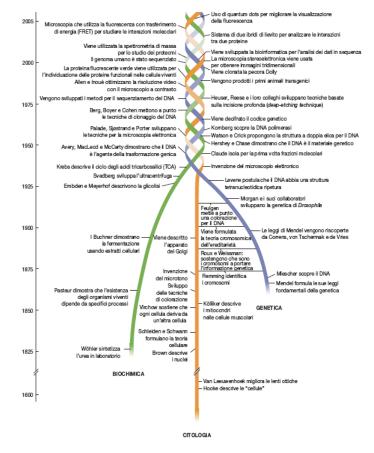
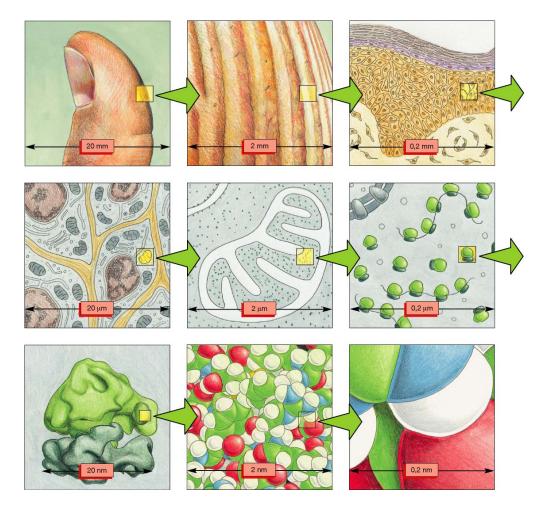
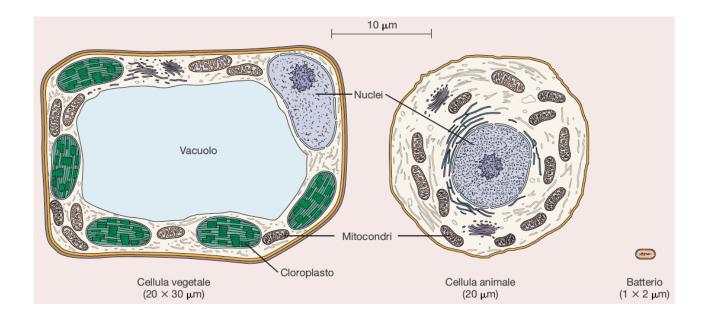
BIOLOGIA CELLULARE







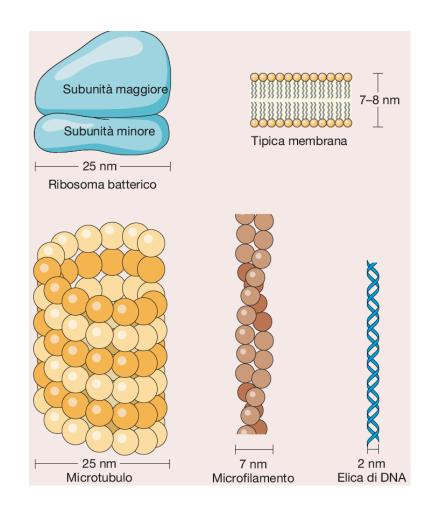


TABELLA 1.1 Diversi tipi di microscopia ottica: un confronto

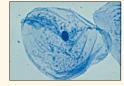
Tipo di microscopia

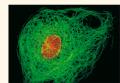
In campo luminoso (campione non colorato): la luce passa direttamente attraverso il campione; a meno che la cellula non sia pigmentata naturalmente o colorata artificialmente, l'immagine presenta uno scarso contrasto.

In campo luminoso (campione colorato): la colorazione con varie sostanze intensifica il contrasto, ma quasi tutte le procedure di colorazione richiedono che le cellule siano fissate (preservate).

A fluorescenza: Mostra la localizzazione di specifiche molecole all'interno della cellula. Le sostanze fluorescenti assorbono le radiazioni ultraviolette ed emettono luce visibile. Le molecole fluorescenti possono essere presenti naturalmente nel campione, ma più spesso si generano marcando le molecole di interesse con coloranti o anticorpi fluorescenti.





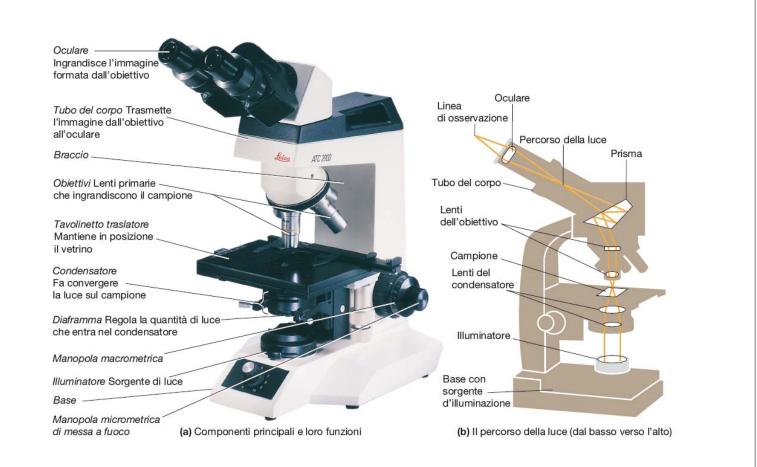




| 50 µm Confocale: utilizza laser e sistemi ottici particolari per focalizzare il fascio di illuminazione su un unico piano all'interno del campione. Si ottengono soltanto le immagini delle regioni all'interno di un ristretto intervallo di profondità rispetto al piano focalizzato. Le regioni al di sopra e al di sotto del piano

selezionato appaiono scure.

Fonte: Campbell e Reece, Biology, 6th ed. (San Francisco: Benjamin Cummings, 2002), pag. 110.



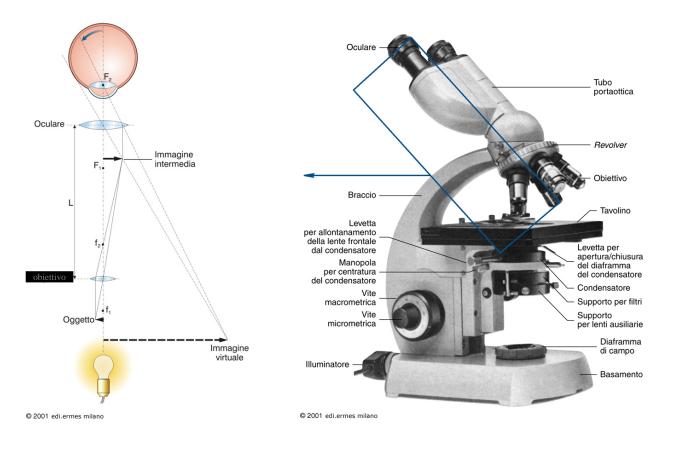
Tipo di microscopia

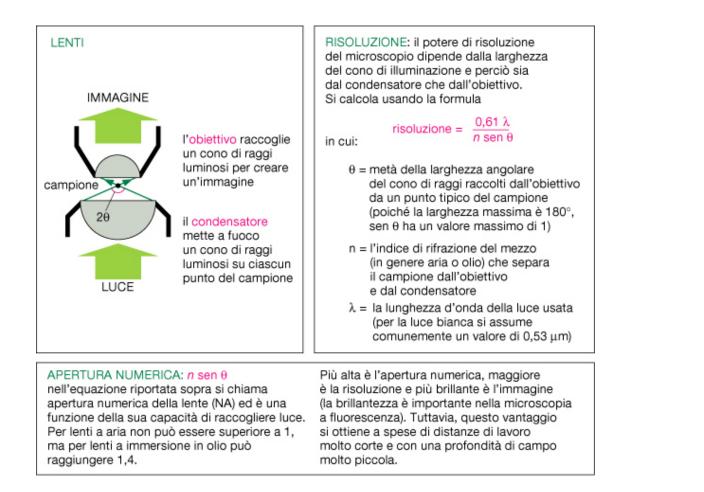
Contrasto di fase: intensifica il contrasto nelle cellule non colorate amplificando le differenze dell'indice di rifrazione all'interno del campione; particolarmente utile per l'osservazione di cellule viventi non pigmentate.

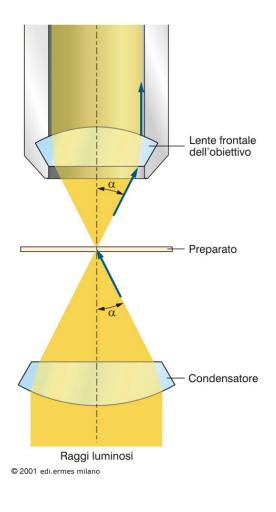
Contrasto interferenziale: utilizza anche modifiche ottiche per esasperare le differenze dell'indice di rifrazione.

Fotografie al microsconia ettico

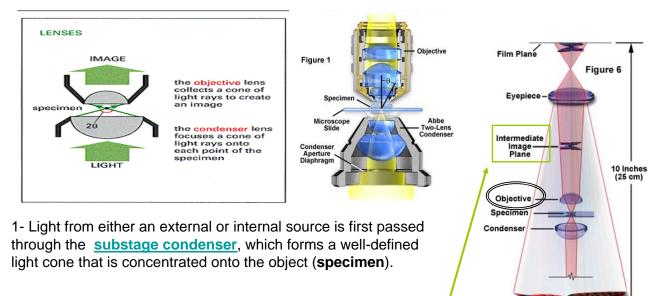
Formazione immagine







Objective Lens



Virtual Image

2- Light passes through the specimen and into the objective which / then projects a <u>real, inverted, and magnified image of the specimen</u> to a fixed plane within the microscope that is termed the **intermediate** image plane (Fig 6).

Objective Numerical Apertures

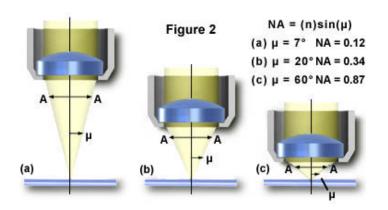
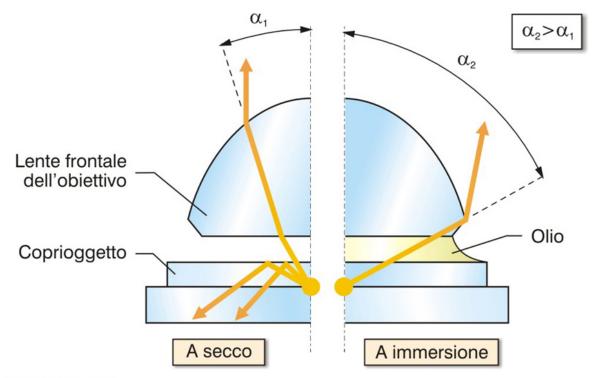


Fig 2 illustrates a series of light cones derived from objectives of varying focal length and numerical aperture. As the light cones change, the angle μ increases from 7° in Fig 2(a) to 60° in Fig 2(c), with a resulting increase in the NA from 0.12 to 0.87, nearing the limit when air is the imaging medium.

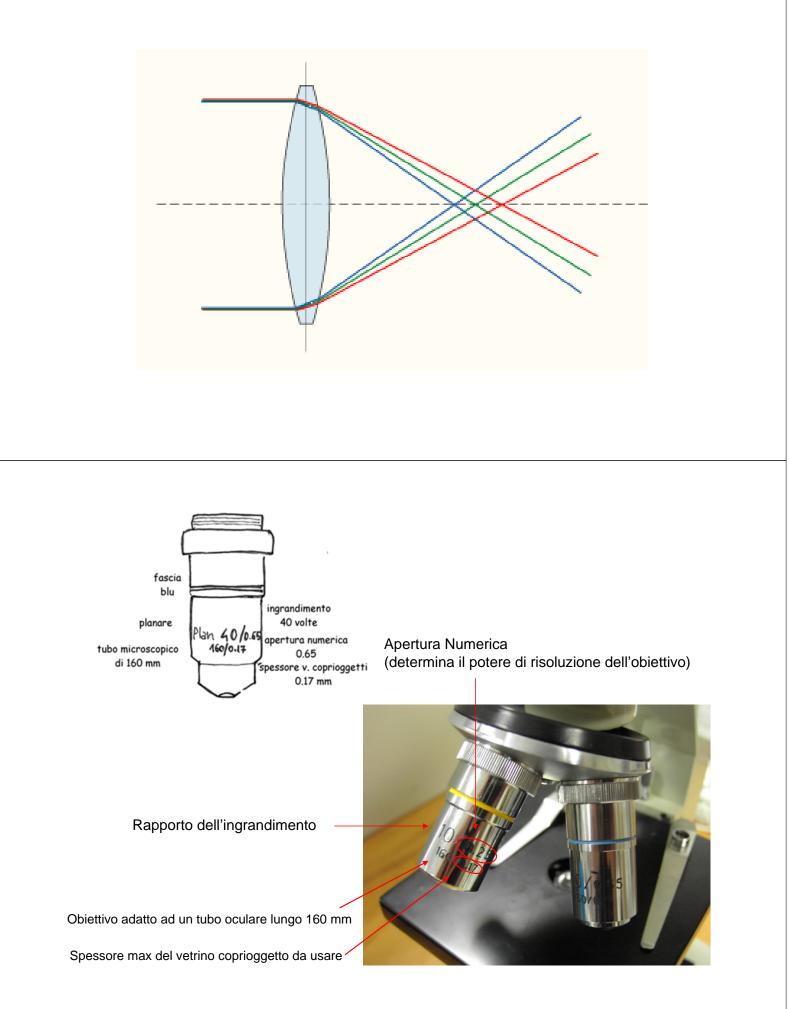
Magnification	Plan Achromat (NA)	Plan Fluorite (NA)	Plan Apochromat (NA)
0.5x	0.025	n/a	n/a
1x	0.04	n/a	n/a
2x	0.06	n/a	0.10
4x	0.10	0.13	0.20
10x	0.25	0.30	0.45
20x	0.40	0.50	0.75
40x	0.65	0.75	0.95
40x (oil)	n/a	1.30	1.00
60x	0.75	0.85	0.95
60x (oil)	n/a	n/a	1.40
100x (oil)	1.25	1.30	1.40
150x	n/a	n/a	0.90

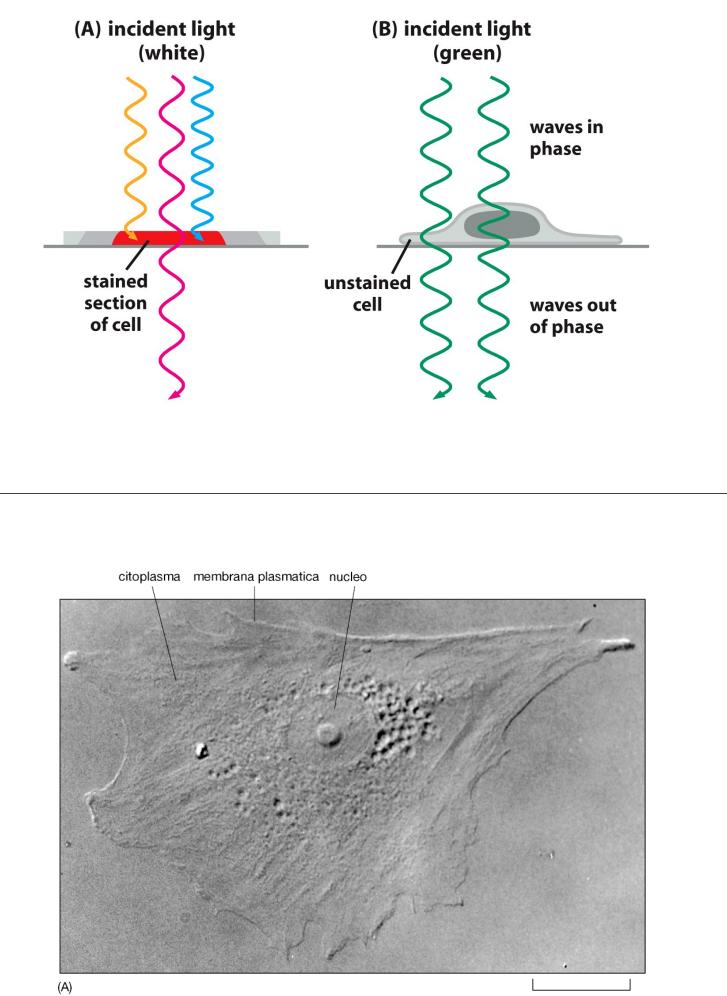
The NA of an objective is also dependent upon the amount of correction for optical aberration



© 2001 edi.ermes milano

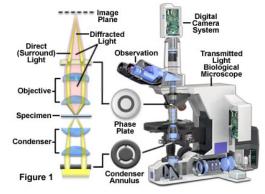
Aberrazione cromatica





Phase contrast

Fritz Zernike developed Phase Contrast in the 1930s and received the Nobel prize in 1953 for the invention.

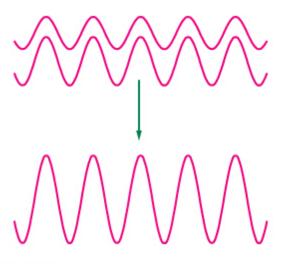


Is a contrast-enhancing optical technique that can be utilized to produce high-contrast images of transparent specimens such as living cells, microorganisms, thin tissue slices, and sub-cellular particles (such as nuclei and other organelles).

One of the major advantages of phase contrast microscopy is that living cells can be examined in their natural state without being killed, fixed, and stained.

As a result, the dynamics of ongoing biological processes in live cells can be observed and recorded in high contrast with sharp clarity of minute specimen detail.

DUE ONDE IN FASE



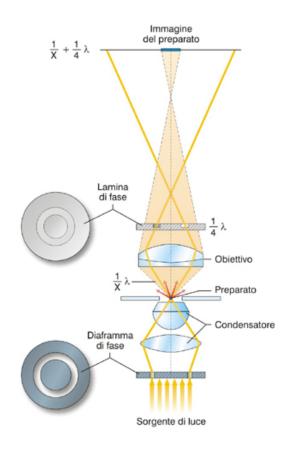
DUE ONDE FUORI FASE

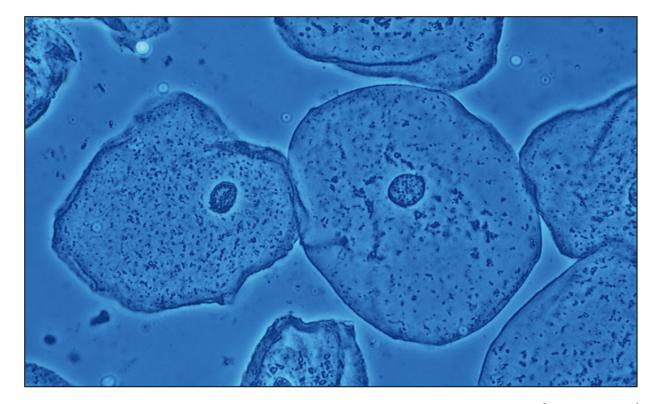
fioca

brillante

Phase Contrast Microscope Configuration

Microscopio a contrasto di fase





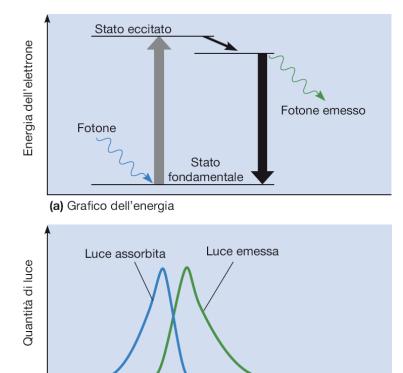


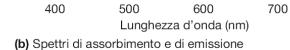
h

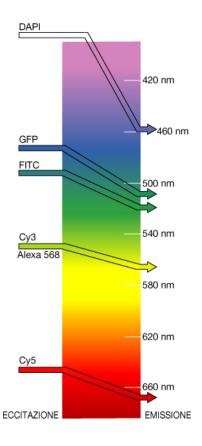
Microscopio rovesciato



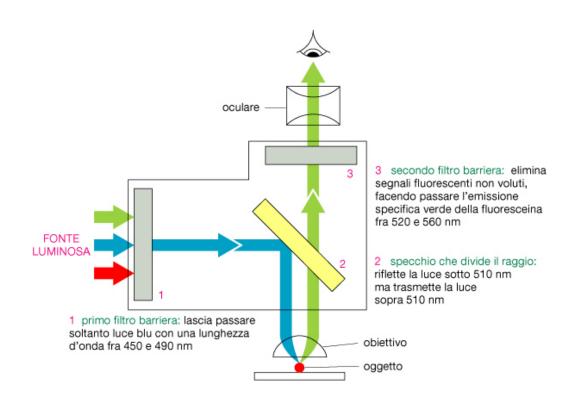
Fluorescenza

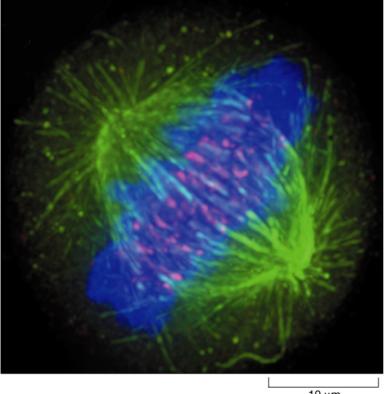




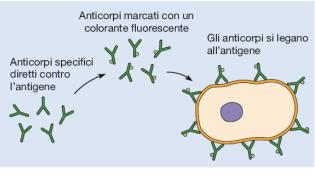


Microscopio a fluorescenza

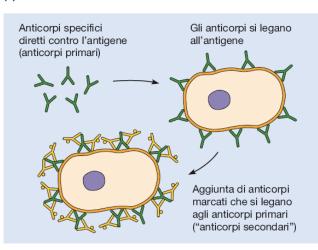




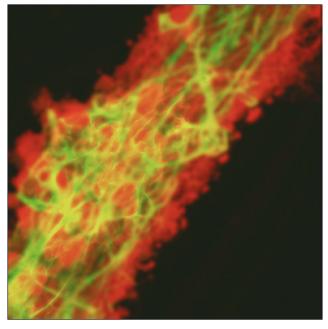
10 µm



(a) Immunofluorescenza

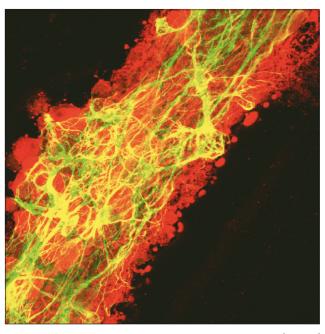


(b) Immunofluorescenza indiretta



(a) Microscopia a fluorescenza tradizionale

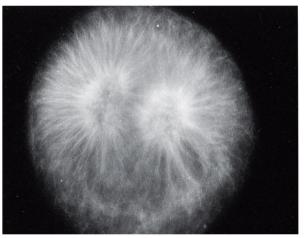
_____ 25 μm



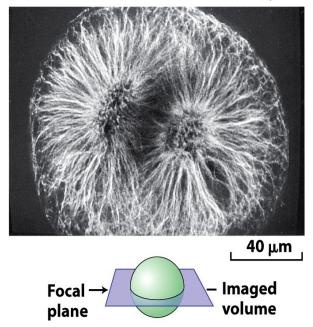
(b) Microscopia a fluorescenza confocale

25 µm

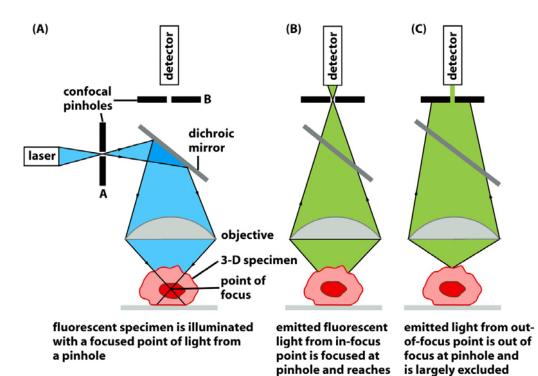
(a) Conventional fluorescence microscopy (b) Confocal fluorescence microscopy



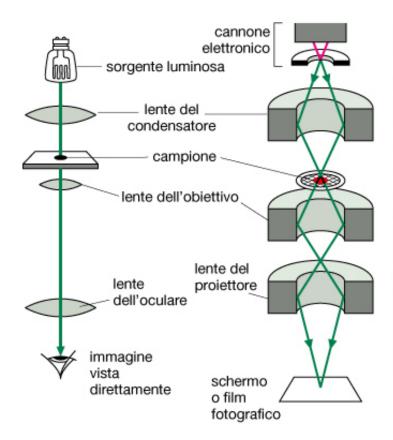
lmaged volume Focal · plane

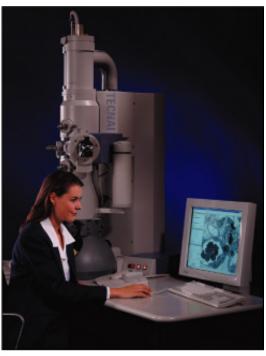


Microscopio confocale

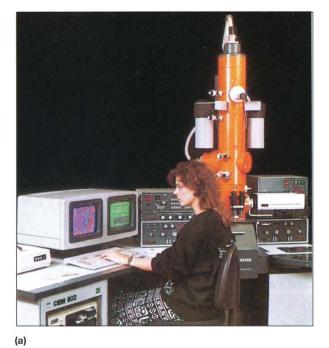


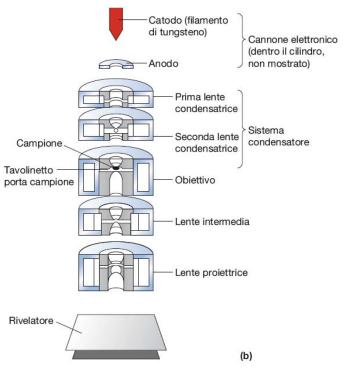
detector





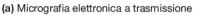
from detector









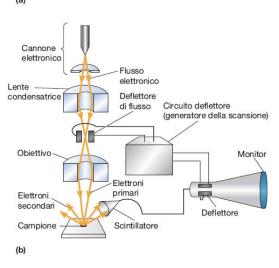




(b) Micrografia elettronica a scansione



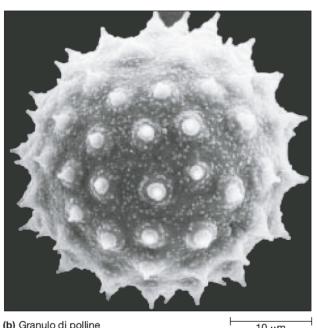






(a) Cellule umane di neuroblastoma

50 µm

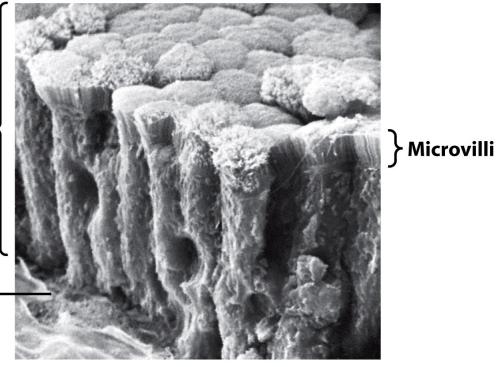


(b) Granulo di polline

10 µm

Absorptive epithelial cells

Basal Iamina



<u>5 μm</u>

Figure 9-24 Molecular Cell Biology, Sixth Edition © 2008 W. H. Freeman and Company