in developmental biologist James Thomson's research lab. The embryonic stem cell colonies are the rounded, dense masses of cells. The flat, elongated cells in between the embryonic stem cell colonies are fibroblasts that are used as a "feeder layer" on which the embryonic stem cells are grown. (Source: University of Wisconsin-Madison.)

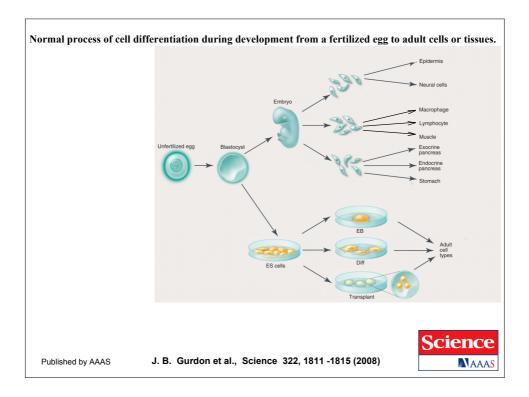


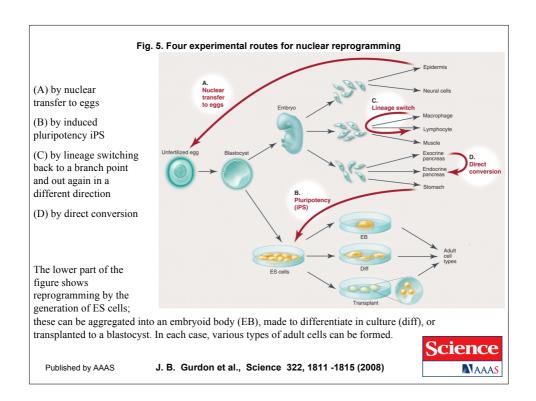


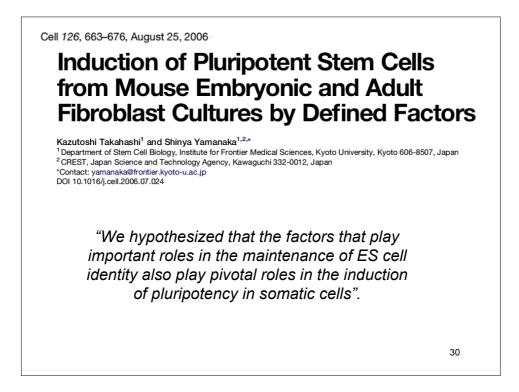






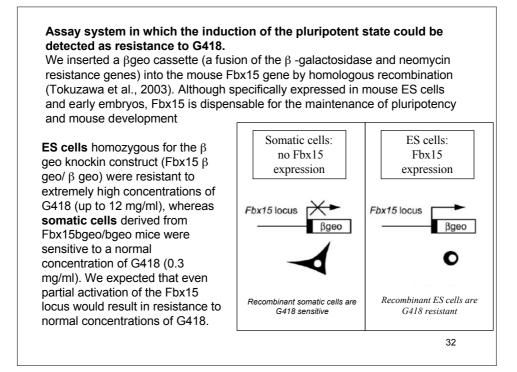


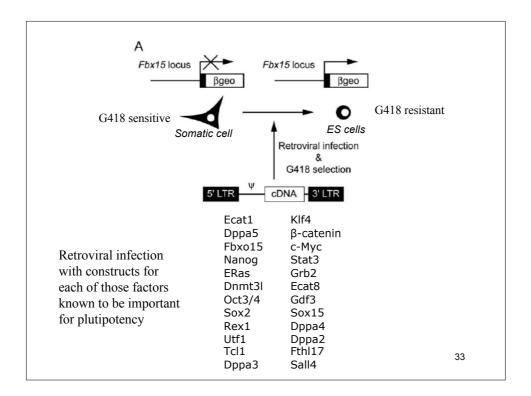


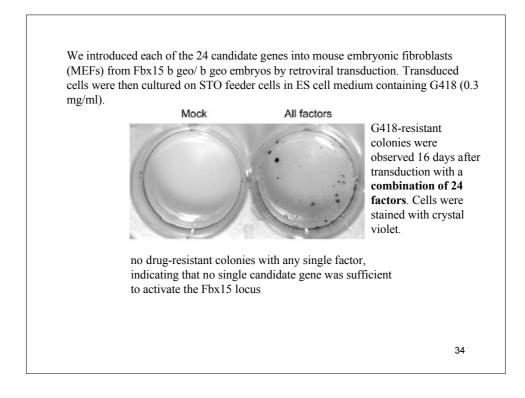


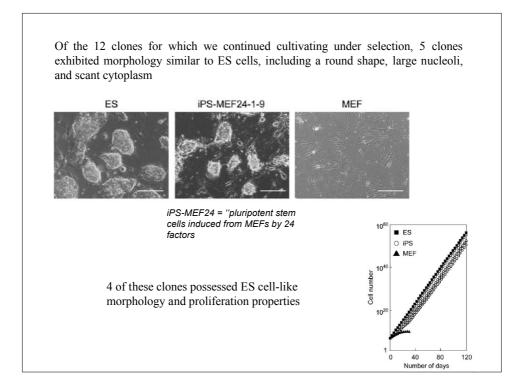
We selected 24 genes as candidates for factors that induce pluripotency in somatic cells, based on our hypothesis that such factors also play pivotal roles in the maintenance of ES cell identity. For b-catenin, c-Myc, and Stat3, we used active forms, S33Y-b-catenin (Sadot et al., 2002), T58A-c-Myc (Chang et al., 2000), and Stat3-C (Bromberg et al., 1999), respectively. Because of the reported negative effect of Grb2 on pluripotency (Burdon et al., 1999; Cheng et al., 1998), we included its dominant-negative mutant Grb2DSH2 (Miyamoto et al., 2004) as 1 of the 24 candidates.

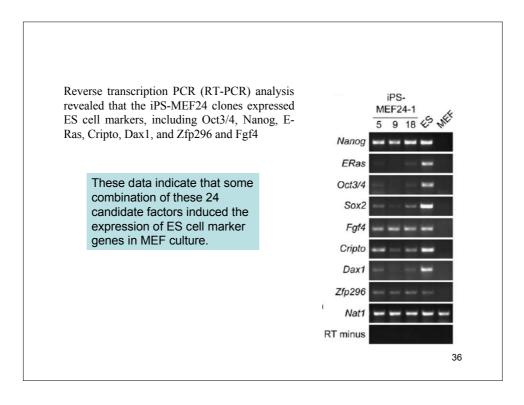
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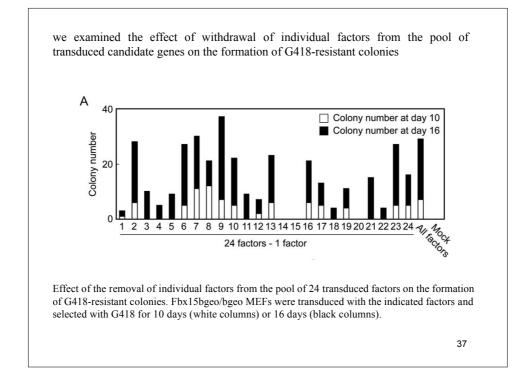


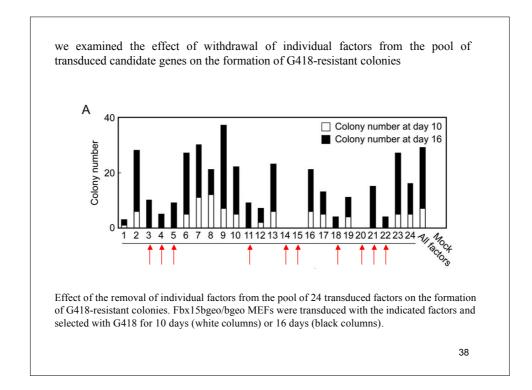


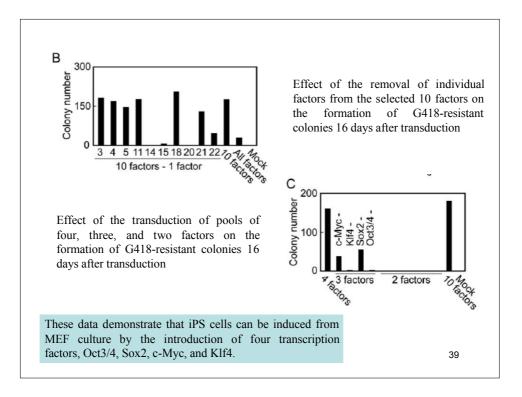


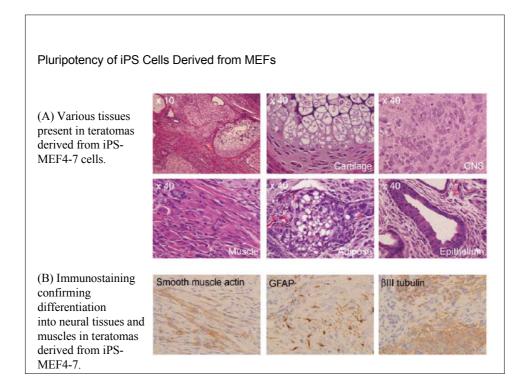


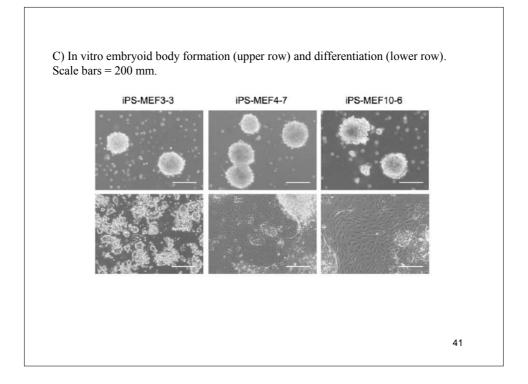


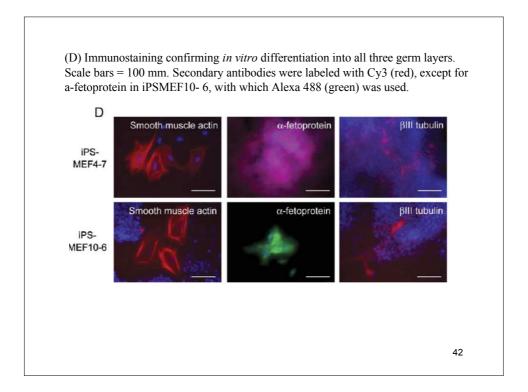








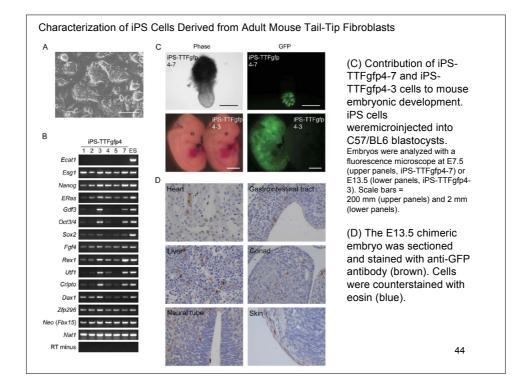




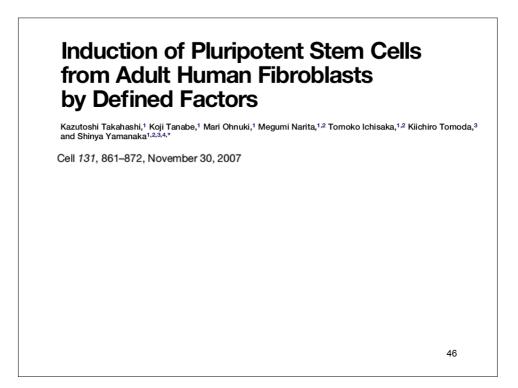
From adult fibroblasts?

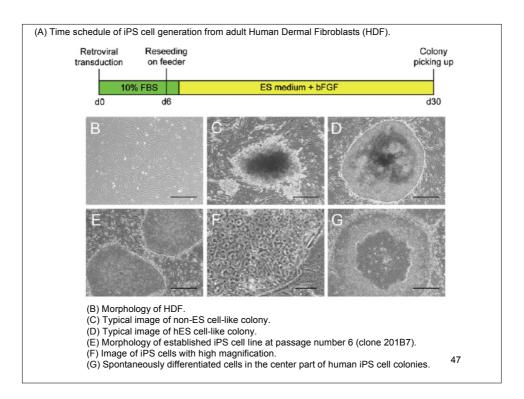
We next introduced the four selected factors into tail-tip fibroblasts (TTFs) of four 7-week-old male Fbx15bgeo/bgeo mice on a C57/BL6-129 hybrid background. We obtained 3 G418-resistant colonies, from each of which we could establish iPS cells (iPS-TTF4). We also introduced the four factors into TTFs from a 12-week-old female Fbx15bgeo/bgeo mouse, which also constitutively expressed green fluorescent protein (GFP) from the CAG promoter and had a C57/BL6-129-ICR hybrid background.

43



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Conclusion: Our study has opened an avenue to generate patient and disease-specific pluripotent stem cells. Even with the presence of retroviral integration, human iPS cells are useful for understanding disease mechanisms, drug screening, and toxicology. For example, hepatocytes derived from iPS cells with various genetic and disease backgrounds can be utilized in predicting liver toxicity of drug candidates. Once the safety issue is overcome, human iPS cells should also be applicable in regenerative medicine. Human iPS cells, however, are not identical to hES cells: DNA microarray analyses detected differences between the two pluripotent stem cell lines. Further studies are essential to determine whether human iPS cells can replace hES in medical applications. 48

